

**NUCLEIC ACIDS AND PROTEINS FROM GROUP B STREPTOCOCCUS**

The present invention relates to proteins derived from *Streptococcus agalactiae*, nucleic acid molecules encoding such proteins, and the use of the proteins as antigens and/or immunogens and in detection/diagnosis. It also relates to a method for the rapid screening of bacterial genomes to isolate and characterise bacterial cell envelope 5 associated or secreted proteins.

The *Group B Streptococcus* (GBS) (*Streptococcus agalactiae*) is an encapsulated bacterium which emerged in the 1970s as a major pathogen of humans causing sepsis 10 and meningitis in neonates as well as adults. The incidence of early onset neonatal infection during the first 5 days of life varies from 0.7 to 3.7 per 1000 live births and causes mortality in about 20% of cases. Between 25-50% of neonates surviving early onset infections frequently suffer neurological sequelae. Late onset neonatal infections occur from 6 days to three months of age at a rate of about 0.5 - 1.0 per 1000 live 15 births.

There is an established association between the colonisation of the maternal genetic tract by GBS at the time of birth and the risk of neonatal sepsis. In humans it has been established that the rectum may act as a reservoir for GBS. Susceptibility in the 20 neonate is correlated with the a low concentration or absence of IgG antibodies to the capsular polysaccharides found on GBS causing human disease. In the USA strains isolated from clinical cases usually belong to capsular serotypes Ia, Ib, II, III although serotype V may be of increasing significance. Type VIII GBS is the major cause of neonatal sepsis in Japan.

25 A possible means of prevention involves intra or postpartum administration of antibiotics to the mother but there are concerns that this might lead to the emergence of resistant organisms and in some cases allergic reactions. Vaccination of the adolescent females to induce long lasting maternally derived immunity is one of the 30 most promising approaches to prevent GBS infections in neonates. The capsular

polysaccharide antigens of these organisms have attracted most attention as with regard to vaccine development. Studies in healthy adult volunteers have shown that serotype Ia, II and III polysaccharides are non-toxic and immunogenic in approximately 65%, 95% and 70% of non-immune adults respectively. One of the 5 problems with using capsule antigens as vaccines is that the response rates vary according to pre-immunisation status and the polysaccharide antigen and not all vaccinees produce adequate levels of IgG antibody as indicated in vaccination studies with GBS polysaccharides in human volunteers.

10 Some people do not respond despite repeated stimuli. These properties are due to the T-independent nature of polysaccharide antigens. One strategy to enhance the immunogenicity of these vaccines is to enhance the T cell dependent properties of polysaccharides by conjugating them to a protein. The use of polysaccharide conjugates looks promising but there are still unresolved questions concerning the 15 nature of the carrier protein. A conjugate vaccine against GBS would require at least 4 different conjugates to be prepared adding to the cost of a vaccine.

Recent evidence also suggests that bacterial surface proteins may be useful to confer 20 immunity. A protein called Rib which is found on most serotype III strains but rarely on serotypes Ia, Ib or II confers immunity to challenge with Rib expressing GBS in animal models (Stalhammar-Carlemalm *et al.*, *Journal of Experimental Medicine* 177:1593-1603 (1993)). Another surface protein of interest as a component of a 25 vaccine is the alpha antigen of the C proteins which protected vaccinated mice against lethal infection with strains expressing alpha protein. The amount of antigen expressed by GBS strains varies markedly.

Approaches to vaccination against GBS infections which rely on the use of capsular 30 polysaccharides have the disadvantage that response rates are likely to vary considerably according to pre-immunisation status and the particular type of polysaccharide antigen used. Results of trials in human volunteers have indicated that

response rates may only be around 65% for some of the key capsule antigens (Larsson *et al.*, *Infection and Immunity* 64:3518-3523 (1996)). It is also not clear whether all individuals responding to the vaccine would have adequate levels of polysaccharide specific IgG which can cross the placenta and afford immunity to neonates. By 5 conjugating a protein carrier to the polysaccharide antigen it may be possible to convert them to T-cell dependent antigens and enhance their immunogenicity.

Preliminary studies with GBS type III polysaccharide-tetanus toxoid conjugate have been encouraging (Baker *et al.*, *Reviews of Infectious Diseases* 7:458-467 (1985), 10 Baker *et al.*, *The New England Journal of Medicine* 319:1180-1185 (1988), Paoletti *et al.*, *Infection and Immunity* 64:677-679 (1996), Paoletti *et al.*, *Infection and Immunity* 62:3236-3243 (1994)) but in developed countries the use of tetanus may be disadvantageous since most adults will have been immunised against tetanus within the past five years. Additional boosters with tetanus toxoid may cause adverse 15 reactions (Boyer., *Current Opinions in Pediatrics* 7:13-18 (1995)). The polysaccharide conjugate vaccines have the disadvantage of being costly to produce and manufacture in comparison with many other kinds of vaccines. There is also the possible risk of problems caused by the cross reactivity between GBS polysaccharides and sialic acid-containing human glycoproteins.

20 An alternative to polysaccharides as antigens is the use of protein antigens derived from GBS. Recent evidence suggest that the GBS surface associated proteins Rib and alpha C protein may be used to confer immunity to GBS infections in experimental model systems (Stalhammar-Carlemalm *et al.*, (1993) [*supra*], Larsson *et al.*, (1996) [*supra*]). However these two proteins are not conserved in all serotypes of GBS which cause disease in humans. Assuming that these antigens would be immunogenic and elicit protective level responses in humans they would not confer protection against all infections as 10% of infectious *Group B streptococci* do not express Rib or C protein alpha.

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This invention seeks to overcome the problem of vaccination against GBS by using a novel screening method specifically designed to identify those *Group B Streptococcus* genes encoding bacterial cell surface associated or secreted proteins (antigens). The proteins expressed by these genes may be immunogenic, and therefore may be useful in the prevention and treatment of *Group B Streptococcus* infection. For the purposes of this application, the term immunogenic means that these proteins will elicit a protective immune response within a subject. Using this novel screening method a number of genes encoding novel *Group B Streptococcus* proteins have been identified.

10 Thus in a first aspect, the present invention provides a *Group B Streptococcus* protein, having a sequence selected from those shown in figure 1, or fragments or derivatives thereof.

15 It will be apparent to the skilled person that proteins and polypeptides included within this group may be cell surface receptors, adhesion molecules, transport proteins, membrane structural proteins, and/or signalling molecules

20 Alterations in the amino acid sequence of a protein can occur which do not affect the function of a protein. These include amino acid deletions, insertions and substitutions and can result from alternative splicing and/or the presence of multiple translation start sites and stop sites. Polymorphisms may arise as a result of the infidelity of the translation process. Thus changes in amino acid sequence may be tolerated which do not affect the protein's function.

25 Thus, the present invention includes derivatives or variants of the proteins, polypeptides, and peptides of the present invention which show at least 50% identity to the proteins, polypeptides and peptides described herein. Preferably the degree of sequence identity is at least 60% and preferably it is above 75%. More preferably still is it above 80%, 90% or even 95%.

The term identity can be used to describe the similarity between two polypeptide sequences. A software package well known in the art for carrying out this procedure is the CLUSTAL program. It compares the amino acid sequences of two polypeptides and finds the optimal alignment by inserting spaces in either sequence as appropriate.

5 The amino acid identity or similarity (identity plus conservation of amino acid type) for an optimal alignment can also be calculated using a software package such as BLASTx. This program aligns the largest stretch of similar sequence and assigns a value to the fit. For any one pattern comparison several regions of similarity may be found, each having a different score. One skilled in the art will appreciate that two 10 polypeptides of different lengths may be compared over the entire length of the longer fragment. Alternatively small regions may be compared. Normally sequences of the same length are compared for a useful comparison to be made.

15 Manipulation of the DNA encoding the protein is a particularly powerful technique for both modifying proteins and for generating large quantities of protein for purification purposes. This may involve the use of PCR techniques to amplify a desired nucleic acid sequence. Thus the sequence data provided herein can be used to design primers for use in PCR so that a desired sequence can be targeted and then amplified to a high degree.

20 Typically primers will be at least five nucleotides long and will generally be at least ten nucleotides long (e.g. fifteen to twenty-five nucleotides long). In some cases primers of at least thirty or at least thirty-five nucleotides in length may be used.

25 As a further alternative chemical synthesis may be used. This may be automated. Relatively short sequences may be chemically synthesised and ligated together to provide a longer sequence.

Thus in a further aspect, the present invention provides , a nucleic acid molecule comprising or consisting of a sequence which is:

- (i) any of the DNA sequences set out in figure 1 herein or their RNA equivalents;
- (ii) a sequence which is complementary to any of the sequences of (i);
- (iii) a sequence which codes for the same protein or polypeptide, as those sequences of (i) or (ii);
- 5 (iv) a sequence which shows substantial identity with any of those of (i), (ii) and (iii); or
- (v) a sequence which codes for a derivative or fragment of a nucleic acid molecule shown in figure 1.

10 The term identity can also be used to describe the similarity between two individual DNA sequences. The 'bestfit' program (Smith and Waterman, *Advances in applied Mathematics*, 482-489 (1981)) is one example of a type of computer software used to find the best segment of similarity between two nucleic acid sequences, whilst the GAP program enables sequences to be aligned along their whole length and finds the 15 optimal alignment by inserting spaces in either sequence as appropriate.

20 The term 'RNA equivalent' when used above indicates that a given RNA molecule has a sequence which is complementary to that of a given DNA molecule, allowing for the fact that in RNA 'U' replaces 'T' in the genetic code. The nucleic acid molecule may be in isolated or recombinant form.

25 The nucleic acid molecule may be in an isolated or recombinant form. DNA constructs can readily be generated using methods well known in the art. These techniques are disclosed, for example in J. Sambrook *et al*, *Molecular Cloning 2<sup>nd</sup> Edition*, Cold Spring Harbour Laboratory Press (1989). Modifications of DNA constructs and the proteins expressed such as the addition of promoters, enhancers, signal sequences, leader sequences, translation start and stop signals and DNA stability controlling regions, or the addition of fusion partners may then be facilitated.

Normally the DNA construct will be inserted into a vector which may be of phage or plasmid origin. Expression of the protein is achieved by the transformation or transfection of the vector into a host cell which may be of eukaryotic or prokaryotic origin. Such vectors and suitable host cells form yet further aspects of the present

5 invention.

The *Group B Streptococcus* proteins (antigens) described herein can additionally be used to raise antibodies, or to generate affibodies. These can be used to detect *Group B Streptococcus*.

10 Thus in a further aspect the present invention provides, an antibody, affibody, or a derivative thereof which binds to any one or more of the proteins, polypeptides, peptides, fragments or derivatives thereof, as described herein.

15 Antibodies within the scope of the present invention may be monoclonal or polyclonal. Polyclonal antibodies can be raised by stimulating their production in a suitable animal host (e.g. a mouse, rat, guinea pig, rabbit, sheep, goat or monkey) when a protein as described herein, or a homologue, derivative or fragment thereof, is injected into the animal. If desired, an adjuvant may be administered together with the protein. Well-known adjuvants include Freund's adjuvant (complete and incomplete) and aluminium hydroxide. The antibodies can then be purified by virtue of their binding to a protein as

20 described herein.

Monoclonal antibodies can be produced from hybridomas. These can be formed by fusing myeloma cells and spleen cells which produce the desired antibody in order to form an immortal cell line. Thus the well-known Kohler & Milstein technique (*Nature* 256 (1975)) or subsequent variations upon this technique can be used.

30 Techniques for producing monoclonal and polyclonal antibodies that bind to a particular polypeptide/protein are now well developed in the art. They are discussed in standard

immunology textbooks, for example in Roitt *et al*, *Immunology* second edition (1989), Churchill Livingstone, London.

In addition to whole antibodies, the present invention includes derivatives thereof which  
5 are capable of binding to proteins etc as described herein. Thus the present invention includes antibody fragments and synthetic constructs. Examples of antibody fragments and synthetic constructs are given by Dougall *et al* in *Tibtech* 12 372-379 (September 1994).

10 Antibody fragments include, for example, Fab, F(ab')<sub>2</sub> and Fv fragments. Fab fragments (These are discussed in Roitt *et al* [*supra*]). Fv fragments can be modified to produce a synthetic construct known as a single chain Fv (scFv) molecule. This includes a peptide linker covalently joining V<sub>h</sub> and V<sub>l</sub> regions, which contributes to the stability of the molecule. Other synthetic constructs that can be used include CDR peptides. These are  
15 synthetic peptides comprising antigen-binding determinants. Peptide mimetics may also be used. These molecules are usually conformationally restricted organic rings that mimic the structure of a CDR loop and that include antigen-interactive side chains.

20 Synthetic constructs include chimaeric molecules. Thus, for example, humanised (or primatised) antibodies or derivatives thereof are within the scope of the present invention. An example of a humanised antibody is an antibody having human framework regions, but rodent hypervariable regions. Ways of producing chimaeric antibodies are discussed for example by Morrison *et al* in *PNAS*, 81, 6851-6855 (1984) and by Takeda *et al* in *Nature*, 314, 452-454 (1985).

25 Synthetic constructs also include molecules comprising an additional moiety that provides the molecule with some desirable property in addition to antigen binding. For example the moiety may be a label (e.g. a fluorescent or radioactive label). Alternatively, it may be a pharmaceutically active agent.

Affibodies are proteins which are found to bind to target proteins with a low dissociation constant. They are selected from phage display libraries expressing a segment of the target protein of interest (Nord K, Gunneriusson E, Ringdahl J, Stahl S, Uhlen M, Nygren PA, Department of Biochemistry and Biotechnology, Royal Institute of Technology 5 (KTH), Stockholm, Sweden).

In a further aspect the invention provides an immunogenic composition comprising one or more proteins, polypeptides, peptides, fragments or derivatives thereof, or nucleotide sequences described herein. A composition of this sort may be useful in the 10 treatment or prevention of *Group B Streptococcus* infection in subject. In a preferred aspect of the invention the immunogenic composition is a vaccine.

In other aspects the invention provides:

- 15      i)     Use of an immunogenic composition as described herein in the preparation of a medicament for the treatment or prophylaxis of *Group B Streptococcus* infection. Preferably the medicament is a vaccine.
- 20      ii)    A method of detection of *Group B Streptococcus* which comprises the step of bringing into contact a sample to be tested with at least one antibody, affibody, or a derivative thereof, as described herein.
- 25      iii)   A method of detection of *Group B Streptococcus* which comprises the step of bringing into contact a sample to be tested with at least one protein, polypeptide, peptide, fragments or derivatives as described herein.
- 30      iv)    A method of detection of *Group B Streptococcus* which comprises the step of bringing into contact a sample to be tested with at least one nucleic acid molecule as described herein.

v) A kit for the detection of *Group B Streptococcus* comprising at least one antibody, affibody, or derivatives thereof, described herein.

5 vi) A kit for the detection of *Group B Streptococcus* comprising at least one *Group B Streptococcus* protein, polypeptide, peptide, fragment or derivative thereof, as described herein.

10 vii) A kit for the detection of *Group B Streptococcus* comprising at least one nucleic acid of the invention.

15 As described previously, the novel proteins described herein are identified and isolated using a novel screening method which specifically identifies those *Group B Streptococcus* genes encoding bacterial cell envelope associated or secreted proteins.

20 15 The information necessary for the secretion/export of proteins has been extensively studied in bacteria. In the majority of cases, export requires a signal peptide positioned at the N-terminus of the precursor protein to target the precursor to translocation sites on the membrane. During or after translocation, the signal peptide is removed by a signal peptidase. The ultimate destination/localisation of the protein, (whether it be secreted extracellularly or anchored to the bacterium's surface, etc) is determined by sequences other than the leader peptide sequence.

25 Recently, Poquet *et al.* (*J. Bacteriol.* **180**:1904-1912 (1998)) have described a screening vector incorporating the *nuc* gene lacking its own signal leader as a reporter to identify exported proteins in Gram positive bacteria, and have applied it to *L. lactis*. Staphylococcal nuclease is a naturally secreted heat-stable, monomeric enzyme which has been efficiently expressed and secreted in a range of Gram positive bacteria (Shortle., *Gene* **22**:181-189 (1983), Kovacevic *et al.*, *J. Bacteriol.* **162**:521-528 (1985), Miller *et al.*, *J. Bacteriol.* **169**:3508-3514 (1987), Liebl *et al.*, *J. Bacteriol.*

174:1854-1861(1992), Le Loir *et al.*, *J. Bacteriol.* 176:5135-5139 (1994), Poquet *et al.*, 1998 [*supra*]). The screening vector (pFUN) contains the pAM $\beta$ 1 replicon which functions in a broad host range of Gram-positive bacteria in addition to the ColE1 replicon that promotes replication in *Escherichia coli* and certain other Gram negative bacteria. Unique cloning sites present in the vector can be used to generate transcriptional and translational fusions between cloned genomic DNA fragments and the open reading frame of the truncated *nuc* gene devoid of its own signal secretion leader. The *nuc* gene makes an ideal reporter gene because the secretion of nuclease can readily be detected using a simple and sensitive plate test: Recombinant colonies 5 secreting the nuclease develop a pink halo whereas control colonies remain white 10 (Shortle, 1983 [*supra*], Le Loir *et al.*, 1994 [*supra*]).

A direct screen to identify and isolate DNA encoding bacterial cell envelope 15 associated or secreted proteins (antigens) in pathogenic bacteria has been developed by the present inventors which utilises a vector-system (pTREP1 expression vector) in *Lactococcus lactis* that specifically detects DNA sequences which are adjacent to, and associated with DNA encoding surface proteins from *Group B Streptococcus*. The screening vector also incorporates the *nuc* gene encoding the *Staphylococcal* nuclease 20 as a reporter gene.

Only the part of the *nuc* gene encoding the mature nuclease protein (minus its signal 25 peptide sequence) is cloned into the pTREP1 expression vector in *L. lactis*. In this form, the *nuc*-encoded nuclease cannot be secreted even when expressed intracellularly. The reporter vector is then randomly combined with appropriately digested genomic DNA from *Group B Streptococcus*, cloned into *L. lactis* and used as a screening system for sequences permitting the export of nuclease. In this way gene/partial gene sequences encoding exported proteins from *Group B Streptococcus* are isolated. Once a partial gene sequence is obtained, full length sequences encoding 30 exported proteins can readily be obtained using techniques well known in the art.

In possessing a promoter, the pTREP1-*nuc* vectors differ from the pFUN vector described by Poquet *et al.* (1998) [*supra*], which was used to identify *L. lactis* exported proteins by screening directly for *Nuc* activity directly in *L. lactis*. As the 5 pFUN vector does not contain a promoter upstream of the *nuc* open reading frame the cloned genomic DNA fragment must also provide the signals for transcription in addition to those elements required for translation initiation and secretion of *Nuc*. This limitation may prevent the isolation of genes that are distant from a promoter for example genes which are within polycistronic operons. Additionally there can be no 10 guarantee that promoters derived from other species of bacteria will be recognised and functional in *L. lactis*. Certain promoters may be under stringent regulation in the natural host but not in *L. lactis*. In contrast, the presence of the P1 promoter in the pTREP1-*nuc* series of vectors ensures that promoterless DNA fragments (or DNA 15 fragments containing promoter sequences not active in *L. lactis*) may still be transcribed. Thus yet another advantage of this invention is that genes missed in other screening methods may be identified.

Hence in a further aspect the present invention provides a method of screening for 20 DNA encoding bacterial cell wall associated or surface antigens in gram positive bacteria comprising the steps of:

- combining a reporter vector including the nucleotide sequence encoding the mature from of the staphylcoccus nuclease gene and an upstream promoter region with DNA from a gram positive bacteria.
- transforming the resultant vector into *Lactococcus lactis* cells.
- assaying for the secretion of *staphlycoccus* nuclease protein in the 25 transformed cells.

Preferably, the reporter vector is one of the pTREP1-*nuc* vectors shown in figure 4.

In another aspect, the present invention provides a vector as shown in figure 4 for use in screening for DNA encoding exported or surface antigens in gram positive bacteria. Examples of gram positive bacteria which may be screened include *Group B Streptococcus*, *Streptococcus pneumoniae*, *Staphylococcus aureus* or pathogenic 5 *Group A Streptococci*.

Given that the inventors have identified a group of important proteins, such proteins are potential targets for anti-microbial therapy. It is necessary, however, to determine whether each individual protein is essential for the organism's viability.

10 Thus, the present invention also provides a method of determining whether a protein or polypeptide as described herein represents a potential anti-microbial target which comprises inactivating said protein and determining whether *Group B Streptococcus* is still viable.

15 A suitable method for inactivating the protein is to effect selected gene knockouts, ie prevent expression of the protein and determine whether this results in a lethal change. Suitable methods for carrying out such gene knockouts are described in Li *et al*, *P.N.A.S.*, **94**:13251-13256 (1997) and Kolkman *et al*

20 In a final aspect the present invention provides the use of an agent capable of antagonising, inhibiting or otherwise interfering with the function or expression of a protein or polypeptide of the invention in the manufacture of a medicament for use in the treatment or prophylaxis of *Group B Streptococcus* infection.

25 The invention will now be described by means of the following example which should not in any way be construed as limiting. The examples refer to the figures in which

Fig 1: (A) Shows a number of full length nucleotide sequences encoding antigenic *Group B Streptococcus* proteins. (B) Shows the corresponding amino acid sequences.

5 Fig 2: Shows a number of oligonucleotide primers used in the screening process

**nucS1** primer designed to amplify a mature form of the nuc A gene

**nucS2-** primer designed to amplify a mature form of the nuc A gene.

**nucS3** primer designed to amplify a mature form of the nuc A gene

10 **nucR** primer designed to amplify a mature form of the nuc A gene

**nucseq** primer designed to sequence DNA cloned into the pTREP-Nuc vector

**pTREPF** nucleic acid sequence containing recognition site for ECORV. Used for cloning fragments into pTREX7.

15 **pTREPR** nucleic acid sequence containing recognition site for BAMH1. Used for cloning fragments into pTREX7.

**PUCF** forward sequencing primer, enables direct sequencing of cloned DNA fragments.

**VR** example of gene specific primer used to obtain further antigen DNA sequence by the method of DNA walking.

20 **V1** example of gene specific primer used to obtain further antigen DNA sequence by the method of DNA walking.

**V2** example of gene specific primer used to obtain further antigen DNA sequence by the method of DNA walking.

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Fig 3: (i) Schematic presentation of the nucleotide sequence of the unique gene cloning site immediately upstream of the mature *nuc* gene in pTREP1-*nuc1*, pTREP1-*nuc2* and pTREP1-*nuc3*. Each of the pTREP-*nuc* vectors contain an

EcoRV (a SmaI site in pTREP1-*nuc2*) cleavage site which allows cloning of genomic DNA fragments in 3 different frames with respect to the mature *nuc* gene.

(ii) A physical and genetic summary map of the pTREP1-*nuc* vectors. The expression cassette incorporating *nuc*, the macrolides, lincosamides and streptogramin B (MLS) resistance determinant, and the replicon (rep) *Ori*-pAM $\beta$ 1 are depicted (not drawn to scale).

(iii) Schematic presentation of the expression cassette showing the various sequence elements involved in gene expression and location of unique restriction endonuclease sites (not drawn to scale).

Fig 4: Shows the results of various DNA vaccine trials;

Fig 5: Shows the results of a second group of DNA vaccine trials;

Figs 6-11: Show various Southern Blot analyses of different Group B streptococcus strains.

### Example 1

Thus far more than 100 gene/partial gene sequences putatively encoding exported proteins in *S. agalactiae* have been identified using the nuclease screening system of the invention. These have been further analysed to remove artifacts. The nucleotide sequences of genes identified using the screening system has been characterised using a number of parameters described below. All of these sequences are novel in that they have not been described previously.

1. All putative surface proteins are analysed for leader/signal peptide sequences. Bacterial signal peptide sequences share a common design. They are characterised by a short positively charged N-terminus (N region) immediately preceding a stretch of hydrophobic residues (central portion-h region) followed by a

more polar C-terminal portion which contains the cleavage site (c-region). Computer software is used to perform hydropathy profiling of putative proteins (Marcks, *Nuc. Acid. Res.*, 16:1829-1836 (1988)) which is used to identify the distinctive hydrophobic portion (h-region) typical of leader peptide sequences. In addition, the presence/absence of a potential ribosomal binding site (Shine-Dalgarno sequence required for translation) is also noted.

5 2. All putative surface protein sequences are used to search the OWL sequence database which includes a translation of the GENBANK and SWISSPROT database.. This allows identification of similar sequences which may have been previously 10 characterised not only at the sequence level but at a functional level. It may also provide information indicating that these proteins are indeed surface related and not artifacts.

15 3. Putative *S. agalactiae* surface proteins are also be assessed for their novelty. Some of the identified proteins may or may not possess a typical leader peptide sequence and may not show homology with any DNA/protein sequences in the database. Indeed these proteins may indicate the primary advantage of our screening method, i.e. isolating atypical surface-related proteins, which would have been missed 20 in all previously described screening protocols.

20 The construction of three reporter vectors and their use in *L. lactis* to identify and isolate genomic DNA fragments from pathogenic bacteria encoding secreted or surface associated proteins is now described.

25 **Construction of the pTREP1-nuc series of reporter vectors**

(a) **Construction of expression plasmid pTREP1**

30 The pTREP1 plasmid is a high-copy number (40-80 per cell) theta-replicating gram positive plasmid, which is a derivative of the pTREX plasmid which is itself a derivative of the the previously published pIL253 plasmid. pIL253 incorporates the

broad Gram-positive host range replicon of pAM $\beta$ 1 (Simon and Chopin, 1988) and is non-mobilisable by the *L. lactis* sex-factor. pIL253 also lacks the *tra* function which is necessary for transfer or efficient mobilisation by conjugative parent plasmids exemplified by pIL501. The Enterococcal pAM $\beta$ 1 replicon has previously been transferred to various species including *Streptococcus*, *Lactobacillus* and *Bacillus* species as well as *Clostridium acetobutylicum*, (LeBlanc *et al.*, *Proceedings of the National Academy of Science USA* 75:3484-3487 (1978)) indicating the potential broad host range utility. The pTREP1 plasmid represents a constitutive transcription vector.

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The pTREX vector was constructed as follows. An artificial DNA fragment containing a putative RNA stabilising sequence, a translation initiation region (TIR), a multiple cloning site for insertion of the target genes and a transcription terminator was created by annealing 2 complementary oligonucleotides and extending with Tfl DNA polymerase. The sense and anti-sense oligonucleotides contained the recognition sites for NheI and BamHI at their 5' ends respectively to facilitate cloning. This fragment was cloned between the XbaI and BamHI sites in pUC19NT7, a derivative of pUC19 which contains the T7 expression cassette from pLET1 (Wells *et al.*, *J. Appl. Bacteriol.* 74:629-636 (1993)) cloned between the EcoRI and HindIII sites. The resulting construct was designated pUCLEX. The complete expression cassette of pUCLEX was then removed by cutting with HindIII and blunting followed by cutting with EcoRI before cloning into EcoRI and SacI (blunted) sites of pIL253 to generate the vector pTREX (Wells and Schofield, *In Current advances in metabolism, genetics and applications-NATO ASI Series. H* 98:37-62. (1996)). The putative RNA stabilising sequence and TIR are derived from the *Escherichia coli* T7 bacteriophage sequence and modified at one nucleotide position to enhance the complementarity of the Shine Dalgarno (SD) motif to the ribosomal 16s RNA of *Lactococcus lactis* (Schofield *et al.* pers. coms. University of Cambridge Dept. Pathology.).

A *Lactococcus lactis* MG1363 chromosomal DNA fragment exhibiting promoter activity which was subsequently designated P7 was cloned between the EcoRI and BglII sites present in the expression cassette, creating pTREX7. This active promoter region had been previously isolated using the promoter probe vector pSB292 (Waterfield *et al.*, *Gene* 165:9-15 (1995)). The promoter fragment was amplified by PCR using the Vent DNA polymerase according to the manufacturer.

The pTREP1 vector was then constructed as follows. An artificial DNA fragment which included a transcription terminator, the forward pUC sequencing primer, a promoter multiple cloning site region and a universal translation stop sequence was created by annealing two overlapping partially complementary synthetic oligonucleotides together and extending with sequenase according to manufacturers instructions. The sense and anti-sense (pTREP<sub>F</sub> and pTREP<sub>R</sub>) oligonucleotides contained the recognition sites for EcoRV and BamHI at their 5' ends respectively to facilitate cloning into pTREX7. The transcription terminator was that of the *Bacillus penicillinase* gene, which has been shown to be effective in *Lactococcus* (Jos *et al.*, *Applied and Environmental Microbiology* 50:540-542 (1985)). This was considered necessary as expression of target genes in the pTREX vectors was observed to be leaky and is thought to be the result of cryptic promoter activity in the origin region (Schofield *et al.* pers. coms. University of Cambridge Dept. Pathology.). The forward pUC primer sequencing was included to enable direct sequencing of cloned DNA fragments. The translation stop sequence which encodes a stop codon in 3 different frames was included to prevent translational fusions between vector genes and cloned DNA fragments. The pTREX7 vector was first digested with EcoRI and blunted using the 5' - 3' polymerase activity of T4 DNA polymerase (NEB) according to manufacturer's instructions. The EcoRI digested and blunt ended pTREX7 vector was then digested with Bgl II thus removing the P7 promoter. The artificial DNA fragment derived from the annealed synthetic oligonucleotides was then digested with EcoRV and Bam HI and cloned into the EcoRI(blunted)-Bgl II digested pTREX7 vector to

generate pTREP. A *Lactococcus lactis* MG1363 chromosomal promoter designated P1 was then cloned between the EcoRI and BglII sites present in the pTREP expression cassette forming pTREP1. This promoter was also isolated using the promoter probe vector pSB292 and characterised by Waterfield *et al.*, (1995) [*supra*]. The P1 promoter fragment was originally amplified by PCR using vent DNA polymerase according to manufacturers instructions and cloned into the pTREX as an EcoRI-BglII DNA fragment. The EcoRI-BglII P1 promoter containing fragment was removed from pTREX1 by restriction enzyme digestion and used for cloning into pTREP (Schofield *et al.* pers. coms. University of Cambridge, Dept. Pathology.).

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**(b) PCR amplification of the *S. aureus nuc* gene.**

The nucleotide sequence of the *S. aureus nuc* gene (EMBL database accession number V01281) was used to design synthetic oligonucleotide primers for PCR amplification. 15 The primers were designed to amplify the mature form of the *nuc* gene designated *nucA* which is generated by proteolytic cleavage of the N-terminal 19 to 21 amino acids of the secreted propeptide designated Snase B (Shortle, 1983 [*supra*]). Three sense primers (*nucS1*, *nucS2* and *nucS3*, shown in figure 3) were designed, each one having a blunt-ended restriction endonuclease cleavage site for EcoRV or SmaI in a 20 different reading frame with respect to the *nuc* gene. Additionally BglII and BamHI were incorporated at the 5' ends of the sense and anti-sense primers respectively to facilitate cloning into BamHI and BglII cut pTREP1. The sequences of all the primers are given in figure 3. Three *nuc* gene DNA fragments encoding the mature form of the nuclelease gene (*NucA*) were amplified by PCR using each of the sense primers 25 combined with the anti-sense primer. The *nuc* gene fragments were amplified by PCR using *S. aureus* genomic DNA template, Vent DNA Polymerase (NEB) and the conditions recommended by the manufacturer. An initial denaturation step at 93°C for 2 min was followed by 30 cycles of denaturation at 93°C for 45 sec, annealing at 50°C for 45 seconds, and extension 73°C for 1 minute and then a final 5 min extension step

at 73°C. The PCR amplified products were purified using a Wizard clean up column (Promega) to remove unincorporated nucleotides and primers.

**(c) Construction of the pTREP1-nuc vectors**

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The purified *nuc* gene fragments described in section b were digested with Bgl II and BamHI using standard conditions and ligated to BamHI and BglII cut and dephosphorylated pTREP1 to generate the pTREP1-*nuc1*, pTREP1-*nuc2* and pTREP1-*nuc3* series of reporter vectors. These vectors are described in figure 4.

10

General molecular biology techniques were carried out using the reagents and buffers supplied by the manufacturer or using standard techniques (Sambrook and Maniatis, Molecular cloning: A laboratory manual. Cold Spring Harbor Laboratory Press: Cold Spring Harbour (1989)). In each of the pTREP1-*nuc* vectors the expression cassette comprises a transcription terminator, lactococcal promoter P1, unique cloning sites (BglII, EcoRV or SmaI) followed by the mature form of the *nuc* gene and a second transcription terminator. Note that the sequences required for translation and secretion of the *nuc* gene were deliberately excluded in this construction. Such elements can only be provided by appropriately digested foreign DNA fragments (representing the target bacterium) which can be cloned into the unique restriction sites present

15

immediately upstream of the *nuc* gene.

20

**(d) Screening for secreted proteins in Group B Streptococcus.**

Genomic DNA isolated from and *Group B Streptococcus* (*S. agalactiae*) was digested with the restriction enzyme Tru9I. This enzyme which recognises the sequence 5'-TTAA -3' was used because it cuts A/T rich genomes efficiently and can generate random genomic DNA fragments within the preferred size range (usually averaging 0.5 - 1.0 kb). This size range was preferred because there is an increased probability that the P1 promoter can be utilised to transcribe a novel gene sequence. However, the P1 promoter may not be necessary in all cases as it is possible that many Streptococcal promoters are recognised in *L. lactis*. DNA fragments of different size ranges were

purified from partial Tru9I digests of *S. agalactiae* genomic DNA. As the Tru 9I restriction enzyme generates staggered ends the DNA fragments had to be made blunt ended before ligation to the EcoRV or SmaI cut pTREP1-*nuc* vectors. This was achieved by the partial fill-in enzyme reaction using the 5'-3' polymerase activity of 5 Klenow enzyme. Briefly Tru9I digested DNA was dissolved in a solution (usually between 10-20 µl in total) supplemented with T4 DNA ligase buffer (New England Biolabs; NEB) (1X) and 33 µM of each of the required dNTPs, in this case dATP and dTTP. Klenow enzyme was added (1 unit Klenow enzyme (NEB) per µg of DNA) and the reaction incubated at 25°C for 15 minutes. The reaction was stopped by incubating 10 the mix at 75°C for 20 minutes. EcoRV or SmaI digested pTREP-*nuc* plasmid DNA was then added (usually between 200-400 ng). The mix was then supplemented with 400 units of T4 DNA ligase (NEB) and T4 DNA ligase buffer (1X) and incubated overnight at 16°C. The ligation mix was precipitated directly in 100% Ethanol and 1/10 15 volume of 3M sodium acetate (pH 5.2) and used to transform *L. lactis* MG1363 (Gasson, *J. Bacteriol.* 154:1-9 (1983)). Alternatively, the gene cloning site of the pTREP-*nuc* vectors also contains a BglII site which can be used to clone for example Sau3AI digested genomic DNA fragments.

*L. lactis* transformant colonies were grown on brain heart infusion agar and nuclease 20 secreting (*Nuc*<sup>+</sup>) clones were detected by a toluidine blue-DNA-agar overlay (0.05 M Tris pH 9.0, 10 g of agar per litre, 10 g of NaCl per liter, 0.1 mM CaCl<sub>2</sub>, 0.03% wt/vol. salmon sperm DNA and 90 mg of Toluidine blue O dye) essentially as described by Shortle, 1983 [*supra*], and Le Loir *et al.*, 1994 [*supra*]). The plates were then incubated at 37°C for up to 2 hours. Nuclease secreting clones develop an easily 25 identifiable pink halo. Plasmid DNA was isolated from *Nuc*<sup>+</sup> recombinant *L. lactis* clones and DNA inserts were sequenced on one strand using the *NucSeq* sequencing primer described in figure 3, which sequences directly through the DNA insert.

Whilst the example described above related specifically to *Group B Streptococcus*, it will be apparent to one skilled in the art that the same screening technique may be used to detect exported and secreted proteins in other gram positive bacteria, for example *Streptococcus pneumoniae*.

5 **Example 2; Screening Group B Streptococcal derived genes in DNA vaccination experiments.**

**pcDNA3.1+ as a DNA vaccine vector**

10 The commercially available pcDNA3.1+ plasmid (Invitrogen), referred to as pcDNA3.1 henceforth, was used as a vector in all DNA immunisation experiments involving gene targets derived using the LEEP system. pcDNA 3.1 is designed for high-level stable and transient expression in mammalian cells and has been used widely and successfully as a host vector to test candidate genes from a variety of pathogens in DNA vaccination experiments (Zhang *et al.*, 1997; Kurar and Splitter, 15 1997; Anderson *et al.*, 1996).

20 The vector possesses a multiple cloning site which facilitates the cloning of multiple gene targets downstream of the human cytomegalovirus (CMV) immediate-early promoter/enhancer which permits efficient, high-level expression of the target gene in a wide variety of mammalian cells and cell types including both muscle and immune cells. This is important for optimal immune response as it remains unknown as to which cells types are most important in generating a protective response *in vivo*. The 25 plasmid also contains the ColE1 origin of replication which allows convenient high-copy number replication and growth in *E. coli* and the ampicillin resistance gene (B-lactamase) for selection in *E. coli*. In addition pcDNA 3.1 possesses a T7 promoter/priming site upstream of the MCS which allows for *in vitro* transcription of a cloned gene in the sense orientation.

30 **Preparation of DNA vaccines**

Oligonucleotide primers were designed for each individual gene of interest derived using the LEEP system. Each gene was examined thoroughly, and where possible, primers were designed such that they targeted that portion of the gene thought to

encode only the mature portion of the protein (**APPENDIX I**). It was hoped that expressing those sequences that encode only the mature portion of a target gene protein, would facilitate its correct folding when expressed in mammalian cells. For example, in the majority of cases primers were designed such that putative N-terminal signal peptide sequences would not be included in the final amplification product to be cloned into the pcDNA3.1 expression vector. The signal peptide directs the polypeptide precursor to the cell membrane via the protein export pathway where it is normally cleaved off by signal peptidase I (or signal peptidase II if a lipoprotein). Hence the signal peptide does not make up any part of the mature protein whether it be displayed on the bacterium's surface or secreted. Where a N-terminal leader peptide sequence was not immediately obvious, primers were designed to target the whole of the gene sequence for cloning and ultimately, expression in pcDNA3.1.

All forward and reverse oligonucleotide primers incorporated appropriate restriction enzyme sites to facilitate cloning into the pcDNA3.1 MCS region. All forward primers were also designed to include the conserved Kozak nucleotide sequence 5'-gccacc-3' immediately upstream of an 'atg' translation initiation codon in frame with the target gene insert. The Kozak sequence facilitates the recognition of initiator sequences by eukaryotic ribosomes. Typically, a forward primer incorporating a BamH1 restriction enzyme site the primer would begin with the sequence 5'-cgggatccgccaccatg-3', followed by a sequence homologous to the 5' end of that part of a gene being amplified. All reverse primers incorporated a Not I restriction enzyme site sequence 5'-ttgcggccgc-3'. All gene-specific forward and reverse primers were designed with compatible melting temperatures to facilitate their amplification.

All gene targets were amplified by PCR from *S. agalactiae* genomic DNA template using Vent DNA polymerase (NEB) or rTth DNA polymerase (PE Applied Biosystems) using conditions recommended by the manufacturer. A typical amplification reaction involved an initial denaturation step at 95°C for 2 minutes followed by 35 cycles of denaturation at 95°C for 30 seconds, annealing at the appropriate melting temperature for 30 seconds, and extension at 72°C for 1 minute (1 minute per kilobase of DNA being amplified). This was followed by a final extension period at 72°C for 10 minutes. All PCR amplified products were extracted once with phenol chloroform (2:1:1) and once with chloroform (1:1) and ethanol precipitated.

Specific DNA fragments were isolated from agarose gels using the QIAquick Gel Extraction Kit (Qiagen). The purified amplification gene DNA fragments were digested with the appropriate restriction enzymes and cloned into the pcDNA3.1 plasmid vector using *E. coli* as a host. Successful cloning and maintenance of genes was confirmed by restriction mapping and by DNA sequencing. Recombinant plasmid DNA was isolated on a large scale (>1.5 mg) using Plasmid Mega Kits (Qiagen).

It was decided to include the *S. agalactiae* rib gene as a positive control in at least one trial of DNA immunisation experiments. Rabbit antiserum against the Rib protein

10 (Stalhammar-Carleman *et al.*, 1993) and highly purified preparations of the Rib protein itself (Larsson *et al.*, 1999; Larsson *et al.*, 1996) have been shown to confer protection against lethal infection with strains expressing the antigen. All serotype III strains have been shown to express the Rib antigen and Southern blot analysis performed in the laboratory has confirmed that *S. agalactiae* serotype III (strain 97/0099) does contain the rib gene, hence the rib gene as part of a DNA vaccine would represent a potential positive control for all DNA immunisation experiments. Oligonucleotide primers were designed (**Appendix I**) that targeted only the mature portion of the rib gene and which included appropriate restriction enzyme sites for cloning into pcDNA3.1. rib was amplified using rTth DNA polymerase (PE Applied Biosystems) using conditions recommended by the manufacturer. Conditions for cloning were similar to that described previously.

#### **Preparation of a *S. agalactiae* standard inoculum**

##### **25 Strain validation**

*S. agalactiae* serotype III (strain 97/0099) is a recent clinical isolate derived from the

cerebral spinal fluid of a new born baby suffering from meningitis. This haemolytic

strain of Group B Streptococcus was epidemiologically tested and validated at the

Respiratory and Systemic Infection Laboratory, PHLS Central Public health

30 laboratory, 61 Collindale Avenue, London NW9 5HT. The strain was subcultured only

twice prior to its arrival in the laboratory. Upon its arrival on a agar slope, a sweep of

4-5 colonies was immediately used to inoculate a Todd Hewitt/5% horse blood broth

which was incubated overnight statically at 37 °C. 0.5 ml aliquots of this overnight

culture were then used to make 20% glycerol stocks of the bacterium for long term

storage at -70 °C. Glyerol stocks were streaked on Todd Hewitt/5% horse blood agar plates to confirm viability.

5      ***In vivo* passaging of Group B Streptococcus**

A frozen culture (described under strain validation) of *S. agalactiae* serotype III (strain 97/0099) was streaked to single colonies on Todd-Hewitt/5% blood agar plates which were incubated overnight at 37°C. A sweep of 4-5 colonies was used to inoculate a Todd Hewitt/5% horse blood broth which was again incubated overnight. A 0.5 ml aliquot from this overnight culture was used to inoculate a 50 ml Todd Hewitt broth (1:100 dilution) which was incubated at 37 °C. 10-fold serial dilutions of the overnight culture were made (since virulence of this strain was unknown) and each were passaged intra-peritoneally (IP) in CBA/ca mice in duplicate. Viable counts were performed on the various inocula used in the passage. Groups of mice were challenged with various concentrations of the pathogen ranging from  $10^8$  to  $10^4$  colony forming units (cfu). Mice that developed symptoms were terminally anaesthetized and cardiac punctures were performed (Only mice that had been challenged with the highest doses, i.e.  $1 \times 10^8$  cfu, developed symptoms). The retrieved unclotted blood was used to inoculate directly a 50ml serum broth (Todd Hewitt/20% inactivated foetal calf serum). The culture was constantly monitored and allowed to grow to late logarithmic phase. The presence of blood in the medium interfered with OD<sub>600</sub> readings as it was being increasingly lysed with increasing growth of the bacterium, hence the requirement to constantly monitor the culture. Upon reaching late logarithmic phase/early stationary phase, the culture was transferred to a fresh 50 ml tube in order to exclude dead bacterial cells and remaining blood cells which would have sedimented at the bottom of the tube. 0.5 ml aliquots were then transferred to sterile cryovials, frozen in liquid nitrogen and stored at -70 °C. A viable count was carried out on a single standard inoculum aliquot in order to determine bacterial numbers. This was determined to be approximately  $5 \times 10^8$  cfu per ml.

30      **Intra-peritoneal Challenge and virulence testing of Group B Streptococcus standard inoculum**

To determine if the standard inoculum was suitably virulent for use in a vaccine trial, challenges were carried out using a dose range. Frozen standard inoculum strain

5 aliquots were allowed to thaw at room temperature. From viable count data the number of cfu per ml was already known for the standard inoculum. Initially, serial dilutions of the standard inoculum were made in Todd Hewitt broth and mice were challenged intra-peritoneally with doses ranging from  $1 \times 10^8$  to  $1 \times 10^4$  cfu in a 500  $\mu$ l volume of Todd Hewitt broth. The survival times of mouse groups injected with different doses of the bacterium were compared. The standard inoculum was determined to be suitably virulent and a dose of  $1 \times 10^6$  cfu was considered close to optimal for further use in vaccine trials. Further optimisation was carried out by comparing mice challenged with doses ranging between  $5 \times 10^5$  and  $5 \times 10^6$  cfu. The 10 optimal dose was estimated to be approximately  $2.5 \times 10^6$  cfu. This represented a 100% lethal dose and was repeatedly consistent with end-points as determined by survival times being clustered within a narrow time-range. Throughout all these experiments, challenged mice were constantly monitored to clarify symptoms, stages of symptom development as well as calculating survival times.

15

#### **Vaccine trials**

20 Vaccine trials in mice were accomplished by the administration of DNA to 6 week old CBA/ca mice (Harlan, UK). Mice to be vaccinated were divided into groups of six and each group were immunised with recombinant pcDNA3.1 plasmid DNA containing a specific target-gene sequence derived using the LEEP system. A total of 100  $\mu$ g of DNA in Dulbecco's PBS (Sigma) was injected intramuscularly into the tibialis anterior muscle of both hind legs. Four weeks later this procedure was repeated using the same amount of DNA. For comparison, control mice groups were included in all vaccine trials. These control groups were either not DNA-vaccinated or were immunised with 25 non-recombinant pcDNA3.1 plasmid DNA only, using the same time course described above. Four weeks after the second immunisation, all mice groups were challenged intra-peritoneally with a lethal dose of *S. agalactiae* serotype III (strain 97/0099). The actual number of bacteria administered was determined by plating serial dilutions of the inoculum on Todd-Hewitt/5% blood agar plates. All mice were killed 3 or 4 days 30 after infection. During the infection process, challenged mice were monitored for the development of symptoms associated with the onset of *S. agalactiae* induced-disease. Typical symptoms in an appropriate order included piloerection, an increasingly hunched posture, discharge from eyes, increased lethargy and reluctance to move which was often the result of apparent paralysis in the lower body/hind leg region. The

latter symptoms usually coincided with the development of a moribund state at which stage the mice were culled to prevent further suffering. These mice were deemed to be very close to death, and the time of culling was used to determine a survival time for statistical analysis. Where mice were found dead, a survival time was calculated by 5 averaging the time when a particular mouse was last observed alive and the time when found dead, in order to determine a more accurate time of death.

### Interpretation of Results

A positive result was taken as any DNA sequence that was cloned and used in

10 challenge experiments as described above and gave protection against that challenge. DNA sequences were determined to be protective;

-if that DNA sequence gave statistically significant protection (to a 95% confidence level ( $p>0.05$ ) as determined using the Mann-Whitney U test.

15 -if that DNA sequence was marginal or non-significant using Mann-Whitney but showed some protective features. For example, one or more outlying mice may survive for significantly longer time periods when compared with control mice. Alternatively, the time to first death may also be prolonged when compared to counterpart mice in control groups.

20 It is acceptable to allow marginal or non-significant results to be considered as potential positives when it is possible that the clarity of some results may be affected by problems associated with the administration of the DNA vaccine. Indeed, much varied survival times may reflect different levels of immune response between different members of a given group.

25

### Results

#### Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 1 (Figure 4a)

30

	Mean Survival Times (hours)				
	pcDNA3.1	17(ID-8)	18(ID-9)	20(ID-25)	rib
1	26.833	14.916	27.750	30.500	88.666

2	42.333	94.000 (T)	34.333	33.333	28.166
3	47.916	45.166	41.083	34.083	37.250
4	28.333	30.750	47.083	23.500	37.250
5	42.333	74.666	94.000 (T)	94.000 (T)	94.000 (T)
6	25.333	25.000	26.166	30.500	45.750
<b>Mean</b>	<b>37.549</b>	<b>51.899</b>	<b>48.849</b>	<b>43.083</b>	<b>57.066</b>
<b>sd</b>	<b>9.3943</b>	<b>32.214</b>	<b>26.257</b>	<b>28.768</b>	<b>31.556</b>
<b>p value 1</b>		<b>0.4049</b>	<b>0.4049</b>	<b>0.5000</b>	<b>0.1481</b>
<b>p value 2</b>	<b>&gt; 39.0</b>	<b>&gt; 39.0</b>	<b>&gt; 39.0</b>	<b>&gt; 39.0</b>	

(T) - terminated at conclusion of experiment but showing symptoms of infection.

5      p value 1 refers to statistical significance when compared to pcDNA3.1 controls.

    p value 2 refers to statistical significance when compared to rib positive control.

10

All DNA vaccine's showed a pattern of protection similar to that obtained with the rib DNA vaccine, which was initially used as a positive control.

15

### **17 (ID-8)**

20

Mice immunised with the '17 (ID-8)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there are two outlying mice one of which survived the term of the experiment despite developing symptoms. The group also exhibited a much wider range of survival times reflected by a mean survival value which is approximately 14 hours higher than that demonstrated by the unvaccinated control group.

25

### **18 (ID-9)**

5 Mice immunised with the '18 (ID-9)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there is one outlying mouse which survived the term of the experiment despite developing symptoms.

## 20 (ID-25)

10 Mice immunised with the '20 (ID-25)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there was one outlying mouse which survived the term of the experiment despite developing symptoms.

15 **Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 2 (Figure 4b)**

	Mean Survival Times (hours)			
	pcDNA	UnVacc	22(ID-10)	28(ID-13)
1	45.000	27.916	44.666	72.000 (T)
2	37.333	45.083	51.416	33.000
3	37.333	37.583	40.791	36.083
4	35.291	24.583	44.666	72.000 (T)
5	24.333	37.583	36.916	49.166
6	45.000	33.166	57.833	36.083
<b>Mean</b>	<b>35.858</b>	<b>34.549</b>	<b>43.691</b>	<b>52.449</b>
<b>sd</b>	<b>7.4342</b>	<b>8.2567</b>	<b>5.3825</b>	<b>18.850</b>
<b>p value 1</b>		<b>&gt; 39.0</b>	<b>0.1137</b>	<b>0.2340</b>
<b>p value 2</b>	<b>0.4679</b>		<b>0.0323</b>	<b>0.1137</b>

20 (T) - terminated at conclusion of experiment but showing symptoms of infection.

**p value 1** refers to statistical significance when compared to pcDNA3.1 controls.

**p value 2** refers to statistical significance when compared to unvaccinated controls.

5 There was no significant difference in the survival times exhibited by the pcDNA3.1 and unvaccinated control groups. This is confirmed by their very similar mean survival times of 35.858 hours (pcDNA3.1) and 34.166 hours (Unvaccinated).

10

### **22 (ID-10)**

15 Mice immunised with the '22 (ID-10)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group but not when compared with the pcDNA3.1 control group. In addition, the time to first death in this group was prolonged by approximately 12 hours when compared to the pcDNA3.1 and unvaccinated control groups. The mean survival time of 43.691 hours is also considerably higher than that determined for both control groups.

20

### **28 (ID-13)**

25 Mice immunised with the '28 (ID-13)' DNA vaccine did not show significantly longer survival times when compared with the pcDNA3.1 and unvaccinated control groups. However there are three outlying mice, two of which survived the term of the experiment despite showing symptoms. In addition, the time to first death in this group was prolonged by approximately 9 hours when compared to the pcDNA3.1 and unvaccinated control groups. The mean survival time of 52.449 hours is also considerably higher than that determined for both control groups, as well demonstrating a wider range of survival times.

30

### **Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 3 (Figure 4c)**

35

	Mean Survival Times (hours)				
	UnVacc.	70(ID-42)	94(ID-48)	86(ID-47)	51(ID-37)
1	27.583	25.166	34.666	32.416	43.749
2	27.583	42.666	49.500	32.416	38.333
3	24.583	34.666	27.000	42.500	50.666
4	22.250	42.666	30.500	34.500	45.166
5	35.916	30.583	30.500	34.500	69.082
6	22.250	25.166	42.666	42.500	31.166
<b>Mean</b>	<b>27.583</b>	<b>35.149</b>	<b>34.433</b>	<b>35.266</b>	<b>49.399</b>
<b>sd</b>	<b>5.1691</b>	<b>7.6444</b>	<b>8.8495</b>	<b>4.1758</b>	<b>11.846</b>
<b>p value</b>		<b>0.0628</b>	<b>0.0321</b>	<b>0.0153</b>	<b>0.0041</b>

5      **p value** refers to statistical significance when compared to unvaccinated controls.

### 70 (ID-42)

10     Mice immunised with the '70 (ID-42)' DNA vaccine, marginally did not show significantly longer survival times when compared with the unvaccinated control group. However, the first death in this group is prolonged (by approximately 3 hours ) when compared with the unvaccinated group. In addition, the group has a mean survival time which is approximately 8 hours longer than the unvaccinated group.

15

### 94 (ID-48)

20     Mice immunised with the '94 (ID-48)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

### 86 (ID-47)

Mice immunised with the '86 (ID-47)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

5

### **51 (ID-37)**

Mice immunised with the '51 (ID-37)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

10

#### **Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 4 (Figure 4d)**

15

	Mean Survival Times (hours)	
	UnVacc	9(ID-6)
1	32.666	35.250
2	21.666	30.958
3	23.916	69.333
4	22.999	52.333
5	25.916	44.916
6	35.916	47.083
<b>Mean</b>	<b>25.432</b>	<b>46.041</b>
<b>sd</b>	<b>4.3291</b>	<b>16.096</b>
<b>p value</b>		<b>0.0101</b>

(T) - terminated at conclusion of experiment but showing symptoms of infection.

20

**p value** refers to statistical significance when compared to unvaccinated controls

### **9 (ID-6)**

Mice immunised with the '39(ID-6)' DNA vaccine showed significantly longer survival times when compared with the control group.

5

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 6 (Figure 4e)**

10

	Mean Survival Times (hours)				
	pcDNA	UnVacc	32 (ID-15)	39(ID-17)	57(40)
1	33.541	36.000	25.041	52.333	28.333
2	36.750	29.999	30.458	44.750	32.708
3	36.750	32.749	44.833	44.750	36.083
4	36.750	44.500	30.458	36.250	40.333
5	29.000	28.333	64.833	36.250	72.000 (T)
6	30.750	31.666	72.000 (T)	28.583	33.750
<b>Mean</b>	<b>34.558</b>	<b>34.316</b>	<b>39.124</b>	<b>44.016</b>	<b>38.103</b>
<b>sd</b>	<b>3.4036</b>	<b>6.3921</b>	<b>16.140</b>	<b>13.833</b>	<b>12.986</b>
<b>p value 1</b>		<b>&gt; 39.0</b>	<b>0.4043</b>	<b>0.1867</b>	<b>0.4044</b>
<b>p value 2</b>	<b>0.2862</b>		<b>0.2873</b>	<b>0.0458</b>	<b>0.2113</b>

15 (T) - terminated at conclusion of experiment but showing symptoms of infection.

**p value 1** refers to statistical significance when compared to pcDNA3.1 controls

**p value 2** refers to statistical significance when compared to unvaccinated controls.

20

There was no significant difference in the survival times exhibited by the pcDNA3.1 and unvaccinated control groups. This is confirmed by their

very similar mean survival times of 34.558 hours (pcDNA3.1) and 34.316 hours (Unvaccinated).

5 **32 (ID-15)**

10 Mice immunised with the '32 (ID-15)' DNA vaccine did not show significantly longer survival times when compared with the pcDNA3.1 and unvaccinated control groups. However, the '32 (ID-15)' group has two outlying mice one of which survived the term of the experiment despite showing symptoms. This group also exhibits a wide range of survival times.

15 **39 (ID-17)**

20 Mice immunised with the '39 (ID-17)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group but was not significant when compared with the pcDNA3.1 control group. The group has a considerably higher mean survival time of 44.016 hours than that determined for either of the control groups.

25 **57 (ID-40)**

30 Mice immunised with the '32 (ID-15)' DNA vaccine did not show significantly longer survival times when compared with the pcDNA3.1 and unvaccinated control groups. However, the '32 (ID-15)' group has one outlying mouse which survived the term of the experiment despite showing symptoms.

35 **SURVIVAL DATA-SET B**

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 2 (Figure 5a)**

	Mean Survival Times (hours)		
	pcDNA	UnVacc	13(ID-72)
1	45.000	27.916	69.166
2	37.333	45.083	36.333
3	37.333	37.583	43.916
4	35.291	24.583	32.166
5	24.333	37.583	36.333
6	45.000	33.166	43.916
Mean	35.858	34.549	43.582
sd	7.4342	8.2567	14.917
p value 1		> 39.0	0.4679
p value 2	0.4679		0.1880

5 **p value 1** refers to statistical significance when compared to pcDNA3.1 controls.

**p value 2** refers to statistical significance when compared to unvaccinated controls.

10 There was no significant difference in the survival times exhibited by the pcDNA3.1 and unvaccinated control groups. This is confirmed by their very similar mean survival times of 35.858 hours (pcDNA3.1) and 34.166 hours (Unvaccinated).

15

### **13 (ID-72)**

20 Mice immunised with the '13 (ID-72)' DNA vaccine did not show significantly longer survival times when compared with the pcDNA3.1 and unvaccinated control groups. However, there is one outlying mouse which survived approximately 24 hours longer than the longest surviving mice in the pcDNA3.1 and unvaccinated control groups respectively. In addition, the time to first death in this group was prolonged when

compared to the pcDNA3.1 and unvaccinated control groups. The mean survival time of 43.582 hours is considerably higher than that determined for both control groups.

5

10

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 3 (Figure 5b)**

	Mean Survival Times (hours)		
	UnVacc	3-60(ID-65)	3-5(ID-66)
1	27.583	54.416	42.916
2	27.583	31.000	42.916
3	24.583	43.000	32.874
4	22.250	34.916	42.916
5	35.916	38.958	27.333
6	22.250	34.916	30.916
<b>Mean</b>	<b>27.583</b>	<b>40.458</b>	<b>37.791</b>
<b>sd</b>	<b>5.1691</b>	<b>8.9959</b>	<b>7.2860</b>
<b>p value</b>		<b>0.0098</b>	<b>0.0215</b>

15

**p value** refers to statistical significance when compared to unvaccinated controls.

20

**3-60 (ID-65)**

Mice immunised with the '3-60 (ID-65)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

5      **3-5 (ID-66)**

Mice immunised with the '3-5 (ID-66)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 4 (Figure 5c)**

	Mean Survival Times (hours)			
	UnVacc	3-40(ID-67)	3-30(ID-68)	3-38(ID-69)
1	32.666	79.750	35.500	68.583
2	21.666	35.833	28.333	29.916
3	23.916	30.500	31.208	29.916
4	22.999	22.708	98.000 (T)	31.041
5	25.916	28.583	73.500	32.166
6	35.916	40.791	32.333	29.916
<b>Mean</b>	<b>25.432</b>	<b>39.474</b>	<b>53.308</b>	<b>38.324</b>
<b>sd</b>	<b>4.3291</b>	<b>22.998</b>	<b>30.961</b>	<b>16.940</b>
<b>p value</b>		<b>0.1149</b>	<b>0.0463</b>	<b>0.1132</b>

5

(T) - terminated at conclusion of experiment but showing symptoms of infection.

10 **p** value refers to statistical significance when compared to unvaccinated controls

### 3-40 (ID-67)

15 Mice immunised with the '3-40 (ID-67)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there is one outlying mouse which survived approximately 43 hours longer than the longest surviving mice in the unvaccinated control group.

### 20 3-30 (ID-68)

Mice immunised with the '3-30 (ID-68)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

**3-38 (ID-69)**

5 Mice immunised with the '2-19 (ID-73)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there was one outlying mouse which survived approximately 32 hours longer than the longest surviving mice in the unvaccinated control group. In addition, the time to first death was prolonged (by approximately 8 hours) when compared to the  
 10 unvaccinated controls.

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 5 (Figure 5d)**

	Mean Survival Times (hours)				
	UnVacc	141(ID-70)	3-20(ID-71)	2-19(ID-73)	3-6(ID-74)
1	27.833	47.500	36.166	36.166	44.666
2	45.666	52.833	44.833	49.833	36.000
3	45.666	49.333	26.750	55.833	75.416
4	34.333	46.250	36.166	68.583	36.000
5	34.333	47.500	55.916	33.333	55.916
6	45.666	36.500	44.833	30.583	36.000
<b>Mean</b>	<b>37.566</b>	<b>48.683</b>	<b>37.234</b>	<b>48.749</b>	<b>49.599</b>
<b>sd</b>	<b>7.8558</b>	<b>2.5672</b>	<b>8.4103</b>	<b>14.497</b>	<b>16.587</b>
<b>p value</b>		<b>0.0101</b>	<b>0.5000</b>	<b>0.2336</b>	<b>0.1854</b>

15

**p value** - refers to statistical significance when compared to unvaccinated controls.

20

**141 (ID-70)**

Mice immunised with the '141 (ID-70)' DNA vaccine exhibited significantly longer survival times when compared with the unvaccinated control group.

**3-20 (ID-71)**

5 Mice immunised with the '3-20 (ID-71)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there is one outlying mouse which survived approximately 10 hours longer than the longest surviving mice in the unvaccinated control group.

**2-19 (ID-73)**

10 Mice immunised with the '2-19 (ID-73)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there are three outlying mouse which survived approximately 4, 10 and 23 hours longer than the longest surviving mice 15 in the unvaccinated control group. This is reflected in the higher mean survival time of 48.749 hours and a much wider range of survival times.

**3-6 (ID-74)**

20 Mice immunised with the '3-6 (ID-74)' DNA vaccine did not show significantly longer survival times when compared with the unvaccinated control group. However, there are three outlying mouse which survived approximately 4, 10 and 23 hours longer than the longest surviving mice 25 in the unvaccinated control group. This is reflected in the higher mean survival time of 49.599 hours and a much wider range of survival times.

**Statistical analysis of survival times - LEEP DNA immunisation and GBS challenge Trial 6 (Figure 5e)**

30

	Mean Survival Times (hours)			
	pcDNA3.1	UnVacc.	3-51(ID-75)	3-56 (ID-76)

1	33.541	36.000	36.333	46.583
2	36.750	29.999	30.291	29.833
3	36.750	32.749	32.000	40.166
4	36.750	44.500	52.333	46.583
5	29.000	28.333	72.000 (T)	46.583
6	30.750	31.666	40.499	---
<b>Mean</b>	<b>34.558</b>	<b>34.316</b>	<b>44.591</b>	<b>40.791</b>
<b>sd</b>	<b>3.4036</b>	<b>6.3921</b>	<b>16.615</b>	<b>7.9070</b>
<b>p value 1</b>		<b>&gt; 39.0</b>	<b>0.1876</b>	<b>0.0386</b>
<b>p value 2</b>	<b>0.2862</b>		<b>0.0867</b>	<b>0.0587</b>

(T) - terminated at conclusion of experiment but showing symptoms of infection.

5      **p value 1** refers to statistical significance when compared to pcDNA3.1 controls

**p value 2** refers to statistical significance when compared to unvaccinated controls.

10     There was no significant difference in the survival times exhibited by the pcDNA3.1 and unvaccinated control groups. This is confirmed by their very similar mean survival times of 34.558 hours (pcDNA3.1) and 34.316 hours (Unvaccinated).

15

### 3-51 (ID-75)

20     Mice immunised with the '3-51 (ID-75)' DNA vaccine did not show significantly longer survival times when compared with the pcDNA3.1 control group but was relatively close to significant when compared with the unvaccinated control group. The '3-51' group has two outlying mouse one of which survived the term of the experiment despite developing symptoms. The mean survival time of 44.499 hours is considerably higher than that determined for both control groups and the group also demonstrates as a much wider range of survival times.

**3-56 (ID-76)**

5 Mice immunised with the '3-56 (ID-76)' DNA vaccine exhibited significantly longer survival times when compared with the pcDNA3.1 control group but were marginally not significant when compared with unvaccinated control group.

10 **Example 3: Conservation and variability of candidate vaccine antigen genes among different isolates of Group B Streptococci**

An initial Southern blot analysis was carried out to determine cross-serotype conservation of novel Group B Streptococcal genes isolated using the LEEP system. Analysing the serotype distribution of a target gene will also determine their potential 15 use as antigen components in a GBS vaccine. The Group B Streptococcal strains whose DNA was analysed as part of this study are listed in **APPENDIX II**.

20 **Amplification and labelling of specific target genes as DNA probes for Southern blot analysis.**

Oligonucleotide primers were designed for each individual gene of interest derived using the LEEP system. Primers were designed to target the whole of the gene being investigated (All primers are listed in **APPENDIX III**). Specific gene targets were amplified by PCR using Vent DNA polymerase (NEB) according to the manufacturers 25 instructions. Typical reactions were carried out in a 100 µl volume containing 50 ng of GBS template DNA, a one tenth volume of enzyme reaction buffer, 1 µM of each primer, 250 µM of each dNTP and 2 units of Vent DNA polymerase. A typical reaction contained an initial 2 minute denaturation at 95°C, followed by 35 cycles of denaturation at 95°C for 30 seconds, annealing at the appropriate melting temperature for 30 seconds, and extension at 72°C for 1 minute (1 minute per kilobase of DNA 30 being amplified). The annealing temperature was determined by the lower melting temperature of the two oligonucleotide primers. The reaction was concluded with a final extension period of 10 minutes at 72°C.

All PCR amplified products were extracted once with phenol chloroform (2:1:1) and once with chloroform (1:1) and ethanol precipitated. Specific DNA fragments were isolated from agarose gels using the QIAquick Gel Extraction Kit (Qiagen). For use as DNA probes, purified amplified gene DNA fragments were labelled with digoxigenin using the DIG Nucleic Acid Labelling Kit (Boehringer Mannheim) according to the manufacturer's instructions.

**5 Southern blot hybridisation analysis of Group B Streptococcal genomic DNA**

Genomic DNA had previously been isolated from all strains of Group B Streptococci

10 which were investigated for conservation of LEEP-derived gene targets. Appropriate DNA concentrations were digested using either *Hin* DIII, *Eco* RI or *Bgl* II restriction enzymes (NEB) according to manufacturer instructions and analysed by agarose gel electrophoresis. Following agarose gel electrophoresis of DNA samples, the gel was denatured in 0.25M HCl for 20 minutes and DNA was transferred onto Hybond™ N<sup>+</sup> 15 membrane (Amersham) by overnight capillary blotting. The method is essentially as described in Sambrook *et al.* (1989) using Whatman 3MM wicks on a platform over a reservoir of 0.4M NaOH. After transfer, the filter was washed briefly in 2x SSC and stored at 4 °C in Saran wrap (Dow chemical company).

20 Filters were prehybridised, hybridised with the digoxigenin labelled DNA probes and washed using conditions recommended by Boehringer Mannheim when using their DIG Nucleic Acid Detection Kit. Filters were prehybridised at 68°C for one hour in hybridisation buffer (1% w/v supplied blocking reagent, 5x SSC, 0.1% v/v N-lauryl

25 sarkosine, 0.02% v/v sodium dodecyl sulphate[SDS]). The digoxigenin labelled DNA probe was denatured at 99.9°C for 10 minutes before being added to the hybridisation buffer. Hybridisation was allowed to proceed overnight in a rotating Hybaid tube in a Hybaid Mini-hybridisation oven. Unbound probe was removed by washing the filter twice with 2x SSC- 0.1% SDS for 5 minutes at room temperature. For increased stringency filters were then washed twice with 0.1x SSC-0.1% SDS for 15 minutes at 68°C. The DIG Nucleic Acid Detection Kit (Boehringer Mannheim) was used to 30 immunologically detect specifically bound digoxigenin labelled DNA probes.

### Results of Southern blot analysis

All genomic digests and their corresponding Southern blots followed an identical lane order as described in Table I.

5

**Table I**

	1 kb molecula r Weight Marker	515	A909	SB35	H36B	18RS21	1954/92
	Ia	Ia	Ib	Ib	II	II	

	118/158	97/0057	BM110	BS30	M781	97/0099	3139
	II	II	III	III	III	III	IV

	1169-NT	GBS 6	7271	JM9	Group A Streptococcu s	<i>Streptococcu</i> <i>pneumoniae</i>
	V	VI	VII	VIII	-	14

10 For comparative purposes, it was decided to analyse the serotype distribution of the GBS *rib* gene, which encodes the known protective immunogen Rib. Rib has previously been shown to be present in serotype III and some strains of serotype II but not in serotypes Ia or Ib (Stalhammar-Carlemalm *et al.*, 1993). Confirmation of this pattern would not only give increased confidence in interpreting subsequent results, it would also determine if a *rib* gene homologue was present in the remaining GBS

serotypes being investigated here. Primers designed for the amplification of *rib* and its subsequent cloning into pcDNA3.1 (Appendix I), were used to generate a *rib* gene probe for Southern blot analysis.

5      **Southern blot analysis - *rib* (Figure 6)**

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

10

Genomic DNA from each strain was digested completely with *Hin* DIII (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled *rib* gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

15      Comment

The Southern blot analysis described in Figure 7 indicates that the *rib* gene is not conserved across all GBS serotypes. *rib* appears to be absent from all serotype Ia and Ib strains (lanes 2 to 5) and from strains 118/158 and 97/0057 of serotype II (lanes 8 and 9). However, *rib* would appear to present in strains 18RS21 and 1954/92 of serotype II (lanes 6 and 7) and in all strains of serotype III (lanes 10 to 13). This is in agreement with previously published data (Stalhammar-Carlemalm *et al.*, 1993). *rib* would also appear to be present in strains representing serotypes VII and VII (lanes 17 and 18) but was absent from strains representing serotypes IV, V and V (lanes 14 to 16) as well as the control strains (lanes 19 and 20). The *rib* gene probe did hybridise with lower intensity to genomic DNA fragments from strains representing serotypes Ia, Ib, IV, VI, VII and serotype II strains 118/158 and 97/0057. This may indicate the presence of a gene in these strains with a lower level of homology to *rib*. These hybridising DNA fragments may contain a homologue of the GBS *bca* gene encoding the Ca protein antigen which has been shown to be closely homologous to the Rib protein (Wastfelt *et al.*, 1996). If this is the case, it would be in agreement with previous work which showed all strains of serotypes Ia, Ib, II and III to be positive for one the two proteins (Stalhammar-Carlemalm *et al.*, 1993). However, the apparent

variable distribution of the *rib* gene amongst different GBS serotypes, makes it a less than ideal candidate for use in a GBS vaccine that is cross-protective against all serotypes.

5 **Southern blot analysis - 4 (ID-1) (photograph 7)**

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

10 Genomic DNA from each strain was digested completely with *Hin* DIII (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled 4 (ID-1) gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

15 Comment

The Southern blot analysis described in Figure 7 indicates that gene 4 (ID-1) is conserved across all GBS serotypes. The gene probe hybridised specifically to a *Hin* DIII-digested genomic DNA fragment of approximately 3.5 kb in DNA digests from all GBS representatives. but was absent from both the control strains (lanes 19 and 20).

25 **Southern blot analysis - 5 (ID-2) (Figure 8)**

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

30 Genomic DNA from each strain was digested completely with *Eco* RI (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled 5 (ID-2) gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

Comment

The Southern blot analysis described in Figure 7 indicates that gene 4 (ID-1) is conserved across all GBS serotypes. The gene probe hybridised specifically to a *Eco* RI-digested genomic DNA fragment of approximately 6.2 kb in DNA digests from all GBS representatives. but was absent from both the control strains (lanes 19 and 20).

5

#### **Southern blot analysis - 15 (ID-7) (Figure 9)**

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

10

Genomic DNA from each strain was digested completely with *Eco* RI (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled 15 (ID-7) gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

15

#### Comment

The Southern blot analysis described in Figure 7 indicates that gene 15 (ID-7) is conserved across all GBS serotypes. The gene probe hybridised specifically to a *Eco* RI-digested genomic DNA fragment of approximately 6.2 kb in DNA digests from all GBS representatives. but was absent from both the control strains (lanes 19 and 20).

20

The gene probe hybridised specifically with *Eco* RI -digested DNA fragments ranging from approximately 3.5 kb to 5.2 kb in size.

25

#### **Southern blot analysis - 17 (ID-8) (Figure 10)**

#### **Figure 5**

1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

30

Genomic DNA from each strain was digested completely with *Hin* DIII (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled

17 (ID-8) gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

Comment

5 The Southern blot analysis described in Figure 7 indicates that gene 17 (ID-8) is conserved across all GBS serotypes. The gene probe hybridised specifically to a *Hin* DIII-digested genomic DNA fragment of approximately 2.3 kb in DNA digests from all GBS representatives. but was absent from both the control strains (lanes 19 and 20).

10

**Southern blot analysis - 22 (ID-10) (Figure 11)**

**Figure 6**

15 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20

20 Genomic DNA from each strain was digested completely with *Bgl* II (NEB) and electrophoresed at 40 Volts for 6 hours in 0.8% agarose, transferred onto Hybond N<sup>+</sup> (Amersham) membrane by Southern blot and hybridised with the digoxigenin-labelled 22 (ID-10) gene probe. Specifically bound DNA probe was identified using the DIG Nucleic Acid Detection Kit (Boehringer Mannheim).

Comment

25 The Southern blot analysis described in Figure 7 indicates that gene 22 (ID-10) is conserved across all GBS serotypes. The gene probe hybridised specifically to a *Bgl* II-digested genomic DNA fragment of approximately 3.1 kb in DNA digests from all GBS representatives except serotype Ib strain H36B, where the gene probe hybridised specifically to a *Bgl* II-digested genomic DNA fragment. Gene 22 (ID-10) was absent from both the control strains (lanes 19 and 20).

30

**Conclusion**

The Southern blot analyses described here, represents a preliminary investigation into the conservation level of LEEP-derived genes amongst different GBS serotypes. Initial results indicate that the genes 4 (ID-1), 5 (ID-2), 15 (ID-7), 17(ID-8) and 22

(ID-10) are present in all GBS serotypes and thus represent potential candidate genes for use in a GBS vaccine. Similar analyses are being currently carried out for each of the genes contained in this patent.

**APPENDIX I****ID-8 (17)**

Forward Primer

5' - cggatccgccaccatgACCACTTCTCAAGCTGTTTAGC - 3'

Reverse Primer

5' - ttgcggccgcACGATTATCAACAAAGTTCTG - 3'**ID-9 (18)**

10 Forward Primer

5' - cggatccgccaccatgGCTACTCATATTGGAAGTTACCAGC - 3'

Reverse Primer

5' - ttgcggccgcAGGGTTTATTGTTGAAGTGTCTTG - 3'**ID-10 (22)**

Forward Primer

5' - cggatccgccaccatgTATCTATATCATTACCAATGCC - 3'

Reverse Primer

5' - ttgcggccgcTTTATGTATAGAACAGCAGTCCC - 3'

20

**ID-13 (28)**

Forward Primer

5' - cggatccgccaccatgAAAGGAAGAACAAACCTATTGTTAG - 3'

Reverse Primer

25

5' - ttgcggccgcAAGAGCAAATTCGTATCTCCTC - 3'**ID-15 (32)**

Forward Primer

5' - cggatccgccaccATGATTGTTGGACACGGAATTG - 3'

30

Reverse Primer

5' - ttgcggccgcTTTTCTTCCTCCAAAATAACACTAGC - 3'**ID-17 (39)**

Forward Primer

5' - cggatccgcaccatgGCGACTAAAGAGTTAGGTGTTAG -3'

Reverse Primer

5' - ttcgccgcTATAGTTTAGTTCAACTTGTCTAGATG -3'

5 ID-25 (20)

Forward Primer

5' - cggatccgcaccatgTATACGAGTTACAACCAAATCATG -3'

Reverse Primer

5' - ttcgccgcGTCAGCTCGTACTGTTTTTAGC -3'

10

ID-37 (51)

Forward Primer

5' - cggatccgcaccatgTGTCAAATGAATAGTGAACATAAAAG -3'

Reverse Primer

15

5' - ttcgccgcCTCAAATAATTACCTCCAATTG -3'

ID-40 (51)

Forward Primer

5' - cggatccgcaccatgGCTCCATTGAAATTAAAGATTG -3'

20

Reverse Primer

5' - ttcgccgcTGATTTACCAGTTGGAAGAGTTC -3'

ID-42 (70)

Forward Primer

25

5' - cggatccgcaccATGAATACTATTATAATACATTGAGAACAG -3'

Reverse Primer

5' - ttcgccgcTTCTTGTCCAACTTCTGG -3'

ID-47 (86)

30

Forward Primer

5' - cggatccgcaccATGATAGAGTGGATTCAAACACATTAC -3'

Reverse Primer

5' - ttcgccgcTTTATGACTCAAGCGACGTGTTA -3'

ID-48 94

Forward Primer

5' - cggatccggcaccATGGAGTTAGTAATTAGAGATATTGTAAG

Reverse Primer

5' - ttcgccgcCTTGTCAATTACATCTCCCTCAACID-67 (3-40)

Forward Primer

5' - cggatccggcaccatgGCTAGTTTGTCAATGAATCATAATGAC -3'

10 Reverse Primer

5' - ttcgccgcGTTATTGCTCGTTAGCTAAATC -3'ID-68 (3-30)

Forward Primer

15 5' - cggatccggcaccatgGCTCTTAGTTTTATGGTTCAAGC -3'

Reverse Primer

5' - ttcgccgcGAAGGCACCGCCACCTCC -3'ID-69 (3-38)

20 Forward Primer

5' - cggatccggcaccatgGGTGAAACCCAAGATACCAATCAAGC -3'

Reverse Primer

5' - ttcgccgcAACACCTGGTGGCGTTGG -3'25 ID-70 (141)

Forward Primer

5' - cggatccggcaccATGGCTGGGAATCGTAATAACG -3'

Reverse Primer

5' - ttcgccgcAGCCGTCTCTAACACAGGCTTG -3'

30

ID-71 (3-20)

Forward Primer

5' - cggatccggcaccatgCTTCCAACCGCAGCCGAAAAC -3'

Reverse Primer

5' - ttgcggccgcATTTAGTGTATTCTCCTGTTGCATAATCC -3'

ID-72 (13)

Forward Primer

5 5' - cggatccaccatgTACACGCATATTGTTGAAAAAAAG -3'

Reverse Primer

5' - ttgcggccgcAAATAATTCTTTGGTTGTTG -3'

ID-73 (2-19)

10 Forward Primer

5' - cggatccgcaccatgAGTAATCAAGAAGTTCAGCAAGC -3'

Reverse Primer

5' - ttgcggccgcCCATTGTGGAATATCAGCTGAAG -3'

15 ID-74 (3-6)

Forward Primer

5' - cggatccgcaccatgGTGCAGGCAGTGGTACCGCT -3'

Reverse Primer

20 5' - ttgcggccgcGCGCATTGTAACAAATTCTCAG -3'

ID-75 (3-51)

Forward Primer

5' - cggatccaccatgGCTGCCGAGAAGGATAAAAG -3'

25 Reverse Primer

5' - ttgcggccgcATTATTTAGCTGCTTTTAATGG -3'

ID-76 (3-56)

Forward Primer

30 5' - cggatccaccatgTGTCAGGTTTTATGCAAGTTTC -3'

Reverse Primer

5' - ttgcggccgcTTTACTAATTGATAAAAGAGCAACTTCG -3'

*rib* (control)

Forward primer

5' - ggggtacggccaccATGGCTGAAGTAATTAGGAAGT -3'

Reverse primer

5' - cggaattccgTTAACCTTTTTCTTAGAACAGAT

## APPENDIX II

Listed below are the details (serotype and strain designation) of Group B Streptococcus strains whose DNA was analysed for gene conservation

5

**SEROTYPE**      **STRAIN**

	Ia	515
	Ia	A909
10	Ib	SB35
	Ib	H36B
	II	18RS21
	II	1954/92
	II	118/158
15	II	97/0057
	III	BM110
	III	BS30
	III	M781
	III	97/0099
20	IV	3139
	V	1169/NT
	VI	GBS VI
	VII	7271
	VIII	JM9

25

A group A Streptococcal strain (serotype M1, strain NCTC8198) and *Streptococcus pneumoniae* (serotype 14) were also included in the analysis for control purposes.

## APPENDIX III

ID-1 (4)

forward primer

5' - atggaaaaaaaaacttggaaaaaaaaattac -3'

reverse primer

5' - ctattttgttttagcgatgtcttatac -3'

ID-2 (5)

10 forward primer

5' - atgtcaaaacaaaaagtaacggcaac -3'

reverse primer

5' - ttatttatggccaataccataagttaattg

15 ID-6 (9)

forward primer

5' - atgaaaaaaaaagttttttctcatggctatg -3'

reverse primer

5' - ttacttcaactgtttagatagaggacttcc - 3'

20

ID-7 (15)

forward primer

5' - ttgttcaattttataggtttttagaacttgg -3'

reverse primer

25

5' - ttaattttcattgcgtctcaaacc -3'

ID-8 (17)

forward primer

5' - atgacaaaaaaaaacttattattgtatattag -3'

30

reverse primer

5' - ttaacgattatcaacaaagttctgtac -3'

ID-10 (22)

forward primer

5' - atgatacggcagttaagagaa -3'  
reverse primer  
5' - ttatgtatgtatagaaacagcagtccc -3'

5 **References**

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## Claims:

1. A *Group B Streptococcus* protein having a sequence selected from those described in fig 1, or fragments or derivatives thereof.

5

2. A *Group B Streptococcus* polypeptide or peptide having a sequence selected from those described in fig 2, or fragments or derivatives thereof.

10 3. Derivatives or variants of the proteins, polypeptides, and peptides as claimed in claims 1 and 2 which show at least 50% identity to those proteins, polypeptides and peptides claimed in claims 1 and 2.

4. A nucleic molecule comprising or consisting of a sequence which is:

15 (i) any of the DNA sequences set out in figure 1 and figure 2 herein or their RNA equivalents;

(ii) a sequence which is complementary to any of the sequences of (i);

(iii) a sequence which codes for the same protein or polypeptide, as those sequences of (i) or (ii);

20 (iv) a sequence which shows substantial identity with any of those of (i), (ii) and (iii); or

(v) a sequence which codes for a derivative, or fragment of a nucleic acid molecule shown in figure 1 or figure 2.

25 5. A vector comprising DNA encoding for the expression of any one or more proteins, polypeptides, peptides, fragments or derivatives thereof, as claimed in claims 1 to 3.

30 6. A vector as claimed in claim 5 further comprising DNA encoding any one or more of the following: promoters, enhancers, signal sequences, leader sequences,

translation start and stop signals, DNA stability controlling regions, or a fusion partner.

7. The use of a vector as claimed in claims 5 and 6 in the transformation or  
5 transfection of a prokaryotic or eukaryotic host.

8. A host cell suitable for the transformation of vector as claimed in claims 5 and  
6.

10 9. An antibody, an affibody, or a derivative thereof which binds to one or more of  
the proteins, polypeptides, peptides, fragments or derivatives thereof, as claimed in  
any one of claims 1 to 3.

15 10. An immunogenic composition comprising one or more of the proteins,  
polypeptides, peptides, fragments or derivatives thereof, or nucleic acid sequences as  
claimed in any one or more of claims 1-3 and claim 4.

11. An immunogenic composition as claimed in claim 10 which is a vaccine.

20 12. Use of an immunogenic composition as a claimed in claim 10 in the  
preparation of a medicament for the treatment or prophylaxis of *Group B*  
*Streptococcus* infection.

25 13. A method of detection of *Group B Streptococcus* which comprises the step of  
bringing into contact a sample to be tested with at least one antibody, affibody, or a  
derivative thereof, as described herein.

30 14. A method of detection of *Group B Streptococcus* which comprises the step of  
bringing into contact a sample to be tested with at least one protein, polypeptide,  
peptide, fragments or derivatives as described herein.

15. A method of detection of *Group B Streptococcus* which comprises the step of bringing into contact a sample to be tested with at least one nucleic acid molecule as described herein.

5

16. A kit for the detection of *Group B Streptococcus* comprising at least one antibody, affibody, or derivatives thereof as claimed in claim 9.

17. A kit for the detection of *Group B Streptococcus* comprising at least one *Group B Streptococcus* protein, polypeptide, peptide, fragment or derivative thereof as claimed in claims 1 to 3.

15

18. A kit for the detection of *Group B Streptococcus* comprising at least one nucleic acid molecule as claimed in claim 4.

20

19. A method of screening for DNA encoding bacterial cell envelope associated or surface antigens in gram positive bacteria comprising the steps of:

- combining a reporter vector including the nucleotide sequence encoding the mature form of the staphylcoccus nuclease gene and an upstream promoter region with DNA from a gram positive bacteria.
- transforming the resultant vector into *Lactococcus lactis* cells.
- assaying for the secretion of staphlycoccus nuclease protein in the transformed cells.

25

20. A method as claimed in claim 19 wherein the reporter vector is one of the pTREP1-*nuc* vectors shown in figure 4.

30

21. A method as claimed in claim 19 or claim 20 wherein the gram positive bacteria is *Group B Streptococcus*, *Streptococcus Pneumoniae*, *Staphylcoccus aureus* or *pathogenic group A streptococci*.

22. A vector as shown in figure 4 for use in screening for DNA encoding bacterial cell envelope associated or secreted antigens in gram positive bacteria.

5 23. A method of determining whether a protein, polypeptide, peptide, fragment or derivative thereof as claimed in claims 1 to 3 represents a potential anti-microbial target which comprises inactivating said protein and determining whether *Group B Streptococcus* is still viable.

ID-1

**FIG. 1**

Clone 4

ATGGAAAAAAATACTTGGAAAAAAATTACTTGTAGTACTGCTGCTCTTCAGTAGT  
 TGCAGGAGGAGCAATTGCTGCTACTCACTCTAACTCAGTTGATGCTGCTCAAAAAA  
 AACTATCAAACCTTGGTCCCAACAGATTCAAAAGCGTCTTATAAAGCAATTGTT  
 AAAAAAATTCGAGAAGGAAAACAAAGGCCTACTGTAAAAATGATTGAGTCTAATG  
 ACTCCAAAGCTCAAGAAAACGTAAAAAAAGACCCAAGCAAGGCAGCCGATGTATT  
 CTCACCTCCACATGACCAACTTGGTCAATTAGTAGAATCTGGTGTATCCAAGAAA  
 TTCCAGAGCAACTCAAAAGAAATTGCTAAAAACGACACTAAACAATCACTTAC  
 TGGTGCACAATATAAAGGGAAAACTTATGCATTCCATTGGTATTGAATCTCAAG  
 TTCTTATTATAATAAAACAAAGTTAACTGCTGACGACGTTAAATCATAACGAAACA  
 ATTACAAGCAAAGGGAAATTCGGTCAACAGCTTAAACAGCTAACTCATATGTAA  
 CAGGTCCCTTTCTGTAGGCACACTTATTGGTAAATCTGGTAAGATG  
 CTAAGGCACTAACTGGGTAATGAAGCAGGTGTTCTGCTCTTAAATGGATTGCA  
 GATCAAAGAAAAATGATGGTTGTCAACTTGACAGCTGAAAATACAATGTCTAA  
 ATTGGCGATGGTCTGTCATGCTTTGAAAGTGGACCATGGGATTACGACGCTG  
 CTAAAAAAGCTGTCGGTAAGATAAAATCGGTGTTACCCACAATGAAA  
 ATCGGTGACAAAGAAGTTCAACAAAAAGCATTCTGGCGTAAACTTATGCCGT  
 TAACCAAGCACCTGCTGGTCAAACACTAAACGAATCTCAGCTAGCTACAAACTCG  
 CTGCATATCTAACTAATGCTGAAAGTCAAAAAATTCAATTGAAAAACGTATATC  
 GTTCTGCTAACTCATCAATTCAATCTCTGATAGCGTCAAAAAGATGAACCTGC  
 AAAAGCAGTTATCGAAATGGTAGCTCAGATAAAATACACCGTTATGCCTAAG  
 TTGAGTCAAATGTCAACATTCTGGACAGAAAGTGCTGCTATTCTTAGCGATACTTA  
 CAGTGGTAAAATCAAATCTAGCGATTACCTAAACGCTAAACAAATTGATAAG  
 ACATCGCTAAAACAAAATAG

MEKNTWKLLVSTAALSVVAGGAIAATHNSNSVDAASKKTIKLWVPTDSKASYKAIVK  
 KFEKENKGTVKMIESNDSKAQENVKKDPSKAADVFSLPHDQLGQLVESGVIQEIPQ  
 YSKEIAKNDTKQSLTGAQYKGKTYAFPFGIESQVLYYNKTKLTADDVKSYETITSKGK  
 FGQLKAANSYVTGPLFLSVGDTLFGKSGEDAKGTNWGNEAGVSVLKWIADQKKND  
 GFVNLTAEANTSFKFGDGSVHAFESGPWDYDAAKKAVGEDKIGAVYPTMKIGDKEV  
 QQKAFLGVKLYAVNQAPAGSNTKRISASYKLAAYLTNAESQKIQFEKRHTVPANSSIQS  
 SDSVQKDELAKAVIEMGSSDKYTTVMPKLSQMSTFWTESAAILSDTYSGKIKSSDY  
 LKRLKQFDKDIAKTKZ

ID-2

Clone 5

ATGTCAAAACAAAAGTAACGGCAACTTGTGTTATCCACTTACTCTTATCGCT  
ATCATCACCTTAGTGACCTTAGCAGAAACTATTAATCCAGAAACAAGCCTGACAA  
TGGCAACAGCATCAACAGAAAGTTCTTCTGAAGCAGAGAAACAGGAAAAACACA  
ACCTACAGATTAGAAACTGCTTCACCTCAGCCAGGAAGTATCTCAACAGAA  
AAAACAGAGATTGGTAGCAGACAGAGACATCATCAAGCAATGAATCATCATCAAGTT  
CATCACATCAATCTTCTTCCAACGAAGATGCTAAAACATCTGATTCTGCTTCAACA  
GCATCTACTCCTAGCACTAAACTACAAACAGTAGTCAAGCAGACAGTAAGCCAG  
GTCAATCAACAAAGACTGAATTAAAACCTGAGCCTACCTTACCAATTAGTAGAGCCT  
AAAATAACTCCCGCTCCGTCTCAGATAGAAAGTGTTCAGACAAATCAGAATGCTTC  
TGTTCCTGCTTATCCTTGATGATAACTTATTATCAACACCGATTCAACAGTGA  
AGCAACGCCATTCTACGTAGAACACTGGTCTGGTCAGGATGCCTACTCTCACTATT  
TATTGTCACATCGTTACGGTATCAAAGCTGAACAATTAGATGGGTACTTAAAATCT  
TTAGGGATTCAATATGATTCTAATCGTATCAATGGTGCTAAGTTATTACAATGGGA  
AAAAGATAGTGGTTAGATGTCCGTGCTATTGTAGCTATTGCTGTCCTGAAAGTTC  
ATTGGGAACTCAAGGAGTGGCTAAAATGCCAGGTGCTAATATGTTGGTTATGGTG  
CCTTGATCATGACTCTAGCCATGCTAGTGCTTATAATGATGAAGAAGCAATTATG  
TTGTTGACAAAAAATACAATTATTAACAAACAAACTCTAGCTTGAAATCCAAGA  
TTTGAAAGCACAGAAATTATCTTCTGGACAACCTTAATACAGTTACTGAGGGTGGTG  
TTTATTATACAGATAACTCTGGAACTGGTAAACGTCGTGCCAGATTATGGAAGAT  
TTAGACCGCTGGATTGATCAACATGGAGGGACACCCAGAAATTCCGTGCCCTGAA  
AGCTTATCGACAGCAAGTTAGCAGATTACCAAGTGGTTAGCTTATCAACAG  
CGGTTAACACACAGCTAGCTATATTGCATCAACTTATCCATGGGGTGAATGTACATGG  
TATGTCTTAACCGCGCTAAAGAGTTAGGTTATACATTGATCCATTATGGGTAAT  
GGTGGAGATTGGCAACATAAGGCTGGCTTGAACAAACACATTACCAAAAGTAG  
GCTATGCTGTATCATTTCACCAAGGACAAGCTGGTGTGATGGCACTTACGGTCAC  
GTAGCTATTGTTGAAGAAGTTAAAAAGATGGTTAGCTTCACTTCAATTGAGAATCTAA  
TGCAATGGGACGTGGTATTGTCTTACCGTACTTTAGTTAGCAGCACAGCTGCAC  
AATTAACTTATGGTATTGGCCATAAAATAA

MSKQKVATLLLSTLVLSSPLVTLAETINPETS LT MATA STESS SEA KQE KT QPT DSE T A P S A E G S I S T E K T E I G T T E T S S N E S S S S S H Q S S N E D A K T S D S A S T A S T P S T N T T N S S Q A D S K P G Q S T K T E L K P E P T L P L V E P K I T P A P S Q I E S V Q T N Q N A S V P A L S F D D N L L S T P I S P V T A T P F Y V E H W S G Q D A Y S H Y L L S H R Y G I K A E Q L D G Y L K S L G I Q Y D S N R I N G A K L L Q W E K D S G L D V R A I V A I A V L E S S L G T Q G V A K M P G A N M F G Y G A F D H D S S H A S A Y N D E E A I M L L T K N T I I K N N N S S F E I Q D L K A Q K L S S G Q L N T V T E G G V Y Y T D N S G T G K R R A Q I M E D L D R W I D Q H G G T P E I P A A L K A L S T A S L A D L P S G F S L S T A V N T A S Y I A S T Y P W G E C T W Y V F N R A K E L G Y T F D P F M G N G G D W Q H K A G F E T T H S P K V G Y A V S F S P G Q A G A D G T Y G H V A I V E E V K K D G S V L I S E S N A M G R G I V S Y R T F S S A Q A A Q L T Y G I G H K Z

FIG. 1 CONT'D

ID-3

Clone 6

GTGCATATGTTACAAACATTGGACAAACAGGCATTCAAGCAACTCGAATTGCTT  
 AGGTTGTATGAGAATGAGTGACTGAAAGGAAACAAGCTGAAGAAGTAGTTGGA  
 ACAGCATTAGATTTGGGTATTATAAATAAAAGTGAAGAAAGTGTCTCTGGCGT  
 CAAAGTGAATAATCATGTGTTATCAAGAACAGAAATTGCTCTTTCAAGAGA  
 TTAATCAGATGACTTCGTGAAGAACATGCGGACCATGACTTATGATGTATGTT  
 GATCCTTAGTTCTTCTTTATAGGTGCCTCACGTATTAACATTGGCTATGGGA  
 GCTTTATGATTCAAAAGGTCAAGTTACTGTTGGTACTTGTAACATTGTGACG  
 TATTAGATATGTTGGTATGGCCCTTGATGGCGATTGGTTCTTGTCAATATGGTA  
 CAGCGTGGTAGTGTCTTATAACCGTATTAATAGTCACTTGAGCAAGAACCGGA  
 TATAACTGATCCTTAAATCCTATCAAACCTGTTGTCAATGGAACATTAAGATA  
 TGATATTGATTCTTAGATACGACAATGAGGAAACCTTAGCCGATATTCAATTAC  
 CTTAGAAAAAGGTCAAACCTTAGGTTGGTAGGTCAAACGGGATCAGGGAAAGACA  
 AGTCTTATTAAGTTATTGCTACGTGAACATGATGTGACTCAGGGAAAATTACTT  
 AAATAAACATGATATACGTGATTATCGATTGTCTGAGTTACGTCAACTAATCGGTT  
 ATGTTCTCAAGATCAGTTTATTGCTACCGTATTAGAAAATGTTGCTTTG  
 GAAATCCAACCTCTATCTATCAATGCTCAAAGAACGAACTAAATTGGCACATGTT  
 TACGATGACATTGAACAGATGCCAGCAGGATTGAGACTCTAATTGGAGAAAAAG  
 GAGTCTCATTATCTGGTGGACAAAAACAAAGGATTGCGATGAGTCGTGCCATGATT  
 TAGATCCAGATATTCTTATTGGATGATTCTCTATCAGCAGTGGACGCTAAAACG  
 GAACATGCTATTGTTGAGAATCTAAAACGAATCGTCAAGGGAAATGACTATT  
 TTTCAGCACATCGTTATCAGCTGTTGCACCGACACCTATCTTAGTTATGCGAG  
 ACGGCAGAGTCATTGAGCGAGGTCAACATCAAGAGTTGCTAAATAAGGTGGTTG  
 GTATGCTGAAACGTATGCCTCACAGCAATTAGAAATGGAGGAAGCATTGATGAA  
 GTCTAA

MHMLQNIGQTGIQATRIALGCMRMSDLKGKQAEVVGTLADLGIINNKVQESVSGVK  
 VTKSLCYQEQLASFQEINQMTFVKNMRTMTYDVMFDPLVLLFIGASYVLTLMGAF  
 MISKGQVTVGDLVTFTYLDMLVWPLMAIGFLNMVQRGSVSYNRINSLLEQESDITD  
 PLNPIKPVVNGLTRYDIDFFRYDNEETLADIHFTLEKGQTLGLVGQTGSGKTSLIKLLR  
 EHDVTQGKITLNKHDIRDYRLSELRQLIGYVPQDQFLFATSILENVRFGNPLTLSINAVKE  
 ATKLAHVYDDIEQMPAGFETLIGEKGVSLGGQKQRIAMSRAMLDPDILILDDSLSAV  
 DAKTEHAIVENLKNRQGKSTIISAHRLSAVVHADLILVMRDGRVIERGQHQELLNK  
 GWYAETYASQQUEMEEAFDEVZ

ID-4

Clone 6b

TTGATGAAGTCTAATCAATGGCAAGTCTTAAGAGATTAATCTCCTATTTACGCCCT  
 TATAAATGGTTACAGTATTAGCTATCTCTTATTGTTGACGACTGTTAAA

**FIG. 1** CONT'D

AATATTATTCTTTAATTGCTTCACATTTATTGATCACTATCTGACAAATGTTAAT  
CAAACAGCAGTCTTATTAGTGGGATATTATTCAATGTATGTCTGCAGACCTTA  
ATTCAATATTGGGAATCTCTTTGCGCGTGTCTTATAGTATTGTTAGAGAT  
ATTCTGAGAGATGCTTGCTAATATGGAAAGGCTAGGCATGTCTTATTGATAAG  
GACACCGGCAGGATCTATTGTGTACGTATTACTAATGATACTGAAGCAATATCTG  
ATATGTTTCGGGTATTATCAAGTTTATCTCGCGATATTATTACAGTTAC  
TCTGTACACTATGTTGATGCTAGACATTAAACTAACAGGACTCGTCGCTTTGTT  
ACCTGTTATCTTATATTAGTGAATGTCTATCGGAAAAAAATCAGTCACTGTCATTGC  
TAAAACGAGAAGTTACTTAGTGAATCAACAGTAAATTATCAGGAAGTATTGAAG  
GAATTCGCATTGTACAGGCTTGGTCAAGAAGAGCGCTGAAGACTGAATTGAG  
GAAATTAAACAAAGAGCATGTTGTATGCCAATCGTTCTATGGCTCTGATAGTCT  
CTTCTTAAGACCGGCATGTCCTTAAACTCCTAGCATATGCTGTTCTATGTC  
TTATTGTTGATTACAGGAGTTAAAGGAGGTCTACGGCAGGATTAAATGATGCTT  
TTATTCACTACGTTAATCGTCTATTGACCTTAAATTGAAGTAACGCAAAATT  
CAACCTTACAAACATCAATGGTATCAGCAGGGCGTGTGTTGATCTGATTGAT  
GAAACAGGTTGAACCAAGCCAAAAAAATACAGAAGCT

MKSQNQWQVFKRLISYLRPYKWFVLALSLLLLTTVVVKNIPLASHFIDHYLTNVNQTA  
VLILVGYYSMYVLQTLIQYFGNLFFARVSYSIVRDIRDAFANMERLGMSYFDRTPAG  
SIVSRITNDTEAISDMFSGILSSFISAIFIFTVTLYTMLMLDIKLTGLVALLPVIFILVNYY  
RKKSVTVIAKTRSSLSDINSKLSGSIEGIRJVQAFGQEERLKTEFEEINKEHVVYANRSM  
ALDSLFLRPAMSLLKLLAYAVLMSYFGFTGVKGGLTAGLMYAFIQYVNRLFDPLIEVT  
QNFSTLQTSMVSAGRVFDLIDETGFEPSQKNTA

ID-5

### Clone 7

ATGAAAAGAAAAGACTTATTGGTGATAAACAAACTCAATACACGAT  
TAGAAAAGTTAAGTGGAGTAGCTTCAGTGCAACAGGGGTATGTA  
TTTTCTTCATAGTCCACAGGTATTGCTGAAGAAGTAAGTGTTC  
CTGCAACTACAGCGATTGCAAAGTCGAATTAAATCAGGGTGACAAC  
CGGCAATCTACTAATTAAAAGATGACATAAAACTCAAACCTGAGAC  
GGTTGTGACACCCCTCAGATATGCCGGATACCAAGCAATTAGTATCAG  
ATGAAACTGACACTCAAAAGGAGTGACAGAGCCGGATAAGGCGAC  
AAGCCTGCTTGAAGAAAATAAGGCTCTGTTTAGATAAAAATACCT  
TAGATTAAAAGTGGCACCATCTACATTGCAAAATACTCCGACAAA  
ACTTCTCAAGCTATAGGTGCTCCAAGTCCGACCTGAAAGTTGCTAAT  
CAAGCTCCACAGATTGAAAATGGTTACTTAGGTTACATCTAAAGA  
ATTGCCTCAAGGTCATCCTGTAGAAAGCACTGGGCTTGGATATGGG  
GAGATGTTGATCAACCGTCTAGTAATTGGCCAATGGTGTATCCCT  
ATGACTAATGCTAAGAAAGATGATTACGGTTATTATGTTGATTAA  
ATTATCTGAAAACAACGAAAACAAATATCTTTTAATTAAATAACA  
AAGCAGGAACAAATTAAAGCGGCATCATATTCCATTATTACGA

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

CCTGAGATGAACCAAGTTGGATTGATGAAAAGTACGGTATACATAC  
TTATCAGCCCCCTCAAAGAAGGGTATGTCGTATTAACATATTGAGTTC  
ATCTGGTAACATATGACCACTTATCAGCATGGCTTTAAAGATGTTGC  
AACCCCCCTCAACAACTTGGCCAGATGGTAGTAATTTGTGAATCAAG  
GACTATATGGAAGGTATATTGATGTACCACTGAAAACAAATGCCAAA  
GAGATTGGTTTCTAATCTTAGATGAAAGTAAGACAGGAGATGCAGT  
GAAAGTTCAACCCAACGACTATGTTTAGAGATTAGCTAACCATAC  
ACCAAATTGTAAAAGATAAGGATCCAAAGGTTATAATAATCCT  
TATTACATTGATCAAGTGCAGCTAAAGGATGCTCAACAAACTGATT  
AACAAAGTATTCAAGCAAGTTTACAACCTAGATGGGTAGATAAAA  
CTGAAATTAAAAGAATTGAAAGTACAGATAAAAAATCAAATGCT  
ATACAAATTCTGATATCACTCTCGATACTAGTAAATCTTTAATA  
ATCAAAGGCAGCTTAATCCTAAACAAGGTCTTCAATATATCTTAT  
AATGGTAACAATGTCACGACAAGGCAATCTGGGAATTAAAGACCA  
ACTTTATGCTTATAGTGGAAATTAGGTGCAGTCTCAATCAAGATGG  
TTCAAAAGTTGAAGCCAGCCTCTGGTACCGAGTGCTGATAGTGTCA  
CTATGATTATTATGACAAAGATAATCAAAACAGGGTTAGCGACT  
ACCCCCCTGTGAAAAATAATAAGGTGTTGGCAGACGATACTTGA  
TACTAAATTAGGTATTAAGGACTATACTGGTTACTATTATCTTACGA  
AATAAAAAGAGGTAAAGGATAAGGTTAAGATTAGTACCTTATGCAA  
AGTCATTAGCAGAGTGGGATAGTAATACTGTTAATGACGATATAAAA  
ACGGCTAAAGCAGCTTTGTAAATCCAAGTCACCTGGACCTAAAAA  
TTAAGTTTGCTAAAATTGCTAATTAAAGGAAAACAAGATGCTGT  
TATATACGAAGCACATGTAAGAGACTTCACCTGATCAATCTTGG  
ACGGAAAATTAAAAATCAACTGGTACCTTGCAAGCCTTTCAGAG  
AAACTAGATTATTACAGAAATTAGGAGTTACACACATTAGCTTT  
ACCGTATTGAGTTATTGTTAATGAAATGGATAAGTCACGCTC  
AACAGCTTACACTCCTCAGACAATAATTACAATTGGGCTATGACC  
CACAGAGCTATTGCTCTCTGGAAATGATGTTAGAGAAAACCAAAA  
GATCCATCAGCACGTATGCCGAATTAAAACAATTACATGATAT  
TCATAAACGTGGCATGGGGTTATACITGATGTCGTCTATAATCACA  
CTGCAAAAACCTATCTCTTGAGGATATAGAACCTAATTATTACT  
TTATGAATGAAGATGGTTACCAAGAGAAAAGTTGGAGGGGACGT  
TTAGGAACCACTCATGCAATGAGTCGTGTTGGTTGATTCATT  
AAATATCTTACAAGTGAATTAAAGTTGATGGTTCCGTTTGATATG  
ATGGGAGATCATGATGCCGCTGCGATTGAATTAGCTTATAAAGAAGC  
TAAAGCTATTAACTCTAATATGATTATGATTGGTGAGGGCTGGAGAA  
CATTCCAAGGCATCAAGGTAAAGCCGGTAAACCAAGCTGACCAAGAT  
TGGATGAAGTCACCGATACAGTTGGCGTCTTCAGATGATATTGCT  
AATAGCTTGAATCTGGTTCCAAATGAAGGTACTCCAGCTTCTAC  
ACAGGTGGCCCACAATCTTACAAGGTATTTAAAAATATCAAAGC  
ACAACCTGGGAATTGAAAGCAGATTGCCAGGAGATGTGGTGCAGT  
ATATTGCTGCACATGATAACCTTACCTGATGATGTGATTGCAAAAT  
CAATTAAAGACCCCTAAGGTAGCTGAAGAAGATATTGATAGACGT

FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

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CTGCGTTAGGAAATGTAATGATTTAACATCTCAAGGGACAGCATT  
CATTCAATTCTGGTCAAGAGTATGGTCGTACGAAGCGTTACTAACCC  
TGATTACATGACAAAAGTTTCAGATGACAAATTGCCTAATAAAGCAA  
CACTTATTGAAGCTGTTAAGAACATACCCATATTTATTGATGATTCA  
ATGATTCTTCAGATGCCATTATCATTGATTGGGCAGCAGCCACAG  
ATAATAACAAACACCCAATTCAACGAAAACACAGGCCATACAGCA  
GGTTTAATCACATTAAGGCAGTCAACAGATGCTTCCGGAAATTGAG  
CAAAGCAGAAATTGATCGTGAGGTTAGCTGATTACAGAGGTAGGTC  
AAGGTGATATTAAAGAAAAAGATTGGTTATTGCTTACCAAACAATA  
GATTCTAAAGGCGATATTACGCAGTATTGTTAATGCTGATAGTAA  
AGCTAGAAACGTTTACTAGGTAAAAAATATAAACACCTTTAAAAG  
GGCAAGTAATTGTTGATGCTGATCAAGCGGGGATTAAACCAATCTCA  
ACTCCTAGAGGTGTTCATTTGAAAAAAGATAGTTGCTGATTGATCCA  
TTAACAGCAATTGTTGATTAAAGTTGGCAAAGTTGCTCTAGCCCTAA  
GGAGGAATTGCAAGCAGATTATCCAAAACACAATCTTCAGGGAT  
CTAAAACGGTAGAAAAAAGTAAATAGAATAGCTAATAAGACCTCAAT  
AACTCCTGTAGTTCTAATAAGACCGATTGATCTGACAAATGAAG  
CTAATTGCCAAAAACTGGAGATAAGTCATCAAAAATACTAAGTGT  
TAGGAATAAGCATTCTAGCAAGTCTACTGCTACTAGGTCTCTCT  
TTAAAGAGGAATCGCACTTAA

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MKRKDLFGDKQTQYTIRKLSVGVASVATGVCIFLHSPQVF  
AAEVSVSPA  
TTAIKSNTNQVDNRQSTNLKDDINSNSE  
TVVTPSDMPDTKQLVSD  
ETDT  
QKGVT  
EPDKATSL  
LEENKG  
PVSDK  
NTLD  
KVAP  
STLQ  
NTPD  
KTSQA  
IGA  
PSPL  
LKVA  
NQAP  
QIENG  
YFRL  
HLKEL  
PQGH  
PVEST  
GLWI  
WGD  
VDQ  
PSSN  
WPNG  
AIP  
MTNA  
KDDY  
GYY  
VDF  
KLSE  
KQR  
KQIS  
FLIN  
NKA  
GTN  
LNG  
DH  
HIPL  
LRPE  
MNQV  
WIDE  
KYGI  
HTYQ  
PLKE  
GYVR  
INYLS  
SSGN  
YDH  
LSA  
WL  
FKD  
VATP  
STTWP  
DGSNF  
VNQGL  
YGRY  
IDV  
PLK  
TNA  
KEIG  
FLIL  
DESK  
TGD  
AVK  
QPNDY  
VFR  
DLAN  
HNQ  
IFV  
KDK  
DPK  
VYNN  
PYY  
IDQ  
VQL  
KDA  
QQT  
DL  
TSIQ  
ASF  
TLD  
GV  
DK  
KTE  
IL  
KEL  
KV  
TD  
DN  
QN  
NA  
IQ  
IS  
DT  
LDT  
SK  
SLI  
KG  
DFNP  
KQGH  
FN  
ISY  
NG  
NN  
VTR  
QS  
WE  
FKD  
DQL  
Y  
AS  
GN  
LG  
AV  
LN  
QDG  
SKV  
EASL  
WSPS  
ADS  
VTM  
IIY  
DKD  
NQ  
NRR  
V  
ATT  
PLV  
KNN  
GV  
WQT  
TLD  
TKL  
GI  
KN  
YT  
GYY  
Y  
EIK  
RG  
KDK  
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K  
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LDP  
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AK  
SLA  
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VN  
PSQL  
GP  
KNL  
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VR  
DFT  
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SLDG  
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KN  
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PV  
LSY  
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EM  
DK  
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SS  
DNN  
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NW  
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QSY  
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FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

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VIA YQTIDSKGDIYAVFVNADSKARNVLLGEKYKHLKGQVIVDADQA  
 GIKPISTPRGVHFEKDSLIDPLTAIVIKVGKAPSPKEELQADYPKTQSFK  
 GSKTVEKVNRRIANKTSITPVVSNKTDSYLTNEANLPKTGDKSSKILSVVG  
 ISILASLLALLGLSLKRNRT\*

ID-6

Clone 9

ATGAAAAAAAGTTTTCTCATGGCTATGGTTGTGAGTTAGTAATGATAGCAGG  
 GTGTGATAAGTCAGCAAACCCAAACAGCCTACGCAAGGCATGTCAGTTGTAACC  
 AGCTTTACCCAATGTATGCGATGACAAAAGAAGTATCTGGAGACCTAAATGATGT  
 GAGGATGATCCAATCAGGTGCAGGCATTCACTCCTTGAACCGTCTGTAAATGATG  
 TGGCAGCTATTATGACGCGGATTGTTGTTACCAATCACATACCTTACAAGGCTT  
 GGGCAAGGGATCTAGACCCCTAATTAAAAAAATCAAAGGTTAATGTGTTGAAGC  
 GTCAAAACCTCTGACACTAGATAGACTAGAAGCTAGAAGATATGGAAGTCACA  
 CAAGGCATTGACCCCTGCGACACTTATGACCCACATACCTGGACGGATCCCGTTT  
 AGCTGGTGGAGGAAGCTGTTAATATCGCTAAAGAGCTAGGACATTGGATCCTAAAC  
 ACAAAAGACAGTTACACTAAAAAGGCTAAGGCTTCAAAAAAGAAGCAGAGCAACT  
 AACTGAAGAATACACTCAAAAATTAAAAAGGTGCGCTCAAAACATTGTGACG  
 CAACACACGGCATTCTTATCTGGCTAACGATTGGCTTGAACAACTTGGTAT  
 CTCGGGTATTCTCCAGAGCAAGAGCCCTCTCGCCAATTGAAAGAAATTCAAG  
 ACTTTGTTAAAGAATACAACGTCAAGACTATTGAGCAAGACAAACGTCAACCC  
 AAAATTGCTCATGCTATTGCGAAATCAACAGGAGCTAAAGTAAAGACATTAAGTC  
 CACTTGAAGCTGCTCCAAGCGGAAACAAGACATATCTAGAAAATCTAGAGCAAA  
 TTTGGAAGTGCTCTATCACAGTTGAAGTAA

MKKVFFLMAMVVSLVMIAGCDKSANPKQPTQGMSVVTFSYPMYAMTKEVSGDLND  
 VRMIQSGAGIHSFEPVNDVAIYDADLFVYQSHTLEAWARDLDPNLKSKVNVFEAS  
 KPLTLDRLVKGLEDMEVQGIDPATLYDPHTWTDPLVAGEEAVNIAKELGHLDPKHKD  
 SYTKKAKAFKKEAEQLTEEYTKQFKKVRSKTFVTQHTAFSYLAKRFLKQLGISGISPE  
 QEPSPRQLKEIQDFVKEYNVKTIFAEDNVNPKIAHAIAKSTGAKVKTLSPLEAAPSGNK  
 TYLENLRANLEVLYQQLK\*

ID-7

Clone 15

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TTGTTCAATAAAATAGGTTTAGAACTGGAAATCAGGAAAGCTTTG  
 GCTTTATATGGGAGTGCTAGGATCAACTATTATTTAGGATCAAGTCC  
 TGTATCTGCTATGGATAGTGTGGAAATCAAAGTCAAGGTAATGTTTT  
 AGAGCGTCGCCAACGTGATGCGGAAACAAAAGTCAGGGTAATGTT  
 TTAGAGCGTCGCCAACGTGATGCGGAAACAAAAGAGGCCAAGGCAATG  
 TTTAGAGCGTCGTCAACGCGATGTTGAGAATAAGAGCCAAGGCAAT

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**FIG. 1** CONT'D

**SUBSTITUTE SHEET (RULE 26)**

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GTTCAGAGCGTCGTCAACGTGATGCGGAAAACAAAAGTCAGGGCA  
 ATGTTCTAGAGCGCCGCCAACGTGATGCGGATAACAAAGAGCCAAGTA  
 GGTCAACTTATAGGGAAAAATCCACTTTTCAAAGCCAACGTATCT  
 AGAGAAAATAATCACTCTAGTCAAGGTGACTCTAACAAACAGTCATT  
 CTCTAAAAAAAGTATCTCAGGTTACTAATGTAGCTAATAGACCGATGT  
 TAACTAATAATTCTAGAACAAATTCACTGATAAATAATTACCTAAA  
 ACAGGTGGTGTCAAAATGTCACTTTAAACTGTAGGTTGGTTA  
 ATTTGTTAACAGTCGCTGCAGTTGAGACGCAATGAAAATTAA

MFNKIGFRTWKSGKLWLYMVGVLGISTIILGSSPVSAMDSVGNQSQGNVL  
 ERRQRDAENKSQGNVLERRQRDAENKSQGNVLERRQRDVENKSQGNV  
 LERRQRDAENKSQGNVLERRQRDADNKSQVGQLIGKNPLFSKPTVSREN  
 NHSSQGDSNKQSFSKKVSQVTNVANRPMLTNNNSRTISVINKLPKTGGDQ  
 NVIFKLVGFGLILLTSRCGLRRNEN\*

ID-8

Clone 17

ATGACAAAAAAACTTATTATTGCTATATTAGCACTATGCACTATCTTAACCACCTCT  
 CAAGCTTTAGCTAAAGAAAAATCACAAACTGTTACCAAAAAACAACTATTCT  
 GGTCTATATTAAAAAGAAAAAGAGACAAGCCGGATAATAAAAGCAAATCAG  
 CGAGACACTTAAAGTCCCTTAAAACCCAAAAAGTAGTTGTTGATATGGGAG  
 CTTTGGATACTATCACAGCTTAGGAGCTGAAAAATCTGTTATTGGTATCCCGAAG  
 GCTAAAAATGCTCTAAGTTATTGCCAATAACGTCAAATCTGTTATAAGCTAA  
 GAGATACCAAGACGTAGGAAGTCTCTCGAACCAAACCTTGAAGCTATTGCTCGTA  
 TGCAACCTGATGTGGTTCTAGGAGCACGTATGGCTTCTGTTGATAATATTGAA  
 AAATTAAAGGAGGCTGCACCTAAAGCAGCATTAGTATATGCTGGAGTCGACTCAA  
 AAAAGTATTGACAAAGGAGTTGCTGAGCGTGTACAATGTTAGGGAAAATCTTC  
 GACCAAAATAAAAGGCAAAACCTTAATAAAAGATATCGCACAGCTGTTCTTA  
 AATTGCAAAAATATTGAGAAAAAAGGTAACCTACAGCTCTATTGTAATGGC  
 AAACAGCGGTGAACCTTAACCTCAATCACCTCTGGTCGTTGGTTGGATTTC  
 TGTAGGGATTAAAGCAGTCATGAAAATGAAAACTAAGTTCACATGGTACTC  
 CCGTATCTTATGAATACATCGCTGAAAAAAATCCTAACTATCTCTTGTGTTAGATC  
 GTGGAGCGACTATTGGACAAGGAGCTTCATAAAAGAACTTTAATAACGATGTT  
 ATTAAAGCAACTGATGCTGTAAAAACAAACGTGTTCATGAGGTAGATGGAAAAG  
 ATTGGTATATCAATTCAAGGCGGAAGCCGAGTAACACTCCGTATGATTAAAGATGTA  
 CAGAACTTGTGATAATCGTTAA

MTKKLIALCTILTTSQAVLAKEKSQTVTIKNNYSVYIKKEKRDKPDN  
 KKQISETLKVPLKPKVVVFDMGALDTITALGAEKSVIGIPKAKNALSLL  
 PNNVKSVDKRYQDVGSLFEPNFEAIARMQPDVVFLGARMASVDNIE  
 KLKEAAPKAALVYAGVDSKKVFDKGVAERVTMLGKIFDQNKKAKTFN  
 KDIAQAVLKLQKTIIEKKKGKPTALFVMANSGELLTQSPSGRFGWIFSVGG

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FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

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FKAVNENEKLSSHGTPVSYEYIAEKNPNYLFVLDRGATIGQGASSKELFN  
NDVIKATDAVKNKRVHEVDGKDWYINSGSRVTLRMIKDVQNFVDNR  
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ID-9

Clone 18

GTGAAGAAAACATATGGTTATCGGCTCAGTTGCTGCTATTTACTAGCTACTCAT  
ATTGGAAGTTACCAGCTTGGTAAGCATCATATGGGCTAGCAACAAAGGACAATC  
AGATTGCCATATTGATGATAGCAAAGGTAAAGGTAAAAGCCCTAAAACAAACAA  
AACGATGGATCAAATCAGTGTGAAGAAGGCATCTCTGCTGAACAGATCGTAGTC  
AAAATTACTGACCAAGGTATGTTACCTCACACGGTACCATTATCATTAA  
GGGAAAGTTCCCTATGATGCGATTATTAGTGAAGAGTTGTTGATGACGGATCCTAA  
TTACCAATTAAACAATCAGACGTTATCAATGAAATCTTAGACGGTTACGTTATTAA  
AAGTCAATGGCAACTATTATGTTACCTCAAGCCAGGTAGTAAGCGCAAAACATT  
CGAACCAAACAACAAATTGCTGAGCAAGTAGCCAAGGAACTAAAGAAGCTAAA  
GAAAAAAGGTTAGCTCAAGTGGCCATCTCAGTAAAGAAGAAGTTGCGGCAGTC  
ATGAAGCAAAAGACAAGGACGCTATACTACAGACGATGGCTATATTAGTCC  
GACAGATATCATTGATGATTAGGAGATGCTTATTAGTACCTCATGTAATCACT  
ATCATTATATTCTAAAAAGATTGCTCCAAGTGAGCTAGCTGCTGCACAAGCC  
TACTGGAGTCAAAACAAGGTCGAGGTGCTAGACCGCTCTGATTACCGCCGACAC  
CAGCCCCAGGTCGTAGGAAAGCCCCAATTCTGATGTGACGCCAACCTGGACA  
AGGTCACTAGCCAGATAACGGGGTTATCATCCAGCGCTCCTAGGCCAAATGATG  
CGTCACAAAACAACACCAAAAGAGATGAGTTAAAGGAAAACCTTAAGGAAC  
TTAGATCATCTACACCGTCTTGATTGAAATACCGTCATGTGGAAGAAGATGGG  
TGATTGAAACCGACTCAAGTGATCAAATCAAACGCTTTGGGTATGTGGTGCCT  
CATGGAGATCATTATCATATTATCCCAAGAAGTCAGTTATCACCTCTTGAAATGGA  
ATTAGCAGATCGATACTTAGCCGGCCAAACTGATGACAACGACTCAGGTTCAGATC  
ACTCAAAACCATCAGATAAAGAAGTGACACATACCTTCTTGGTCATCGCATCAA  
GCTTACGGAAAAGGCTAGATGGTAAACCATAATGATACGAGTGATGCTTATGTT  
TAGTAAAGAATCCATTCTAGTGGATAAAATCAGGAGTTACAGCTAAACACGGA  
GATCATTCCACTATATAGGATTGGAGAACTTGAAACATATGAGTTGGATGAGGT  
CGCTAACTGGGTGAAAGCAAAAGGTCAAGCTGATGAGCTTGTGCTTGGATC  
AGGAACAAGGCAAAGAAAAACCACTCTTGACACTAAAAAAGTGAAGTCGCAAAGT  
AACAAAAGATGGTAAAGTGGCTATATTATGCCAAAAGATGGCAAGGACTATTC  
TATGCTCGTTATCAACTTGATTGACTCAGATTGCCATTGCCGAACAAGAACTAATG  
CTTAAAGATAAGAAGCATTACCGTTATGACATTGTTGATACAGGCATTGAGCCACG  
ACTTGCTGTAGATGTGTCAGTCTGCCGATGCATGCTGGTAATGCTACTTACGATA  
CTGGAAAGTTCGTTGTTATCCCACATATTGATCATATCCATGTCGTTCCGTATT  
GGTTGACGCGCAATCAGATTGCAACAAATCAAGTATGTGATGCAACACCCCGAAGT  
TCGTCCGGATGTATGGTCTAAGCCAGGGCATGAAGAGTCAGGTTGGTCATT  
ATGTTACGCCCTTGATAAACGTGCT

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FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

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GGTATGCCAAACTGGCAAATTATCCATTCTGCTGAAGAAGTTCAAAAAGCCCTAGC  
 AGAAGGTCGTTGCAGCACCAAGACGGCTATACTTCGATCCACGAGATGTTGG  
 CAAAAGAAACTTTGTATGGAAAGATGGCTCCTTAGCATCCCAAGAGCAGATGGC  
 AGTCATTGAGAACCATTAATAAAATCCGATCTATCCCAAGCTGAGTGGCAACAAGC  
 TCAAGAGTTATTGGCAAAGAAAAATGCTGGTATGCTACTGATAACGGATAACCT  
 GAAGAAAAGCAACAGGCAGATAAGAGCAATGAAAACCAACAGCCAAGTGAAGCC  
 AGTAAAGAAGAAAAAGAATCAGATGACTTATAGACAGTTACCAGACTATGGTC  
 TAGATAGAGCAACCCCTAGAAGATCATATCAATCAATTAGCACAAAAAGCTAATAT  
 CGATCCTAAGTATCTCATTTCACCAGAAGGTGTCCAATTATAATAAATG  
 GTGAATTGGTAACCTATGATATCAAGACACTCAACAAATAACCCCTAA

MKKTYGYIGSVAAILLATHIGSYQLGKHHMGLATKDNQIAYIDDSKGKVAPKTNKT  
 MDQISAEEGISAEQIVVKITDQGYVTSHGDHYHFYNGKVPYDAIISEELLMTDPNYHFK  
 QSDVINEILDGYVIKVNGNYYVYLKPGSKRKNIRTKQQIAEQVAKGTKEAKEKGLAQV  
 AHLSKEEVAAVNEAKRQGRYTTDDGYIFSPTDIIDDLGDAYLVPHGNHYHYIPKKDLS  
 PSELAAAQAYWSQKQGRGARPSDYPRTAPGRKAPIPDVTPNPGQGHQPDNGGYHP  
 APPRPNDASQNKHQRDEFKGKTFKELLDHLHRLDLKYRHEEDGLIFEPTQVIKSNAF  
 GYVVPHGDHYHIIPRSQLSLEMEADRYLAGQTDDNDGSDHSKPSDKEVHTFLGH  
 RIKAYGKLDGKPYDTSDAYVFSKESIHSVDKSGVTAKHGDHFHYIGFGELEQYELDE  
 VANWVKAKGQADELVAALDQEQQKEPLFDTKKVSRKVTKDGVGYIMPKDGDKY  
 FYARYQLDLTQIAFAEQELMLKDKKHRYDIVDTGIEPRLAVDVSSLPMHAGNATYD  
 TGSSFVIPHIDHIHVVPSLTRNQIATIKYVMQHPEVRPDVWSKPGHEESGSVIPNVT  
 LDKRAGMPNWQIIHSAAEVQKALAEGRFAAPDGYIFDPRDVLAKETFVWKDGFSIPR  
 ADGSSLRTINKSDLSQAEWQQAQUELLAKKNAGDATDTDKPEEKQQADKSNENQQPSE  
 ASKEEKESDDFIDSPLDYGLDRATLEDHINQLAQKANIDPKYLIFQPEGVQFYNKNGEL  
 VTYDIKTLQQINP\*

ID-10

Clone 22

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ATGATACGCCAGTTTAAGAGAACACTGATTGGTATATTATATATCATGATG  
 TTTGT CCTATT TTTATTAGTTCTATCTATATCATTACCAATGCCCTATTGTTA  
 ATTCCCTAGGTTAAATGTTATTGTTACTAGGAATTAGTATTGGCAATACAGTC  
 GTTACAGGAAAAAAATGTTACATCTCAAATATTAAATAGTAGTCAGGACCCCTCT  
 TTCGAACCTCAACCGAGTGATTACGCTTATTAAATATTACACAATTAGAAGCT  
 AGAGAACGCAAAAGTTCTGAAACAATTGAACAAACCAATCATGTTGCACTTA  
 TGATAAAAGATGTGGTCGCACCAATGAAAGTTCCATTGGCAGCTATTCTTAATG  
 GCCCAGACAAATCATCTGATCCTAAGGAAGTTGAACAAACAAATTATTGAAATTGCA  
 ACATTATCTGAAACGTTAGCATTTGAAATTAGACAATATCGTGACGATT  
 TCGTTTGAAAGCTGTTAGCCTAGAGAAGTAGTAGTAGAAATTATAAAATCGTATA  
 AGGTTATTGTCTATCCAAAAGCTTATCTATCATAATTGAAGGCGATAATATCTGG  
 AAAACAGACAAAAAGTGGTTAACCTTGCTCTTACAGGTGCTAGATAATGCCAT

FIG. 1 CONT'D

AAAATATTCTAATCCTGAGTCAAAGATAATAATAAGCATAGGAGAAGAGAGTATT  
AGAATACAAGACTACGGTATCGGCATACTCGAAGAGGGATATCCCTAGACTTTTGA  
AGATGGCTTACGGGTTACAACGGTCATGAGCACCAAAAGGCAACAGGCATGGGG  
TTATATATGACAAAAGAAGTCTTATCTAGTCTGAATTGTCCATTCTGGTGGATAGC  
AAAATTAATTATGGGACTGCTGTTCTATACATAAATAA

MIRQFLREHLIWYILYIMMFVLFFISFYLYHLPMPYLFNSLGLNVIVLLGISIWQYSRYRKKMLHLKYFNSSQDPSFELQPSDYAYFNIITQLEAREAQKVSETIEQTNHVALMIKMWSHQMKVPLAAISLMAQTNHLDPKEVEQQLLKLQHYLETLLAFLKFRQYRDDFRFEAVSLREVVVEIIKSYKVICLSKSLIIIEGDNIWKTDDKKWLTFAISQVLDNAIKYSNPESKIIISIGEESIRIQDYGIGILEEDIPRLFEDGFTGYNGHEHQKATGMGLYMTKEVLSSLNLSIVDSKINYGTAVSIHKZ

ID-11

Clone 23

ATGACTTATCAAAAAACAGTTGGCTGGTATTCTACATTAGACAAATT  
GAAACCACATTAAATCTCTCTGTCTATCATGAGAATCTCTCAATT  
AATCAAGATATTCTCAAGAATGGTTAGCTATGAAAGATAGGGTGG  
TGGAAATCAAATTCAAGGATGTAAAGCTCTTCCATGATCACTT  
AAAATAAAAAGCTTAATCATATTAAATTATGACCTATGCTCGT  
AGTACATCTCAGCTGATACTAGTTATATCTTGA  
ACTCTGACTTAGTTGTTACTACTA  
ATTTAGATAACCTCTTCAAATTCACTAGACAATGCATATTAG  
CTGAGTCAGTCCAGCTCTTTGGGCTTGGATATGGGTT  
CGTTGGCGACAAGAAAATATGACTATTAAATTAAATT  
TTGAGAATGCCAACGAAGGGGATCAAACAATTCTTAA  
ATCGCATGTTGAAAATCAG  
GTAATTATTAGATGATACCTACAATTCTAAATTGGTT  
ATCGATGGGCATAAATTATTGGTGTGATGGGAGCTGCT  
CACTACATTCTGGGAATCAAACCTTGGCAA  
GGTATGGTGGCACTATAATTACTTGA  
TATTGGTTAGACCACCCATT  
CTACAACCTCTGATTGTATACCATCT  
GTCTATTACATTGCATGCACCAACAGTTATGTCTGA

MTYQKTVVLAGDYSYRQIETTLKSLCVYHENLSIFNQDIPQEWFAMKDRVGQTG  
NQIQDVKLFHDHLSPKWENKLNHINYMTYARYFIPQYISADTVLYLDSLTVVTTNLD  
NLFQISLDNAYLAAPALFGLGYGFNAGVMVNNQRWRQENMTIKLIEKNQKEIENAN  
EGDQTILNRMFENQVIYLDDTYNFQIGFDMGAAIDGHKFIFDIPPLPKIHYISGIKPW  
QTLSNMRLREVWWHYNLLEWSSISSKKVFGLDHPIKTQNYRLNFLIATTSDCIPSISEL  
VTALPDCLFHIACTNSVV\*

ID-12

## FIG. 1 CONT'D

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Clone 27

GTGAAGAAAACATATTGTTATATCGGCTCAGTGCTGCTATTTACTAGCTACTCAT  
ATTGGAAGTTACCAAGCTTGGTAAGCATCATGGGCTAGCAACAAAGGACAATC  
AGATTGCCTATATTGATGATAGCAAAGGTAAGGTAAGGCTAAAGCCCTAAACAAACAA  
AACGATGGATCAAATCAGTGCTGAAGAAGGCATCTCTGCTGAACAGATCGTAGTC  
AAAATTACTGACCAAGGTTATGTTACCTCACACGGTACCATTATCATTAA  
GGGAAAGTTCTTATGATGCGATTATTAGTGAAGAGTTGTTGATGACGGATCTAA  
TTACCATTTAAACAATCAGACGTTATCAATGAAATCTTAGACGGTTACGTTATTA  
AAGTCAATGGCAACTATTATGTTACCTCAAGCCAGGTAGTAAGCGAAAAACATT  
CGAACCAAACAAACAAATTGCTGAGCAAGTAGCCAAAGGAACAAAGAAGCTAAA  
GAAAAAAGGTTAGCTCAAGTGGCCCCTCTCAGTAAGAAGAAGTTGCGGCAGTCA  
ATGAAGCAAAAAGACAAGGACGCTAACTACAGACGATGGCTATTTTACTCC  
GACAGATATCATTGATGATTAGGAGATGCTTATTAGTACCTCATGGTAATCACT  
ATCATTATATTCTAAAAAGATTGCTCTCAAGTGAAGCTAGCTGCTGCACAAGCC  
TAAGGACTCAAAACAAAGGTCGAGGTGCTAGACCGTCTGATTACCGCCCGACAC  
CAGCCCCAGGTCTAGGAAAGCCCCACTCCTGATGTGACGCCAACCCCTGGACAA  
GGTCATCAGCCAGATAACGGTGGTTATCATCCAGCGCCTCTAGGCCAAATGATGC  
GTCACAAAACAAACACCAAAGAGATGAGTTAAAGGAAAACCTTAAGGAACCT  
TTAGATCAACTACACCGCTTGATTGAAATACCGTCATGTGGAAGAAGATGGGTT  
GATTTTGAAACCGACTCAAGTGATCAAATCAAACGCTTTGGGTATGTGGTGCCTC  
ATGGAGATCATTATCATATTATCCCAAGAAGTCAGTTATCACCTCTGAAATGGAA  
TTAGCAGATCGATACTTAACCCGGCCAACTGA

MKKTYCYIGSVAAILLATHIGSYQLGKHHMGLATKDNLQIAYIDDSKGKVKA  
PKTNKT  
MDQISAEEGISAEQIVVKITDQGYVTSHGDHYHFYNGKVPYDAIISEELLMTDP  
NYHFK  
QSDVINEILDGYVIKVNGNYYVYLKPGSKRKNIRTKQQIAEQVAKGTKEAKEKGL  
AQV  
AHLSEEVAAVNEAKRQGRYTTDDGYIFSPTDIIDDLGDAYLVPHGNHYHYIPKK  
DLS  
PSELAAAQAYWSQKQGRGARPSDYRPTPAPGRRKAPLPDVTPNPGQGHQPDNGGYH  
APP  
PRNDASQNKHQRDEFKGKTFKELLDQLHRLDLKYRHVEEDGLIFEPTQVI  
SN  
GYVVPHGDHYHIIPRSQLS  
PLEMELADRYL  
TRPN\*

ID-13

Clone 28

ATGGTAAATGATATATTAGAAAGAATGTATAAGAGAAATATTCCAAAATCTTACCT  
TACATCCGTCCCATTAGTTATTCTAAAAAGGAAGAACACCTATTGTTAGTAT  
GA  
CTGGTGGTCAACAAATAGATGGAGTGAAATTCACACAGATATATGAGGACTAT  
ATGAAATTACTCAGTCAGGTAAGGATATCGCAGAGTTATATCAAAAATATTCTAA  
AGAAGAGTTGGCAAATCTAGGCATTAATATTATCAATCCAATGATATAGAAAGG  
ACTGAGGAAAGAACTTTGATGAAATTATCAGTTGGTTCCAACCCTATGCAAC  
AAGACCAATTCAAGAAAGGCACACTATTCAATTAGAGCCAACAAGATTTC  
ACTA

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FIG. 1 CONT'D

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GAGGATAAGAAAAGAATTGAAGAAGACTGCAGCTCAAGGACTAAGCGAAATCGAC  
 CTTATTGATTAGTTGACCTATATGATATTAAATTAGACAATACAAGCGTCAATCGC  
 CATATTGTGGGGTTATTGACTAATAACACCCAAGTAACATACTATTTCCAAGAAC  
 ATTAAATAAGGAGTTGCTGTCAATGGCTCACGCTTAGATAACGTACAACAGGCCT  
 TTATTAAATTATTAAAGTGAAGAGGAGATACGAAAATTGCTTTAA

MVNDILERMYKENIPKSYLTSVPLVISQKGRRTYSFSMTGGQQIDGVKFTQIYEDYMK  
 LLSQGKDIAELYQKYSKEELANLGINYQSNDIERTEERTFDEIISWVSNPYATRPIQERH  
 TIQLEPTRFSLEDKKRIEEAAAQGLSEIDLIDLVLDYDINLDNTSVNRHIVGLTNNTQV  
 TYYFQEQLNKELLSMAHALDNVQQAFIKLLSEEIRKFAL\*

ID-14

Clone 31

ATGAATAAAAGAAGAAAATTATCAAAATTGAATGTAAAAAACACATTAGCTT  
 ATGGAGCTATCACTTAGTAGGCCCTTTTCATGTATTTGGCTGTAACGGTCATCT  
 TAAAAAGTTACAAGTTACTACTGAATCTTGTCAAAAGCAGATAAAGTCGCGTA  
 GCCAAAAAAATCAAAATGACTAAGGCGACATCTAAATCAAAAGTAGAAGATGTAA  
 AACAGGCTCCAAAACCTCTCAGGCATCTAATGAAGCCCCAAATCAAGTTCTCAA  
 TCTACAGAAGCTAATTCTCAGCAACAAGTTACTGCGAGTGAAGAGGGCGGTGAG  
 AACAAAGCAGTTGTAACAGAAAATACCCCTGCTACCAGTCAGGCACAACAAACTTA  
 TGCTGTTACTGAGACAACTTACAAACCTGCTCAACACCCAGACAAGTGGCCAAGTAT  
 TGAGCAATGGAAATCTGCAGGGCGGTGGATCTGCTGCTGCAGCACAAATGGC  
 TGCTGCAACAGGAGTCCTCAGTCTACTTGGGAACATATTATTGCCGTGAATCAA  
 ATGGTAATCCTAATGTTGCTAATGCCCTCAGGGAGCTCAGGACTTTCCAAACGAT  
 GCCAGGTTGGGTTCAACAGCTACAGTTCAGGATCAAGTTAA

MNKRRKLSKLNVKQHLAYGAIHLVALFSCILAVTVIFKSSQVTTELSKADKVRVAK  
 KSKMTKATSKSKVEDVKQAPKPSQASNEAPKSSQSTEANSQQVTASEEEAVEQAV  
 VTENTPATSAQQTYAVTETTYKPAQHQTSGQVLSNGNTAGAVGSAAAAQMAAATG  
 VPQSTWEHIIARESNGNPVANASGASGLFQTMPWGSTATVQDQVNSAIKAYRAQG  
 LSAWGY\*

ID-15

Clone 32

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ATGATTGTTGGACACGGAATTGATTACAAGAGATAGAGGGCGATTACTAAAGCAT  
 ATGAGCGTAATCAACGTTTGCAAGACGCGTTTGACCGAACAGAATTGCTTCTT  
 TTTAAAGGAATTCCAATCCCAAGCGTCAGATGTCTTTAACAGGGCGATGGGC  
 AGCAAAAGAGGGCTTATAGCAAAGCACTGGAACAGGAATTGGAAAGTTAATT  
 CATGATATCGAAATTATCGGATGATAAAGGAGCGCCCTTGATTACAAAAGAAC

FIG. 1 CONT'D

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GTAAATGGAAAATCTTTGTTCAATATCTCATAGTGGTAATTATGCACAAGCTAG  
TGTTATTTGGAGGAAGAAAAATGA

MIVGHGIDLQEIEAITKAYERNQRFAERVLTEQELLFKGISNPKRQMSFLTGRWAAKE  
AYS KALGTGIGKVNFHDIEILSDDKGAPLITKEPPNGKSFVSIHSGNYAQASVILEEK\*

ID-16

Clone 35

ATGATTTGTCACAGTGGGACACATGAACACAGCAGTTCAACCGTCTTATTAAAGA  
AGTTGATAGATTAAAAGGGACAGGTGCTATTGATCAAGAAGTGTTCATTCAAACG  
GGTTACTCAGACTTCGAACCTCAGAATTGTCAGTGGTCAAAATTCTCTCATATGAT  
GATATGAACCTTACATGAAAGAAGCTGAGATTGTTATCACACATGGCGGCCAGC  
GACGTTATGTCAGTTATTCTTAGGGAAATTACAGTTGTTCTAGGAGAAA  
GCAGTTGGTGAACATATCAATGATCATCAAATACAATTAAAAAAATTGCC  
ACCTGTATCCCTGGCTGGATTGAAGATGTAGATGGACTTGCAGGAAGCGTTGAAA  
AGGAATATAGCTACAGAAAAATATCAGGGAAATAATGATATGTTTGTCAAAATT  
AGAAAAAAATTATAGGTGAAATATGA

MIFVTVGTHEQQFNRLIKEVDRLKGTGAIDQEVIQTGYSDFEPQNCQWSKFLSYDDM  
NSYMKEAEIVITHGGPATFMSVISLGKLPVVVPRRKQFGEHINDHQIQFLKKIAHYPL  
A WIEDVDGLAEALKRIATEKYQGNNDMFCHKLEKIIGEI\*

ID-17

Clone39

TTGGAAGACAAATTATTCAACAAACATTAGGCATTACTATTTAAACTTTATT  
GTTTATATGGTCTATTATTGTTACCGTTATCATAGCTTTATTGCGACTAAAGAG  
TTAGGTGTTAGCACTAGCCAAGCAGGATTAGCAACGGGGATTATATTGTAAGGGAC  
TTGATTGCTCGTCTTATATTGTAAGCAATTAGAAGTTCTAGGACGTAAGTTAGT  
TTTACGTGGAGGGCTATTTTACTTAACAACTTAGCTTATTATATGCC  
AACTATCGGAGTAATGTATTAGTCGTTCTAAATGGTTTGGTTATGGCGTCGT  
GTCAACAGCAACTAATCTATTGTAACAGCCTATATACCAAGCTGATAAAAGAGGTG  
AGGGGATTAACCTTACGGTCTATCAACAAAGTTAGCCGCAGCTATTGGCCTTTG  
TAGGAACATTATGCTAGACAACCTCATATTAACCTTAAATGGTTATTGTATTAT  
GTACTTTAAATCCAGAACAGTTAGCTAAATCAACATTGCTTGGACTATTGATAGTT  
ATTGAGAAAAAAAGCAATTATCACAATTATGCACTTGTGATGGGTATCTCCTAT  
GCTTCCGTGTTAGGTTCCAAAAATTATACAAACAGAAATTAAATTGATGACAGT  
AGGAGCTTATTCTTATTGTTATGCACCTGTCATCACTTAACCAGACCATCTAT  
GGGAAGATTAAATGGACGCTAAGGGAGATAAGTGGGTGCTTATCCAAGTTATCTGT  
TCTTAACCTTGGGACTTGCTTATTAGGGAGTGCTATGGGAAGTGTTACCTACCTTC

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FIG. 1 CONT'D

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TATCAGGTGCTTGATTGGTTGGTATGGCACCTTATGTCCTGGCCAAGCAG  
 CATCAATCAAAGGTGTTGAGGAACATCGTTCAATACAGCCATGTCAACTACATG  
 ATAGGTCTTGATTAGGGTTAGGTGCTGGACCTACATTTGGGACTTGTAAAGAT  
 GGTTTCTGGAGCTGGTGTGCAATCCTTAGAGAATTATTCTGGATAGCAGCGATT  
 ATTCCTGTTGTTGTGGTATTCTATATTCTTAAATCATCTAGACAAGTTGAAACT  
 AAAACTATA  
 TAA

MEDKLFNKHFIGITLNFIVYMVYLFVIIAFIAKELGVSTSQAGLATGIYIVGTLIARL  
 IFGKQLEVLGRKLVLRGGAIFYLLTLAYFYMP SIGVMYLVRFLNGFGYGVVSTATNTI  
 VTAYIPADKRGEVINFYGLSTSLAAIGPFVGTMFMLDNLHINFKMVILC SILIAJVVLG  
 AFVFPVKNITLNPEQLAKSKSWTIDSFIEKKAI FITIIAFLMGISYASV LGFQKL YTTEINL  
 MTVGAYFFIVYALVITLTRPSMGR LMDAKGDKWVLYPSYLF TLGL ALLGSAMGSVT  
 YLLSGALIGFGYGT FMSCGQAASIKGVEHRFNTAMSTYMI GLDLGLGAGPYILGLVK  
 DGFLGAGVQSFRELFWIAAIIPVVC GILYFLKSSRQVETKTIZ

ID-18

Clone 47

ATGAATAGTGAACCTAAAAGTCAGTCACGAAAGTAAAAAATAGCAAGCAATCAG  
 AAGTGAAGAAAGATAAAAAAATGACAAAAAAGAACATTAGCCTATCTCAAAG  
 AGCATGAGCAAGAAATCATAGATTATGAAAATTACATAACAAACCAATTGAGTC  
 CGTTCAATTGATTGGTCAAGTGTAAAAGTAGAACAAAGCGGGATGGAACTCCA  
 CAAGGGGGT GATTATAATCTTCACTGAGAGGAAAGTTAATCATCTACAAAATT  
 AAAATTAAAGTTGATTTTATTAGCTCATAAAAATGATATCCC AAATATCAAAT  
 CAATGGGAATGCTAAATAAGCCATATACATAAAAATGGTATTGGCACATTAT  
 GAATAG

MILGGCQMN SEP KSQSNEVKNSQSEVKDKKMTKKEQLAYLKEHEQEIIDYVKLHN  
 NQIESVQFDWSSVKVEQSGNGTPQGGDYNLSRGKFNHLQNSKLIVDFYLAHKNDIPN  
 IKSMGMLNKPYIHNGIWHIYEZ

ID-19

Clone 102

ATGAAAAAGATTGATTATCAAAGTTATTAAAATGATTGTTGTTATTGTTTTA  
 ATTAGTGTAGCAGCTAGTTTATTCCACGTTGCCAAGTCAGAGATGATAAAA  
 TCCTTATTCAAATGGTCAACGTAAGCCTGGAAACTCTTATATGCTTATGATAAAA  
 TCCTTGATAAGCTATTAAAGCAAAAATAGAAATGACAAACCAAAATATAAAGC  
 AAGTTGCTGGTATGTTCTGCTGCTAAGAAAACTCATAAGACAGTTGTTGTCGTT  
 ATGGTTTGCATAGCAAAGAGAATATGAAGGCATATGGTTGGCTGTTCATAAG  
 TTAGGATACAATGTTCTATGCCTGACAACATTGCACATGGT GAAAGTCATGGCA

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FIG. 1 CONT'D

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GTTGATAGGCTATGGCTGGAACGACCGCGAGAACATTATCAAATGGACAGAAAATG  
 ATAGTGGATAAGAACATCCATCAAGCCAAATTACTTATTGGTGTTCATGGGTGG  
 AGCAACAGTCATGATGGCTAGTGGTAAAAATTACCTAGTCAGGTTGTTAATAT  
 CATTGAAGATTGTGGTATTCTAGTGTGTTGGGATGAATTAAAATTTCAGGCTAAAG  
 AGATGTATGGTTACCAAGCCTCCACTCTTATATGAAGTTCAACAATTCTAAAA  
 TCAGAGCAGGTTTCGTATGGACAAGCAAGTAGTGTGCAACAATTGAAAAAGAA  
 TAATTACCAAGCCCTCTTATTCATGGTATAAGGATAATTGTTCCAACAAGTAT  
 GGTATGACAACATAAAGCTACAGCAGGTAAGAAAGAGCTTATATTGTAAGA  
 GGGGCAAAACATGCGAAATCTTGAAACAGAGCCAGAAAAATATGAGAAACGTA  
 TCTCTAGTTTGAAAAATATGAAAAATAA

MKKIRLSKFIKMIVVILFLISVAASFYFFHVAQVRDDKSFISNGQRKPGNSLYAYDKSFD  
 KLLKQKIEMTNQNIKQVAWYVPAAKKTHKTVVVHGFANSKENMKAYGWLHKLGYNV  
 LMPDNIAHGESHGQLIGYWNDRENIWKTEMIVDKNPSSQITLFGVSMGGATV  
 MMASGEKLPQVNNIEDCGYSSVWDELKFQAKEMYGLPAFPLLYEVSTISKIRAGFSY  
 GQASSVEQLKKNNLPALFIHGDKDNFVPTSMVYDNYKATAGKKELYIVKGAKHAKSF  
 ETEPEKYEKRISFLKKYEK\*

ID-20

Clone 120

TTGAGGAGTAATATGGTAAAGACAGCAGTTAATGGCGACATACAATGGCGAAA  
 AATTATATCTGAACAACTTGATTCAATTGCCAACAGACATTAAAACCAGATTAT  
 GTATTATTGAGGGATGATTGTTCAACGGATGAAACAGTCATGTCGTCATAACTA  
 TATCGAAAACATGAGTTAGAAGGCTGGAAAATTGTTAAAACGACAAAAACTTA  
 GGCTGGCGTTAAATTTCGTCAATTACTTATTGATGTGTTAGCCTATGAGGTTGAC  
 TATGTCCTTTAGTGTCAAGATGATATTGGTATCTGATAAAAACGAACGACA  
 GTTGCCATTATGTCAGATAACCCTCAAATTGAGGTTTGAGTGCAGACGTTGATA  
 TCAAAACGATGTCAGTACAGAACGCCAGTGTCCACATTCTAACCTTTCTTAGTG  
 ATAGAACATCAGTCAGTACCTAAAGTATATGATTATCAAACATTCCGCCCCGATGG  
 ACCATTGCTATGAAGAGAGATTGCGCAAGCTATCGCTTGA

MRSNMVKTAVLMATYNGEKFISEQLDSIRQQLKPDYVLLRDDCSTD  
 DETVNVVNNYIAKHELEGWKIVKNDKNLGWRNLFRQLLIDV  
 LAYEVDYVFFSDQDDIWYLDKNERQFAIMSDNPQIEVLSAD  
 VDIKTMSSTEASVPHFLTFS  
 SDRISQYPKVYDYQTFRPGWTI  
 AMKRDFAQAJAZ

ID-21

Clone 143

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ATGATTGAGATTGACGATTGTCAATTATTGAAAAAGGAAGTTACGTTATT  
 GAATTATATTAAATGCTGAGGGCGAGAGAGTAGTTATTATAATCATAGATTGTCC

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**FIG. 1** CONT'D

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GTAGTGTAGTCCTATTTATATCGTCTATTTATGATTTACTTGCACAAGAAGTAC  
 CTCACTTGCATGATTACATCTATAATGCAAGAGATGATCACTACGATACTTCCAAG  
 TTAAAGAATTAAAGGAGTCACCAACCATCCAGTCCTTGGCATTCTCTGAAAGGTG  
 GCACGATAGTCGCTTGACTTCTAAAAGCCTGCAGAATGTTACAATTAAACCGACC  
 TTGATGAAGAAGTGAATCGACCATCATTCAATTAAAGACAGTCGAAAAATCAGTC  
 AGAAATCCTTGGCTCACCTGATTAAACCTTTGATGAGCAAGAACTATATCGTAC  
 AACTCAATTCTTCTCAAGCATTAGACCAAGATTATCTTCTGGCAAAGGTAAT  
 TGGTGTGAGTATGATACTGTTAATTTCACTACGATAACGGTTAACAGCTTATTAT  
 AAAGATACTTGAGTAA

MIHEIHDCQFIEKGSYVYLNYINAEGERVVIIIDFVRSVSPILYRLFMILLAQEVPHLHD  
 YTYNARDHYDTWKFKELKESNHPVLLAFSERWHDSRLTSKSLAECLQLTDLDEEVKS  
 TIIQLRQFEKSVRNPLAHLIKPFDEQELYRTTQFSSQAFLDQIIFLAKVIGVEYDTVNFHY  
 DTVNKLIKILE\*

ID-22

Clone 1

ATGGTAAAAGTTCAAATTAGGGTATCCACGTCTGGTGAACAGCGCGAATGGAA  
 GCAAGCGATCGAAGCTTCTGGGCAGGGAATCTGAAACAAAAAGATTAGAAAAAA  
 CAACTAAAACAATTACGTATCAATCATTAAAGAAACAAAAGAGGCAGGTATTG  
 ACCTTATTCCAGTGGGGATTCTTCTGTTATGATCATGTTGGATTGTCATTCA  
 ATTCAATGTAATCCCAAAGCGTTCGATGAGTATGAGAGGAATTAGACCTTATT  
 TTGCTATTGCAAGAGGTGACAAAGATAATGTCGCATCATCTATGAAAAAGTGGTTT  
 AATACCAACTACCAACTACATAGTCCCAGAATGGAGGTTGAGACTAAACCTCACTT  
 GCAGAATAATTACTTACTGATCTTATCTAGAAGCTAGGGAAGTAGTTGGTGATA  
 AAGCAAAGCCGGTTATC

MEEIMVKVSNLGYPRLGEQREWQIAIEAFWAGNLEQKDLEKQLKQLRINHLKKQKE  
 AGIDLIPVGDFSCYDHVLDLSFQFNVIPKRFDEYERNLDLYFAIARGDKDNVASSMKK  
 WFNTNYHYIVPEWEVETKPHLQNNYLLDLYLEAREVVGDKAKPVI

ID-23

Clone 2

ATGGTGTACTTTATTGCTAATGGTAGCCAAGTCAAGTTGATGGTTACATGGCTG  
 TTTATAACGATACTGACAAAAATAAAATGTTACCAAGATATGGAGGAAGGAGAAAG  
 TTATCAAGTTAA

MVLLLLLMAKSSLMVTWLFITILTKIKCYQIWRKEKVIKL

ID-24

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FIG. 1 CONT'D

## Clone 14

ATGAACAAAAAAATTCCGGGATCGGCTGGCTTCGATTGCAGTACT  
 TAGTTAGCTGCATGTGGACATCGTGGTGCCTCTAAATCTGGTGGTAA  
 ATCAGATAGCTGAAGGTTGCAATGGTAACAGATACCGGTGGTGTG  
 ATGATAAATCATTAAACCAATCTGGTGGGAGGTATGCAAGCTTGG  
 GGCAGAAGAATGGCCTTAAAAAAGGAGCTGGTTTGACTATTTCCA  
 ATCGGCAAGTGAATCTGATTATGCAACTAACTAGATAACAGCTGTGT  
 CTAGTGGTTATAAATTGATTTCGGTATTGGATTTCCTCTCATGATG  
 CTATTGATAAAGCAGCAGACAATAACAAAGATGTTAATTACGTCATC  
 GTTGATGATGTTATTAAAGGGAAAGATAATGTTGCAAGTGTGTCTT  
 GCGGATAATGAATCAGCTTACTTAGCAGGTATTGCAGCCGCTAAAAC  
 TACCAAAACAAAAACAGTTGGCTTGTAGGTGGTATGGAATCTGAGG  
 TTATTACCGTTTGAAAAAAGGTTTGAAAGCAGGTGTCAAATCAGTTG  
 ATAAATCAATTAAAATTAAAGTTGACTATGCTGGTTATTGGTGTGAT  
 GCTGCTAAGGGTAAGACAATTGCAGCCGCACAATATGCTCTGGCGC  
 AGATATT

MNKKISGIGLASIAVLSAACGHRGASKSGGKSDSLKVAMVTDGGVD  
 DKSFNQSGWEGMQAWGKKNGLKKGAGFDYFQSASESDYATNLDTAVS  
 SGYKLIFGIGFSLHDAIDKAADNNKDVNYYVIVDDVIKGKDGVASVVFAD  
 NESAYLAGIAAAKTTKTKTVGFVGGMESEVITRFEKGFEAGVKSVDKSI  
 KIKVDYAGSFGDAAKGKTIAAAQYASGADI

ID-25

## Clone 20

ATGTTACATTCTAAAAAAATACATTCTTATCGCTTATTGCCGTTCTC  
 TCTTAGCAACATATACGAGTTACAACCAAATCATGTAGCGGCTGA  
 ACAATCACAAAAACATCAACTGTTCTTATGAGTCAAAAACTATTG  
 AACATAAGTTAAAAGTTGCAGATAAAGAAGCTGCTCCTCTACGCT  
 AAAATCGACCATATCCAACGACATATTGAAGTCAAAAAAGCAAAAG  
 ATTTAAAAGTTATTGAATTGTATTTAACAAAGATATCAACCAACTA  
 GAGAAGCAAAATAAACGTCTACTAACTAAATTCTATACTTCTATTGA  
 TAATCAAACATGGGATAGCACAAGTGAAGTCAAAAATTGATTGATA  
 AGACAACCTATCCACTAACGAAAAAGATAGATTAAAATTATTTT  
 GAACAAACGTGCTTACCTTGAGACAAGGTTGAACGACCGCTATCAAA  
 ATTTGATAACTCTATTGAAAACCAAAATAAGAACTAAAATTAA  
 CGTAAAAATAGAAAAAAATCTATCAAAAACATGGTATTACAAAAGA  
 GGTATTAAAACCTACTATGCTAAAAAACAGTACGAGCTGACTGA

FIG. 1 CONT'D

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MLHSKKIHSLSLIAVLSLATYTSQPNVAAEQSQKTSTVLMQSQKTIEHK  
 LKVADKEAAPLYAKIDHIQRHIEVKKAKDLVIELYINKDINQLEKQNK  
 RLLTKFYTSIDNQTWDSTSEVKKLIDKTTLSTNEKDRKLKYFEQRAYLET  
 RLNDRYQKFDNSIENQNKEKILTSKIEKIYQKHGKTVKEVLKTYYAKKTV  
 RAD\*

ID-26

Clone 25

Clone 25 (partial sequence)

CTGAATTCCAAAAACGCTACAATCAAACCTGGTATCCTACTTATGGTTTCTGAT  
 ACTTATGCATTGTTACTAAAGAGTTGCCAGACAGAATAAAATACCAAGAT  
 CTCTGATCTCAAAAGTTATCAACAACATGAAGGCAGGGGTTGATAGTTCATGGA  
 TGAATCGCGAGGGAGATGGATACACTGATTCGCTAAAACATACGGTTTGAATT  
 TCACATATTCACCCTATGCAAATTGGCTTAGTCTATGATGCGGTTGAAAGTAACAA  
 AATGCAATCTGTATTAGGCTACTCCACTGACGGTCGTATTCGAGCTATGATTAG  
 AAATTAAAGGGATGATAAAAAAATTCTTCCTCTATGAAGCCTCTATGGTTGTCA  
 ACAATTCTATCATCAAAAAAGATCCTAAACTAAAAAATTACTCCATCGACTCGAT  
 GGTAAAATCAATTAAAAACGATGCAAAACCTTAATTATATGGTAGATGATAAAACT  
 TTTAGAAGCTTGGCGTAATCATGGCATAGCTGTTCTGTGTGAAATTGTTATCCG  
 CTCACAATTCCACACACATACGAGCCGGAAGCATAA

LNSQKRYNQTWYPTYGFSDTYAFMVTKEFARQNKITKISDLKKLSTTMKAGVDSSWM  
 NREGDGYTDFAKTYGFEFSHIYPMQIGLVYDAVESNKMQSVLGYSTDGRISYYDLEILR  
 DDKKFFPPYEASMVVNNSIKKDPKLKKLHRLDGKINLTMQNLNYMVDDKLEAW  
 RNHGHSCFLCEIVRSQFHTTYEPEA\*

ID-29

Clone 37

ATGAAAAAATTACTTCCCTAACATGTCTAACATGATGTCTTATGT  
 TTAGTGGCATGTTACTAAGCAAGCAATGTCGTCTAACAGCAAGCAATGTC  
 GTCTAACAGCAAATTAAAGATAAGAATAGTAAAGAAAAGGTGATTACT  
 GTTGCAACTTACAGCAAACCTACATCTACCTTTAGATTGATTAAA  
 GATAATGTAAAAGAAAAAGGATATACTTAAAGGTTGTCTGGTCTC  
 TGACTATATTCAAGGCTAACATTGCTTAAAGGTTGTCTGGTCTC  
 CTAACCTTTACAACATGAATTTCATGAGTATCTTAATAAGGAAA

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FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

ATGATGGTCATCTAGTGTCAATTACACCAATTATCATTCAATTGGCTG  
GTTTTATGGTCAACATTGAAAAAATTGCCGAGCTAAAGACGGT  
GCTAAGGTAGCGATTCCGTCTGATCTGCCAATATGACTAGAGCTCT  
GCTATTATTGCAAGAAAAGAAACTTATCACCTTAAAGAATACGTCCA  
AAAAGACCAAGGCTATCGAAGATATTACTAACCTTAAAAAATTAA  
CGAATTGAACCTGTAGCATTACTAACCTCAATCAGGCCTATTTGAA  
TATGACCTTGTCTTAATTCCCTGGATATGTGACAAAAATCAATCTA  
GTTCCCTAAAAGGGATAGATTATTATGAGAAAAAAACCAGATATCCG  
TTTGCAGGTGCCTGGTAGCTCGTGAAGATAATAAAAATAGTGATA  
AAATAAAAGTACTTAAAGAAGTACTAACAGTAAAGAGATTCTGTCA  
CTATATCACTAAGGAGATTCCAAGTGAAGCAGACGTTGCGTTCTAG

MKKLLSLTCLIMMSLCLVACTKQAMSSKQAMSSKQIKDKNSKEKVITV  
ATYSKPTSTFLDLIKDNVKEKGYTLKVVMSDYIQANIALENKEHDANL  
LQHEFFMSIFNKENDGHLVSITPIYHSLAGFYQHLKNIAELKDGA  
PSDPA  
LNQAYF  
NKS  
DKIKVL  
KEV  
LT  
SKE  
IRHY  
IT  
KEIP  
SEAD  
VAF  
\*

ID-30

Clone 38

CTGTTGGCTAAGGAAACCACTATGTCCTGCTTGGTATCAAAATTCTGCAGAAGC  
CAAGGCTTATATTACAAGGTTATAATGTTGCTAAAATGAAGTTAGATGATTGGT  
TACAAAAGCCCAGTGAAAAACCATATTCAATTATCTTAGATTTAGATGAAACAGTT  
TTAGATAATAGCCCATATCAAGCAAAGAATTAAAGATGGCTCTAGTTCACGCC  
AGAGAGTTGGGATAAAATGGGTGCAAAAGAAATCAGCTAAGGCTGTTGCCGGGTGCC  
AAAGAATTTGAAGTATGCTAATGAAAAGGAAATAAAATTATTATGTCAGA  
TCGTACAGATGCTCAAGTTGATGCGACTAAAGAAAATTAGAGAAGGAAGGTATA  
CCTGTTCAAGGGAAAGACCACCTGCTTTCCCTAAAAAAGGAATGAAATCTAAAGA  
GAGTCGCCGTCAAGGCAGTTCAAAAGATACCAATTATTATGCTTTGGAGATA  
ATTTAGTTGATTTGCTGATTTCTAAATCATCTAGTACAGATAGAGAACAACTAC  
TAACTAAACTCAAAGTGAGTTGGTAGTAAATTATTGTTCCCAAATCCTATGT  
ACGGTTCTGGAAAGTGCTATTATCAAGGAAAACATCTGGATGTTCAAAAACAA  
TTGAAAGAACGACAAAAATGTTGCATTGTTGATTGATTAA

MAKLTVKDVLKGKKVLVRVDFNVPLKDGVTNDNRITAALPTIKYIEQGGRAILFSHLGRVKEEADKEGKSLAPVAADLAALKLGQDVVFPGVTRGAKLEEAINALEDGQVLLVENTRFEDVDGKKEKNDEELGKYWASLGDGIFVNDAFGTAHRAHASNVGISSNVEKAVAGFLLENEIAYIQEAVETPERPFVAILGGSKVSDKIGVIEENLLEKADKVLI  
GGGMTYTFYKAQGIEIGTLEKEDKLDVAKDSZ

ID-31

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

Clone 41

ATGGATAATAAAGGTATAACGCCAATGTGATTGATGCAATCGCTGAGGGTGCAA  
GCACAGGTGCACAAATGGCTTCTCAATTGGTGCTAGTTGATTGCCTTGTGGTT  
TAGTTCTTGATTAA

MDNKGNANVIDAIAEGASTGAQMAFSIGASLIAFVGLVSLI

ID-32

Clone 42

ATGAAAAAGAAAAACAAATCCTCTAACATTGCTATAATTGCAATCTT  
TTTGCTATTATGCTTGTCAATTCTTGTCAATTATTAGTT  
TTGGTTAGCCCTATTAAACCTACTTTGATGCATATCCAGTTATTAA  
TTGCATCTATAGCCTATGGACCTCGTATTGGTGCAACTCTAGGCGCCT  
TAATGGGGGGGATCAGCGTAGCTAACAGCAGCATTGTTCTATTACCA  
ACGAGTTACCTCTCACCTTGTGAAAATGGTAAATTATTACCG  
CTAATTATTGCACTGTACCACTGATTCTAACATGGGATTATTCTTAT  
TTCGTTTACAAATTACTACACAACCGCTTGGTTGGCTATCTCAGGT  
GCTATAGGCTCTAACAAACACAGTATTGTTTATCTGGAATT  
ATCTTTTCAAGTACTTATAATGGGAATATCAAGCTAACGCTCGCT  
GGGATTATTCTAACATTGAGATGGTATTGCAAGCTATC  
ATTGTATATCTAACGTACCTCGTATTCTCAATTAAACATTAA

MKKKNKSSNIAILIAFFAIMLVIHFLSSFISFWLVPIKPTLMHIPVILASIA  
YPRIGATLGALMGGISVANSSVLLPTSYLFSPFVENGNFYSLIIALVPRILI  
GIIPYFVYKLLHNRFGLAISGAIGSLNTVFLSGIFIFFSSTYNGNIKML  
AGIISNSLAEMVIAAIIVYLTDPRLNIKH\*

ID-33

Clone 43

TTGAATATGACATTACAAGACGAAATCAAAAAACGCCGTACTTTGCCATCATCTC  
TCACCCGGATGCTGGTAAGACGACTATTACTGAGCAATTATTATATTGGTGGTG  
AAATTAGAGAAGCAGGGACAGTAAAAGGGAAAAATCAGGTACTTTGCAAAGTC  
CGACTGGATGGATATTGAAAAGCAACGGGTATCTCTGTTACTTCATCTGTTATGC  
AATTGATTACCGGGTAAACGTGTTAA

MNMLQDEIKKRRTFAIISHPDAGKTTITEQLLYFGGEIREAGTVKGKSGTFAKSDW  
MDIEKQRGISVTSSVMQFDYAG

**FIG. 1** CONT'D

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KRV

ID-34

Clone 44

ATGGCAGATAAAAACAGAACATTAAACTTGTAGGTGCAGGATCTC  
TAGCACACAAGAAAAAATTGAAAAGCCTGCTCTTCGTTATGCAAG  
ATGCGTGGCGTCGCTTGAAAAAAAACAAATTAGCAGTAGTTCACTC  
TATTATTAGCTCTTACTTACTTTCTGTTAGCCTCAAATTATTG  
TAACTCAGAAGGATGCTAATGGGTTGATTGAAAAAGTAACGACA  
TATCGCAACTTACCACTAAATTGAGTTCAAACCTCCTTTGGAAT  
GGTAGCATTAAATCCATCA

MADKNRTFKLVGAGSSSTQEKIEKPALSFMQDAWRRLKKNKLA  
VVSLLALLLTFSLASNLFVTQKDANGFDSKKVTTYRNLP  
KLSSNLPFWNGSI  
NPS

ID-35

Clone 46

ATGAAAAGAAAACAGTTATAAAATTAGGAATTGCAACCTTACTAACGGTTATTTC  
GCTTACACACCAATAAACCTAGCTACAAATCATA  
CCACAGAAAATATTGTTACTG  
CTCAAGAGTATAAAACAAAGAGAATGGTACTTACCTTTAA  
MKRKQFIKLGIA  
LLTVISLYTPINLATNHTTENTVTAQEYKTKENILFLL

ID-36

Clone 50

ATGTTTATAATCCTTACTTTATTGACTAATTACAATTGCTGTATTTCTTAG  
CTAAGAAAAAATGGCAATTACCGACATTACTTCATTGGTTGCTATTATCTATA  
ACCAAGGGCTGTGGAACAGTTGATTAAT  
MFYNPLLFI  
VLTIAVFFLA  
KKWQLPTFT  
FIGLLFI  
YNQGLWEQLIN

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FIG. 1 CONT'D

ID-37

Clone 51/52

GTGGTGCAAATAATGAAAAAAACATATAAAAGTATCATACCAATAGT  
 TCTTATTGGTATGATACTAGGAGGCTGCAAATGAATAGTGAACATA  
 AAAGTCAGTATAATGAAACAAAAAGTAGCAAGCAATCAGAAGTGAA  
 GAAAGATAAAAAAAATGACAAAAAAAGAACAAATTAGCTTATCTCAA  
 GAGCATGAACAAGAAATAATTGATTTGTAAAATCTCAGAATAAAA  
 GATAGAATCTGTACAAATTGATTGGAATGATGTTGATGGAGTAAAG  
 GGGGAAATGGTACACCTCAAGGAGGGAGAGGGGATTTACTTTT  
 GGGGAGATTAATAATGATTCTGAATCAAGTTGGAGAGTTGATATTGA  
 TATAGAAAAAGGACGGCTAGACCTAAAAAATATGTATTTAGGACAA  
 CCTATACGAATTGGAGGTAAATTATTTGAGTAA

MVQIMKKHIKSIPIVLIGMILGGCQMNSEHKSQYNETKSSKQSEVKDK  
 KMTKKEQLAYLKEHEQEIIDFVKSQNKIESVQIDWNDVRWSKGNGT  
 PQGGGEGILLFGEINNDSESSWRVDIDIEKGRDLKNMYLGQPIRIGGKLF  
 E\*

ID-38

Clone 53

ATGGAATTTGGCTTATAATGCTTCACAGCAATCGGTGTTCTATT  
 CCGCACGGTAATCATTCCACTTTATTCACTATAAGGATATGCTCCA  
 TTAGAGTTAGAACACAAGGATGGTGGCAGAGCATAGAGGACATC  
 ATATTGATGCATTAGGGAAAAAGATTCTACAGAGAACCAAAGCA  
 TATTCTCATGAACCTAATAAGGAACCTCACACAGAGGAAGAACACC  
 ATGCAGTAACACCGAAAGACCAACGTAAGGCAAACCAAATAGCCA  
 GATTGTCTACAGTGTCAAGAAATTGAAGAGGGAAAAAGCTGGT  
 AAATACACAACATCTGATGGTTACATTGATGCTAAAGATATTAA  
 AAAAGATAACAGGTACAGGTTATGTCATTCCACATATGACACATGAGC  
 ATTGGGTACCAAAGAAAGATTATCAGAGTCGGAATTAAAAGCAGCT  
 CAAGAATTCTTCAGGAAAATCTGAAGCAAATCAAGACAAACCAA  
 AACAGGTAAAACAGCTCAAGAAATCTATGAGGCAATTGAACCAAAA  
 GCAATTGTTAAACCTGAAGATTATTATTGGAAATTGCACAAAGCGAC  
 AGACTATAAGAATGGTACATTGTAATTCTCTATAAAGATCATTACC  
 ATTATGTGGAATTAAAATGGTTGATGAAGAAAAAGATCTTAGCT  
 GATTCAAGATAAGACATATTCTTAGAAGACTATTAGCTACGGCTAA  
 ATATTACATGATGCACCCAGAAAAACGTCTAAAGTTGAAGGATGGG  
 GTAAAGATGCTGAAATTATAAGGAAAAGGACTCTAATAAAGCAGA  
 TAAACCAAGTCCTGCACCAACTGATAATAACATCAAATTCTA

FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

GTGACAAAAACTTAAGTGCTGCAGAAGTATTCAAACAGCAGCAAAACC  
AGAAAAAAATTGTACCGCTTGATAAAATTGCTGCTCACATGGCATATG  
CAGTTGGATTGAGATGATCAATTGATTGTTCCCTCATCATGATCATT  
ATCATATAATGTTCCATGGCATGGTTGACAAGGGTGGTTATGGAAA  
GCACCAAGGCTATACATTACAACAACCTCTCTCAACAATTAAATA  
CTACATGGAACATCCTAATGAATTACCAAAAGAAAAGGGTGGGA  
CACGACAGTGATCATACAAAGGCTCAAATAAGACAATAAGCCA  
AAAATTATGCTCCAGATGAAGAACCTGAAGATTCAAGGAAAGTAAC  
CACAACTATGGTTTATGATGTTAATAAAGGTTAGACGAAGAAGA  
ACCAAGAAAAACAAGAAGATGAATCAGAGCTAGATGAATATGAAC  
GGAATGGCACAAACGCTAAGAAATATGGTATGGATAGACAATCTT  
TGAAAAGCAACTCATCCAATTATCAAATAATAGTGTAAAGTTTG  
AAAGC

MEFLAYNAFTAIGVSIPHGNHFHFIHYKDMSPLEATRMVAEHRGHHI  
DALGKKDSTEKPKHISHEPNKEPHTEEEHHAVTPKDQRKGKPNSQIVYS  
AQEIEEAKKAGKYTTSDGYIFDAKDIKKDTGTGYVIPHMTHEHWVPKK  
DLSESELKAAQEFLSGKSEANQDKPKTGTAQEIYEAEIPEKAVKPEDLL  
FGIAQATDYKNGTFVIPHKDHYHYVELKWFDEEKDLLADSDKTYSL  
YLATAKYYMMHPEKRPKVEGWGKDAEIYKEKDSNKADKPSAPTDNK  
STSNSSDKNLSSAAEVFKQAKPEKIVPLDKIAAHMAYAVGFEDDQLIVPH  
HDHYHNVPMWFDFKGGLWKAPEGYTLQQLFSTIKYYMEHPNELPKEK  
GWGHDSDHNKGSNKNKAKNYAPDEEPEDSGVTHNYGFYDVNKGS  
DEEEPEKQEDESLDEYELGMAQNAKKYGMDRQSFEKQLIQLSNKYSV  
SFES

ID-39 (Same as ID-76)

Clone 56

ATGAGGAAACGTTTCCTGCTAAATTATTGTGTTACTTTATT  
TCTTTTCTTATTCTTTCCGCTTTAAGGCCAAGATTGTCAGGT  
TGTTATGCAAGTTCAAGGAGATCATTGGGACATTGTAACGCATT  
TGATTITCCGTATTACATCGCTTGATCTATTAAAGGTAAGAAAA  
TCAACTTACTTATAGGTTGATAAAGGCCGAATTITGCTAACAGTAAAGCCTACAC  
TGAGGATTGGAGTGATAAAGGCCGAATTITGTTGCTCGTTAATAC  
TCAAAACCATACATTGGAAGGATTGCAACAATTGCCTCAAACATTAT  
TAAAAAAATCATGGATACTATGCCATTAGGATGAAGGATATTGATTG  
ATTACTTCAGTAGAAGGGGACTCAAACACTCAATTATCCAGAATTTC  
ACTACAGGCGACTGGCAATTAGAACGGCTTTCGATGAGGAGACAAG  
CGATGTGGTAAAGGGTATTAAATCAGGATGGTAAGGATGAGTATG  
TGATCATCCAAGGTTTCACTGGAGATCGTTACGTATCTCACTGAAG  
ATTCGGTCGAGAATTATTCCATTATCCTGAAAAAACCCATTGGTC  
ACGCTATTGGAGTGGTCGTTACTTAATCAGACTGTTCGTATTG

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

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GGTGGCGATCAGAAAAAGCAGAATTAAAGGCTTTCACTTGTAGAT  
 GGGCACTTGGTTCAGAATTAGATAGATGCAAAAGCAGCTCTAGTAA  
 TGTCTTAGCTTTGAAAAAGATGGAAAAGCTTATCTTTCTCAGCCAA  
 TAACGGACGTGGCGAAGTTGCTTTATCAATTAGTAAAATAA

MRKRFSLNFIIVTFIFFFILFPLFKAKDCQVYASFQGDHWDICNAFDF  
 PYLHRFDLIGKENQLYFIGCTIANSKAYTEDWSDKGRIFVARFNTQNHT  
 LEGLQQLPQTLLKNHGYYAIQDEGYSLITSVEGVLKLTYPFSTTGDWQ  
 LERLFDEETSDVVKVDINQDGKDEYVIIQGFHGDRRLRIFTEDFGRELFHY  
 PEKTPFGHAIWSGRLLNQTCFVFGWRSEKAELRLHFVDGHLVSELVDA  
 KAASSNVLAFAEKDGKAYLFSANNGRGEVALYQLVK\*  
 ID-40

Clone 57

ATGAAGCACAGTTAAAAGCTTTACGCTTGCTTACTCTCAATATTCTTGTGTTGGTGGAAAGGTAGTCAGCAGAGACTGTGAATATTGTTCTGATACAGCATACTGCTCCATTGAAATTAAAGATTCTGATCAAACCTTAAAGGAATCGATGTTGACATCGTTAACGAAGTCGCTAACGCGTGGCTGGAAATGTTAACATGACGTATCCAGGTTGATGCCGAGTTAACGCTGTTCAATCTGGACAGGCAGATGCGCTAACGGCCGGAACTACTGTTACTGAAGCACGTTAAAAGTCTTAATTCTCAGATACTTATTACGATACTCCGTTATTCTTACTAAAAATAATAATAAAAGTCACAAACTAACACAACATAAAAGGAAAAGTAGTCGGTGTAAAAATGGAACAGCTGCTCAAAGCTTCTAGAAGAAAATACTAAATACGGCTATAAGTTAAAACATTGATACAAGCGACCTAACATGAATAACAGCCTGATTCTGGTTCTATTACGCCGCTATGGACGATCAACCAGTTGTGCAATTGCGATAATCAAGGAAAAGCTACGCCATTAAACATGGAAAGGCGAACAGAGTTGGAGCTTGCATTTGCTGTCAAAAAAGGTAGTGGACACGATAATCTAATTAAAGAATTAAACACAGCTTGCACAAATGAAATCAGATGGCACTTATAATGACATCATGGATAAATGGCTTGGAAAAGACGCTACAAAAACAAGCGGCAAAGCAACAGGTAAATGCCAATGAAAAGCAACTCCTGTTAAAGCCAAGTTATAAAATTGTTCTGATTCTCATTGCAACCATTGCAATATCAAACGGTAAAGGGAAATATACTGGTTTGATATGGAATTATCACGAAAATTGCTAACACAGCAAGGTTAAACTTGATATCTCAATTCCAGGTTTGATGCCGCTTAAATGCTGTCCAATCTGGCAAGCTGACGGTGTATTGCAGGAGCCACAATCACAGAACGACGCCAAAAATCTTGATTCTGATCCTTATTACACATCTAGCGTTATCTAGCGGTTAAAAAAGGAAGCAATGTCAAATCATACCAAGATTAAAAGGAAAAACAATTGGTGCTAAAATGGTACTGCCTCATATACTTGGTTATCAGACCCACGAGATAAGTACAACATCATGTTAAAGCATTGATGAAGCATTCTACAAATGTATGATAGTATGAACTCAGGTTCAATTGATGCTCTAATGGATGACGAAGCCGTTCTGCTTACGCTTAAATCAAGGTCGAAATTGAAACACCTATCAAAGGTAAAAATCAGGCGATATCGGATTGCACTGAA

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FIG. 1 CONT'D

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AAAAGGGGCAAATCCAGAATTAAATTAAAATGTTAACAAACGGTCTTG  
 CTTCACTCAAAAAATCGGGTGAATCGATAAACTGTTAAAAAATAC  
 CTTCCACAGCCAGCACTCTTCAAACGATAAAGCTGCTAACCTGT  
 AGATGAATCAACTATTTAGGGTTAATTCTAATAACTACAAACAATT  
 GCTATCTGGTATTGGAACACTTTAAGTTAACCTTATCTCGTTGC  
 GATTGCTATGGTTATTGGTATTATCTTGGTATGATGAGCGTATCACC  
 AAGTAATACTCTCCGCACAATTCAATGATTTGTTGATATTGTCG  
 TGGTATTCCACTCATGATTGTGGCCGCTTTATTTCTGGGTATTCCCT  
 AATTAAATCGAAAGCATCACAGGTACCCAAAGTCCAATTAAATGACTT  
 CGTTGCTGCTACTATCGCTCTTAAATGGTGGTGCACATTGC  
 TGAAATTGTACGTGGTGGTATTGAAGCTGTTCTTCTGGTCAAATGGA  
 AGCAAGTCGCAGCTTAGGTATTCTACGGCAAAACTATGCAAAAGG  
 TTATCTTACCTCAAGCAGTACGCCTATGTTACCAAACATTATCAACC  
 AATTGTCATCTCATTAAAGGATAACAACAATTGTATCAGCAATCGGA  
 CTTGTGGAACCTTCCAAACTGGTAAATCATAA

MKHKLKAFTLALLSIFFVFGGKVSAETVNIVSDTAYAPFEFKDSQTYK  
 GIDVDIVNEVAKRAGWNVNMTYPGFDAAVNAVQSGQADALMAGTTV  
 TEARKKVFNFSDTYYDTSVILYTKNNNKVTNYKQLKGKVGVVKNGTA  
 AQSFLEENKSKYGYKVKTFDSDLMNNSLDSGSIYAAADDQPVVQFAI  
 NQGKAYAINMEGEAVGSFAFAVKKGSGHDLIKEFNTAFAQMKSQDGTY  
 NDIMDKWLKDATKTSKATGNANEKATPVKPSYKIVSDSSFAPFEYQ  
 NGKGKYTGFDMELITKIAKQQGFKLDISNPGFDAALNAVQSGQADGVIA  
 GATITEARQKIFDFSDPYYTSSVILAVKKGSNVKSYQDLKGKTVGAKNG  
 TASYTWLSDHADKYNHVKAFDEASTMYDSMNSGSIDALMDDEAVLA  
 YAINQGRKFETPIKGEKSGDIGFAVKKGANPELIKMFNNGLASLKKSGEY  
 DKLVKKYLSTASTSSNDKAAKPVDESTILGLISNNYKQLSGIGTTLSTL  
 ISFALAMVIGIIFGMMVSPSNTLRTISMIFVDIVRGIPLMIVAAFIFWGIPN  
 LIESITGHQSPINDFVAATIALSLNGGAYIAEIVRGGIEAVPSGQMEASRSL  
 GISYGKTMQKVILPQAVRLMLPNFINQFVISLKDTTIVSAIGLVELFQTGK  
 S\*

ID-41

Clone 58

TTGGAAGGTTACTTATTGCATTGATTCCATGTTGCGTGGGGAAAGTATTGGATT  
 GTTAGTAATAAAATTGGAGGGCGTCAAATCAACAAACATTGGAATGACTTTAGG  
 AGCATTGCTATTGCGATTATCGTATGTTATTAA

MEGLLIALIPMFAWGSIGFVSNKIGGRPNQQTGGMTLGALLFAIIVCLF

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**FIG. 1** CONT'D

ID-42

Clone 70

ATGAATACTATTATAATACATTGAGAACAGATAAAGGTTATAAAGT  
 TTATGAGGGGTATTTATGAAATTACTGGTGAAGAATGTGAAGAAG  
 CCTAGACCTTGTGATTCTAAGAATATTGTATTGCAGATAACAGATA  
 CTTGTGGCTACACTTTTACTCAATGAAGATGGAACAGTTATGATG  
 ATGTGACTTCTACAAATTGATGATAAATATTGGTTGGCTAGTCATA  
 AAGCTTGGATTCTTATTAGACAACATCAATTGACTATACCGTAA  
 CAGATATTCTGACGAGTATAAAATGCTGCAAATTGAAGGAAGATAT  
 TCGGGAGAAATTGCTCAGTCATTATGAATATGATATTCAACACTT  
 AATTTCGTACTCTCGCATAGAGATGGACTTCATCAAAGGTGAGGA  
 AAGGTTATCTTGGCGTAGATTGGTTTCTGGAGAATTGGCTATCA  
 ATTTCCTACCATCTCTATTGCTACTTTGTTGGATGTCTGT  
 GAAGGTATAGCAGAGTGTGGGGATGAACTTGATAGATATTAAAGGTT  
 TGAAGTGGGACAACCCATTACTGATATTATCAACAAAGAAGAATATT  
 CTTATATGAAATAGGTATTCTGGAATCTAGATTCAAAAGGAA  
 GAATTAGAGGTCGCGATAGCTGTTAGAGCACATCAGATCAGCAC  
 AGTAAAAGTGTGGATTCTCAACGAAGGAAAACCTCGCTTCAGGAA  
 CACCAGTGTCTTGTGATGACCAAATTGTTGGAAAGATTGGATAG  
 CAGACGAGAACACTCTCGAAAATTACCTAGGTTGATGATTGTT  
 AACCAAACATATGCTCATTAGGAGTTACTTTGTAACAGAAGATGG  
 CCAAATTGAAAACACAATCAAGCCATTATTGTATCCCAGAAAGTT  
 GGAACAAAGAATGA

MNTIYNTLRTDKGYKVYEGLYEITGEECEEALDLVIPKNIVADTDTCG  
 YTFLLNEDGTVYDDVTFYKFDDKYWLASHKALDSYLDNINFDTVTDIS  
 DEYKMLQIEGRYSGEIAQSFYEYDISTLNFRTRIEMDFIKGEERLSWRRF  
 GFSGEFGYQFFLPSSIFATFVSDVCEGIAECGDELDRLRFEVGQPITDIY  
 QQEEYSLYEIGYSWNLDFTKEEFRGRDSLLEHIRSATVKSVGFSTKEKLA  
 SGTPVLFDDQIVGKIFWIADEKHSENLYLGLMIVNQTYAHSGVTFVTED  
 GQILKTQSSPYCIPESWNKE\*

ID-43

Clone 78/94

ATGGAGTTAGTAATTAGAGATATCGTAAGCGGTTCAAGGAAACAGA  
 GGTCTTGAGAGGGAGCAAGTTACCGATTATTCAAGTAAATAACAG  
 GGGTCTTAGGTAGGAATGGTGTGGAAAACAACTTATTAAATA  
 CTTATGGGGATCTGAGCTGACAACGGGACCATTGTTATTGAAG  
 GATAATCACGAGTATCCTTACCGATAAGGATATTGGTATTGTTAT

**FIG. 1** CONT'D

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TCCGAAAACACCTTCCAGAATTAAACAGGGTATGAATTGTAAA  
 ATTTACATGGATTACATCCTCAGATGATTAATGACAATAGATGA  
 TTATTAGATTTATGAAATAGGACAAACAGAGCGTCATAGAATT  
 TCAAAGGATATTCTGATGGAATGAAGAGTAAGCTCTCATTAAATTGC  
 CTGATGATTCTAACGAAAAGTAATTACTAGATGAGCCACTGAC  
 TGCAAGTTGATGTTGATCAAGTATTGCAATAAAACGCCTTTGTTGGA  
 ATTAAGTGAGGATCATATTATTATATTATCAACTCATATAATGGCCTT  
 AGCAGAAGATCTATGTGATATTGTTGCTGATTAGACAAAGGAAAAC  
 TCCAAACATTAGATATTGATCGTAAACATGAACAAATCGAAGAGCGT  
 CTTCTCAAGTGTGAAGGGAGATGAATATGACAAGTAA

MELVIRDIRKRFQETEVLRGASYRFYSGKITGVLRNGAGKTLFNILYGD  
 DLAADNGTICLLKDNEHEYPLTDKDIGIVYSENYLPEFLTGYEFVKFYMD  
 LHPSDLMLTIDDYLDFMEIGQTERHRIKGSMDGMSKLSLICLMSKPK  
 VILLDEPLTAVDVVSSIAKRLLELSEDHIIILSTHIMALAEDLCDIVAVL  
 DKGKLQTLIDRKHEQFEERLLQVLKGDEYDK\*

ID-44

Clone 80

TTGTTTATGAGATACAAATGAAATTGAAAGCCTTGCAAGACCT  
 CGAAAACCTGAAGGTGTGGATAAAAAATCCGCTTATATTGTTGGTTC  
 TGGTTAGCAGGATTAGCTGCCGCTGTCTTTAATACGTGACGGTCA  
 AATGGATGGTCAACGTATTCAATTGAAAGAACTACCTCTTCTGG  
 AGGATCACTTGACGGTGTCAAACGACCTGATATCGGTTTGTAAACGC  
 GTGGTGGTCGTGAAATGAAAATCACTTCGAATGTATGTGGGATATG  
 TACCGTTCCATCCCCCTCTCGAAGTCCAGATGCTCTTATCTAGAT  
 GAATTATTGGCTTGACAAGGATGATCCAATTCACTCTAATGTCGC  
 CTCATTCAAACAGGGGAATCGCTTAGAATCTGATGGTATTTCAC  
 ACTCGGAACACATTCAAAGAGTTAGTTAAGCTAGTCATGGAGACTG  
 AAGAGTCTTAGGTGCTAAGACGATTGAAGAAGTTTCAAAAGAA  
 TTTTGAAAGTAATTGGACTTATTGGCTACTATGTTGCCTTG  
 AGAAATGGCATTAGCGATTGAAATCGCTCGATATGCTATGCGCTT  
 ATCCATCATATTGGTGGTCTGCCTGATTCACTTCATTAAAATTAAAT  
 AAATATAATCAATATGATTCTATGGTAAACCAATCATCAGTTATT  
 GAGTCTCATATGAGATGTTCAATTGATAGCAAGGTAACATAAT  
 CTCCGTAGACTTT

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MFMRYTNGNFEAFARPRKPEGVDKSAYIVGSGLAGLAAVFLIRDGQ  
 MDGQRIHIFEELPLSGGSLDGVKRPDIGFVTRGGREMEMHFECMWDMY  
 RSIPSLEVPDASYLDEFYWLKDPPNSNCRLIHKQGNRLESMDGDFTLGT  
 HSKELVKLVMETEESLGAKTIEEVFSKEFESNFWTYWATMFAFEKWH

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FIG. 1 CONT'D

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AIEMRRYAMRFIHHIGGLPDFSLKFNQYNQYDSMVKPIISYLESHNVDV  
QFDISKVTNISVDF

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ID-45

Clone 81

TTGTTGGCTTCTTATTATCGTCCGTTGTCAAAATCGCTTCGCTAA  
GGAGGAGCAATATGAAAAAAATTACTTAGATGGCTCCTCCTGTACTT  
TTCATTATTATCCTTATAGGAATGACTATCTTAGGTAAAGTCCTATATC  
AATAAAGTAACAGCTCACAAATAAAACTCTATAACTCTCGAATGAC  
TCCTACTATTTAATTCAAGGATCCAGTGCTACTCAAGAACGATTAA  
CAGCATGTTAGCACAGCTCAACCAATGGGAGAAAAACATAGCGTT  
TAAAGTTAACTGTCAAAAAAGACAATAGCATTATCTACAATGGACAA  
ATTAGCGGCAATGACCACAAACCCCTACATTGTCAATTGGATTGAAAA  
TAATGAAGATGGTTATAGTAACATCAAAAAACAAACAAATGGCTA  
CAGATTGCTATGAATGATCTTCAGAAGAAATATAAATTAAACGTTT  
TAACGCTATCGGTCAATTCAAATGGTGGCTATCATGGACTATTTCT  
AGAAGATTATTACGACTCTGATGAATTGATATGAAATCATTGTTAA  
CAATGGGAACACCTTTAACTTGAGAAAGTAACACCTCAAATCAT  
ACTCAAATGCTAAAGATTTAACAGTAATAAAGGAATATTCCATC  
AAGTCTCATGGTACAAATTGGCAGGAACTAATTATGATGGTG  
ATAAAATTGTTCCATTGCTAGTGTGGAGACTGGTAAATATATTCC  
AAGAAACCGCTAAACACTATACCCAACTAACAGTAACTGGTAAAT  
GCTACACATTCTGACTTGCCTGATAATTCTGAAGTTATCCAATATGTC  
GCAGAAAAAAATTCTAAAAATGAGAAAGGTAAATTACCAAAACCTC  
ACTAA

MLASLFIVRLSKSLRNSNMKLLRWLPPVLFIIILIGMTILGKSYINKVT  
AHKIKLYNSRMTPTILISGSSATQERFNSMLAQLNQMGEKHSVLKLTVK  
KDNSIYNGQISGNDHKPYIVIGFENNEDGYSNIKKQTKWLQIAMNDLQK  
KYKFKRFNAIGHNSNGGLSWTIFLEDYYDSDEFDMKSLLTMGTPFNFEES  
NTSNHTQMLKDLISNKGNIPSSLMVYNLAGTNSYDGDKIVPFASVETGK  
YIFQETAKHYTQLTVTGNNAHTSDLPDNPEVIQYVAEKILKNEKGKLPK  
PH

\*

ID-46

Clone 83

TTGAAATTAGGTATTACAACATCGGAGAGACAACAATCCTGAAGAAACAAACC  
AAAGCTATTACACATCCTGAGAGGATTGCCAATTAGTTGCTGAGATTGAACTAGCT  
GATCAAGTTGGTTAGATGTATATGGTATTGGAGAGCACCACATCGTGAAGATTTGC

**FIG. 1** CONT'D

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GGTCTCTGCACCGAAATTATCCTAGCAGCAGGAGCGGTTAGAACTAATAATATCC  
 GTTTATCTAGTCAGTAACGATTCTCTCTTCCAATGATCCTATTGCGCTATCAGC  
 AATTTCAACGATTGACGCACTTCAAATGGTAGAGCAGAAATTATGGCAGGGCGT  
 GGTCCTTATTGAGTCTTCATTGTTGGATACGATTAGCGGATTATGATGAT  
 TTATTAATGAAAAAATGGATATGTTAGCAATTAACTCAGCGACAAATCTGA  
 TTGGAAAGGTCAATTGACACAAACAGTTAATGAGCGACCAATTATCCAAGAGCAT  
 TACAAAGACAGTTATCAATATGGGTGGCAACAGGAGGAAATGTTGATTCTACAATT  
 CGTATTGAGAACAGGTTGCAATTGTTATGCAACTATTGGTGGGAATCCAA  
 AGCCTTCGTCAATTGGTCATATTATAAAGAAGTTGTAAGTCCGTAATGGACA  
 CAAACCAGGAACAACAACTAAAGTTGCTGCTCACTCTGGGGATGGATTGAAGAGGA  
 TAATCAAACCGCTATTGACCGTTATTTTCCCTACGAAACAGACCGTCGATAATAT  
 TGCTAAGGGACGCCCTATTGGTCTGAAATGACTAAAGAGCAGTATTACGTCAA  
 TAGGTCCAGAAGGTGCTATTTGTAGGAAATCCTGAAGTGGTGCACATAAAATT  
 ATAGGACTTTGGTGA

MKLGITTFGETTILEETNQSYSHPERIRQLVAEIELADQVGLDVFYIGEHIREDFAVSAP  
 EIILAAGAVRTNNIRLSSAVTILSSNDPIRVYQQFSTIDALSNRAEIMAGRGSFIESFPLF  
 GYDLADYDDLNEKMDMLLAINSATNLWKHLTQTVNERPIYPRALQRQLSIWVAT  
 GGNVDSTIRIAEQGLPIVYATIGGNPKAFRQLVHIYKEVGKSVMDTNQEQLKVAAHSW  
 GWIEEDNQTAIDRYFFPTKQTVDNIAKGRPHWSEMTKEQYLRSIGPEGAIFVGNPEVV  
 AHKIIGLW

ID-47

Clone 86

ATGATAGAGTGGATTCAAACACATTACCAAATGTATCAAATGGG  
 TTGGGAAGGTGCTTACGGCTGGCAGACAGCTATTGACAAACCCCTT  
 ATATGACTTTGGTCCTTATTGGAGGTTAACAGCTTAAATGGGATTGTTAG  
 GAGGTTATTCCCTGTTAACAGCTTAAATGGGATTGCTTAAAGCTC  
 AATTAGTATTGGAGTTAACAGCTTAAATGGGATTGCTTAAAGCTC  
 TGCCCTTCATTATTCTTCTTGCTTGAATTGCCAGTAACCTCGCGTAAT  
 TGTAGGAACAAACACTTGGTCACCAGCAGCTTGGTACCTCTTCTT  
 GGCAGTTCCCATTGGCTCGTCAAGTCAAGTTGTTAGCTGA  
 ACTTGATGGTGGAGTTATTGAGGCTGCACAAGCCTCAGGTGGAACAC  
 TTTGGGATATTATTGAGTTATCTCGTGAAGGTCTACCAGATTAA  
 TTCGAGTATCAACGGTACTTGAGTTCTTAGTGGCTATTACTA  
 TGGCTGGCGCTATTGGTGCAGGAGGATTGGGTTCTGGCTATTACTA  
 AAGGATATAACTATTCTCGTGAAGGTCTACCAGATTAA  
 TGATTTATTATAATTCTTATCCAATTAGGTGATTAAAC  
 ACGTCGCTTGAGTCATAAAATAA

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MIEWIQTHLPNVYQMGWEGAYGWQTAIVQTLYMTFWSFLIGGLMGLL  
 GGLFLVLTSPRGVIANKLVFGVLDKVSVFRALPFIILLALIAPVTRVIVG

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FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

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TTLGSPAALVPLSLAVFPFFARQVQVLAELDGGVIEAAQASGGTLWDII  
 VVYLREGLPDLIRVSTVTLISLVGETAMAGAIGAGGLGSVAITKGYNYSR  
 DDITLVATILILLIIFIQFLGDFLRRLSHK\*

ID-48 (same as ID-43)

ID-49

Clone 96

TTGGCAGTTAGTTTCAATGAAGTATTGGTGGGATTCTGCTTTTTA  
 TTATGATTATCAATATTCCATTGCTCCTCTTGCTACTTGGCTTAGG  
 TAAACAAACCTTTAAAAACTGTCTATGGTCTTGGATTTCCTGT  
 TTTTATTAAGTTAACACAAAGTGTACCAACTTGACCCACAACACT  
 CCTCGCAGCACTTTGGAGGTGTTAGTACAGGATGTGGTTGGGAT  
 TGTTTTGGAGCGACTCTCAACTGGTGGAACGGGATTATCATTCA  
 ATTCTTAGGAAAATATACTCCTATAAGCCTGGACAAGGGTTATAT  
 TGATTGATGGACTTGTACAAATTGGGTTCTAGCTTGTACAGTG  
 ATACGGTTATGTTCTATTATTGGGTTGATAACTATTAGTTATATT  
 TAATGCTATCCAAACTGGATTACAACCTTAAGCACTGTCTAATCGT  
 TTCTCAAGAGCACCAAAAAATTAAAGACATATATCAATACTGTCGAG  
 ATAGAGGAGTAACAGAAATTCCCGTTAAAGGGGGATTCTGGAACT  
 AATCAAATCATGCTTATGACAACATTGCTGGTTATGAGTTGCTAAA  
 TTACAAGAGGCAATAGCAGAAATTGACGAAACAGCCTTCATAACAGT  
 AACTCCAACATCACAAAGCTTCTGGACGTGGATTAGTCTTCAAAAAA  
 ATCATGGACGTCTGATGAAGACATTCTATGCCAATGTAA

MAVSFHEVFGWDSAFFIMIINIPLLLKYFGLGKQTFLKTVYGSWIFPVFI  
 KLTQSVPTLTHNSLLAALFGGVIVGCGLGIVFWSDSSTGGTGIIQFLGKY  
 TPISLGQGVILIDGLVTIVGFLAFSDTVMFSIIGLITISYINAIQTGFTTLST  
 VLIVSQEHQKIKTYINTVADRGVTEIPVKGGYSGTNQIMLMTTIAGYEFA  
 KLQEAIAEIDETAFITVTPSQASGRGFSLQKNHGRLEDILMPM\*

ID-50

Clone 99

ATGAAAGAAAAACAGTCAAAAGGCTTATTATATACTACTGATTGTTCCCATTAT  
 CTTTATAAGTGTACATACAGTATTAGCCAGCCTCTAAACTACTTCCACCAAA  
 AGAATTAGTTATTCTAACGTTAAAGCTAACAGGAACGATTCCAG  
 CTTTGAGGAAAAATACGGTATAAAAGTTAACAGCTTATTCAAGGTGGACAGGGCA  
 ACTAATAGATAGATTAAGTAAGGAGGGTAAGCAGTTGAAGGCAGGATATTCTTG  
 GAGGAAATTATACGCAATTGAAAGTCATAAGGCATTGTTGAGTCTTACGTATCA

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FIG. 1 CONT'D

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AAGAATGTTCATACTGTTATTCCAGACTATATCCATCCGAGTGATACGGCGACACC  
 TTATACTATAAAATGGGAGTGTCTGATTGTAATAACGAATTAGCTAAGGGACTTA  
 CCATCAAGAGTTATGAAGATTATTACAGCCTCCTAAAAGGTAAAATTGCCTT  
 GCAGATCCTCTAGAGTCGACCTGCAAGCATGCAAGCTGGCGTAA

MKEKQSKRLIYILLIVPIIFISVFTYSISQPSKLLPPKELVILSPNSQAILTGTTIPAFEEKYGI  
 KVVKLIQGGTGQLIDRLSKEGKQLKADIFFGGNYTQFESHKALFESYVSKNVHTVIPDYI  
 HPSDTATPYTINGSVLIVNNELAKGLTIKSYEDLLQPSLKGKIAFADPLESTCKHASLA

ID-51

Clone 103

CCTCCTATCAAATGATGACAAACGTGAGAGGTACATGGAACAAATGCTCTTAAAAA  
 TTGAAAATGCAACCTGGCAGCGTGTGGTAAGAGCACTTTATCGTAAATACAATAAG  
 GAATTTTTACATATCCAGCCGAAAACAAACACCACGCTTTGAATCAGGATT  
 GGCATATCACACGGCAACAATGGTCGTTGGCAGATAGTATCGGAGATATCTATC  
 CAGAACTTAATAAAAGTTGATGTTGCTGGTATTATGCTACATGATTAGCCAAG  
 GTCATAGAGTTATCGGGTCCTGATAATACAGAATATACTATTGAGGTAATCTTAT  
 CGGTATTTCACTTATTGATGAGGAATTAA

LLSNDDKRERYMEQMLFKIENATWQRVVRALYRKYNKEFFTYPAAKTNHHAFESGL  
 AYHTATMVLADSIGDIYPELNKSLMFAGIMLHDLAKVIELSGPDNTEYTIRGNLIGHIS  
 LIDEEL

ID-52

Clone 104

ATGAAAAAAAATAAAATTATCCGATTCACTCAGTTAGTTGGTGTCTACTT  
 GCGATACTATGCTTAGTCTTTGCTTATTGAAGCCTAACAGTCAA  
 CAATCATCATCTCAAAAGTTGAGGAATGAGGATATAAAAAGACATC  
 CTCTCAAAAAGAAATAAGAAATTACGATTACCAAGCTGTATCATCAA  
 AAGATTGGAACCTGATTTGGTCAATCGTGACCATAAACATGAAGAA  
 TTAAGTCCAGATGTGGCCTGTTGAAAATATTATTGGATAAACGT  
 ATTACGAAGCAAGCTACTCAGTTTAGAGGCTGCTAGAGCAATTGA  
 TTCACGAGAACATTAAATTCCGGTTATCGTAGTGTGCTATCAGGA  
 GAAGTTGTTCAATTCTATGTTACTCAAGAGATGACTAGTAACCTAA  
 TTTGACGAGGGACAAGCAGAAAAGTTGGTAAAACCTACTCTCAGC  
 CTGCAGGTGCTAGTGAACACCAGACTGGATTAGCGATGGATATGAGT  
 ACTGTAGATTCTTGAATGAGAGCGATCCTAGAGTAGTCAGTCAGTT  
 GAAAAAGATAGCTCCACAATATGGTTTGTCTACGGTTCCGGATG  
 GTAAAAACAGCAGAACAGGGTAGGTTATGAAGATTGGCATTACCG

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FIG. 1 CONT'D

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CTATGTTGGGGTAGAGCTGCAAAATATGGTCAAACATCATTAA  
CATTAGAAGAATACATAACTTATTAAAGGAGAATAACCAATGA

MKKNKIIRFSLVGVLLAILCFSLFALLKPNSQQSSQKLRNEDIKKTSSQK  
RNKKLRLPAVSSKDWNLILVNRDHKHEELSPDVVPVENIYLDKRITKQA  
TQFLEAARAIDSREHLISGYRSVAYQEKLFSYVTQEMTSNPNLTRGQA  
EKLVKTYSQPAGASEHQTGLAMDMSTVDSLNESDPRVVSQQLKIAPOY  
GFVLRFPDGKTAETGVGYEDWHYRYVGVESAKYMVKHHTLEEYITLL  
KENNQ\*

ID- 53

Clone 106

CTGTTATGTGGATTCTTCCATCAATTCTGTGCTAATTCCGGGGGG  
TATGGTATAATAACAGTTATGAAAAATAAAAAAAATCTTATTGGGAC  
TGGCCTTGGTGTGGGTTACTGGCAGCTGCTGGTTACCTAAC  
TAAAAAAAGTAACAGATTATAAACGTCAGCAAATCACTCAGACCTTAA  
GAGAACTTTTAGTCAGATGGGTGATATTCAAGGTATTTATTTAATG  
AATTGAATCTGATATTAAATGACCAGTGGTGGTCTTGTCTTGGAA  
GATGGCAGAATTTCGAATTCAATTACGTCAAGGTGTTCTGATTAT  
GTGGAGGTGAGCAAATGA

LLCGFLPSIPVNSGGYGIITVMKNKKILFGTGLAGVGLAAAGYLT  
KVTDYKRQQITQTLRELFSQMGDIQVFYFNEFESDIKMTSGGLV  
LEDGRIF  
EFTYRQGVLDYVEVSK\*

ID-54

Clone 108

ATGTATCAAACTCAGACAAATAAGGAAAAATTGTTTATTGAAATTATTC  
CCAGTATTGATTCAATTGCTAATTTCAGCTACTTTATTGATTGGTTATGA  
CTGGACAGTATAGTCAGCTACATTGGCAGGTGTCAACTGCTAGTAATTATGG  
ACTCCGTTTCGCTTATTAGTAGGTATGATTCAAGCATTAGTACCAAGTAGTTGGT  
CAACATTGGTAGAGGAAATAAAGAACAAATTGCACAGAATTCAATTCT  
ATATTAGGTTGATACTGTCCTAA

MYQTQTNKEKFVLFLKFIPVLIYQFANFSATFIDSVMTGQYSQLHL  
AGVSTASNLWTP  
FFALLVGMISALVPVVGQHLGRGNKEQIRTEFHQFLYLGILSL

ID-55

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FIG. 1 CONT'D

## Clone 112

CTGCTCTTTAGCTAACCTTCTAATTATGGTATAATTGTATGGATT  
 GTTAGCTAGAATGGAGAAGATGATGCAAGATGTTTCAATTATAGGA  
 AGTAGAGGGTTGCCAGCTCGTTACGGTGGTTGAAACCTTTGTTCA  
 GAATTGATTAATCATCAAAAAAGTCCGACATAAAATACCATGTTGC  
 ATGCCTTAGTGATAAAGAACATCATACTCATTAACTTGTGCTGACGC  
 TGATTGTTTACTATAAATCCTCCCAATTAGGGCCAGCACGTGTGAT  
 TGCTTATGATATTATGCCATTAAATTATGCCCTGACTTGGTTAAGAC  
 ACATGATTAAGAGCCTATTAAATTTAGGAAATACAATTGG  
 TGCCTTATTGGCATTGGCCAATAAAACATAAAAGTCGGTGGCTT  
 ATTGTATGTTAATCCGGATGGTTAGAGTGGAAAGCGATCAAAGTGGT  
 CTCGCCCCACACAGCGTTATTAAATACGCCAAAAATGTATGACT  
 AAAATGCAGACCTAATTATTCTGATAATTGGTATTGAAAATTA  
 CATTCAATCTACCTACTCTAATGTGAAGACAAGGTTATTGCTTACGG  
 TACAGAGATTAAATTCTAGGAAATTATCGTCAGATGATCCACGTGTCA  
 AACAGTTGTTAAAAAATGGAATTAAAGTCTAAGGTTACTATCTA  
 ATCGTTGGTCGATTGTCCTGAAAACAATTATGAAACGGCTATTAG  
 GGAGTTCATGGCTTCAGACTAAAGCGTGATTAGTTATTATCTGTA  
 CCATCAAAATAACCCCTACTTGTAAAAGTTGTCTAAAGACAAACC  
 TTCAACAAGATAAAAGAGTTAAGTTGAGGTACGCTCTATGAAAAAA  
 GATCTGCTGGATTATGTCGTCAACAAGCCTTGCTTATATTATGGG  
 CATGAAGTTGGCGGTACTAATCCAGGACTGCTTGAGGCTTAGCTAA  
 TACTGATTGAATCTTGTCTAGATGTTGATTCAACAAATCAGTAGC  
 AGGTCTCTCAAGTTTACTGGACTAAAAAAGAGGGGGATTAGCTA  
 AGCTT

MLFLANFSNLWYNCMDCLARMEKMMQDVFIIGSRGLPARYGGFETFVS  
 ELINHQKSSDIKYHVACLSDKEHHHTHFNFADADCFTINPPQLGPARVIALY  
 DIMAINYALDLVKTHDLKEPIFYILGNTIGAFIWHFANKIHKVGGLLYVN  
 PDGLEWKRSKWSRPTQRYLKYAEKCMTKNADLIISDNIGIENYIQSTYSN  
 VKTRFIAYGTEINSRKLSDDPRVKQLFKWNIKSKGYYLIVGRFVPENN  
 YETAIREFMASDTKRDLVICNHQNNPYFEKLSLKTNLQQDKRVKFVGT  
 LYEKDLLDYVRQQAFAYHGHEVGGTNPGLEALANTDLNLVLDVDFN  
 KSVAGLSSFYWTKKEGDLAKL

ID-56

## Clone 120

TTGAGGAGTAATATGGTAAAGACAGCAGTTTAATGGCGACATACAA  
 TGGCGAAAAATTATATCTGAACAACTTGATTCAATTGCCAACAGA  
 CATTAAAACCAGATTATGTATTATTGAGGGATGATTGTTCAACGGAT  
GAAACAGTCAATGTCGTCAATAACTATATCGCAAAACATGAGTTAGA

FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

AGGCTGGAAAATTGTTAAAAACGACAAAAACTTAGGCTGGCGTTAA  
ATTTCTGTCAATTACTTATTGATGTGTTAGCCTATGAGGTTGACTATG  
TCTTTTTAGTGTCAAGATGATATTGGTATCTTGATAAAAACGAAC  
GACAGTTGCCATTATGTCAGATAACCCTCAAATTGAGGTTTGAGTG  
CAGACGTTGATATCAAAACGATGTCTACAGAAGGCCAGTGTCCACAT  
TTTCTAACTTTTCTTAGTGTAGAATCAGTCAGTATCCTAAAGTA  
TATGATTATCAAACATTCCGTCCCGGATGGACCATTGCTATGAAGAG  
AGATTTGCGCAAGCTATCGCTTGA

MRSNMVKTAVLMATYNGEKFISEQLDSIRQQTLKPDYVLLRDDCSTD  
VNVVNNYI<sup>A</sup>KHELEGWKIVKNDKNLGWR<sup>B</sup>LNFRQ<sup>C</sup>LLIDV<sup>D</sup>LA<sup>E</sup>YEV<sup>F</sup>DYV<sup>G</sup>FF  
SDQDDIWYLDKNERQFAIMS<sup>H</sup>DNPQIEV<sup>I</sup>LSADVD<sup>J</sup>IKTM<sup>K</sup>STEAS<sup>L</sup>VPHFLTFS  
SSDRISQYPK<sup>M</sup>VYDYQTFRPGWT<sup>N</sup>IA<sup>O</sup>M<sup>P</sup>KRDEAOAJA\*

ID-57

Clone 123

GTGATTATGGATAAGTCTATCCTAAAGCAACTGCTAACGTTATCA  
CTGTACTACCGTATTAAACGTTAATACTGATGGCATCGAAAAAA  
GCTAGTTCAAACAAATTGCAGATGCCCTAGGTATCGATTCTGCTACT  
GTTCGACGTGATTTCTTATTTGGTGAACTAGGACGCCGTGGTTT  
GGTTATGATGTCAAAAAACTTATGAACCTCTTGCAGAAATATTGAA  
CGATCATTCTACAACAAATGTTATGCTGGTGGGGTGTGGAAATATCG  
GTAGAGCTCTTGCAATTATCGTTCCACGATCGCAATAAAATGCAA  
ATTCATGGCTTTGATTAGATAGCAATGATTAGTTGGTAAACACA  
ACCGAGGATGGAATTCTGTCTACGGTATTGACTATCAATGACCA  
TTAATAGATAGTGTATTGAAACTGCTATCCTAACAGTACCTAGTAC  
AGAAGCCCCAAGAAGTTGCTGACATCTTAGTCAAAGCAGGTATAAAA  
GGCATCTTGAGTTCTCCAGTTCAATTAAACATTACCAAAAGATATC  
ATTGTCAGTATGTAGATTAAACAAGCGAATTACAAACTTACTTTAT  
TTCATGAACCAGCAGCGATAA

MIMDKSIPKATAKRLSYYRIFKRFNTDGIEKASSKQIADALGIDSATVRR  
DFSYFGEGLRGFGYDVKKLMNFFAEILNDHSTTNVMLVGCGNIGRALL  
HYRFHDRNKMQISMAFDLDSNDLVGKTTEDGIPVYGISIINDHLIDSIE  
TAILTVPSTEAQEVADILVKAGIKGILSFSPVHLTLPKDIIVQYVDLTSELQ  
TLLYFMNQQR\*

ID-58

Clone 125

FIG. 1 CONT'D

ATGGGTGCTAAAGGAGCAGATGTCATTCTCGTTTATCACACTCTGGCATTGGAGA  
TGATCGATATGAAGAAGGTGAAGAAAACGTTGGCTATCAAATTGCCAGCATCAAG  
GGAGTGGATGCCGTTACGGGACACTCACACGCTGAATTCCATCAGGTAAACGG  
TACTGGCTTCTATGAAAAAATACACTGGAGTTGATGGTATCAATGGAAAAAATAATG  
GAACACCTGTTACAATGGCAGGCAAGTACGGGGATCACCTGGTATTATTGATTAA  
GGACTTAGTTATACTAATGGAAAATGGCAAGTCTCCGAAAGCAGTGCTAAAATCC  
GTAAAATTGATATGAACTCAACAACGTGCTGACGAGCGTATCATTGCAATTGGCTAAG  
GAAGCACACGATGGCACTATCAACTATGTTGCCAACAGTAGGTACAACAACTG  
CGCCAATTACAAGTTACTTGCACTAGTTAA

MGAKGADVLVLSHSGIGDDRYEEGEENVGYQIASIKGVDAVVTGHSHAEFPSGNGTGFYEKYTGVDGINGKINGTPVTMAGKYGDHLGIIDLGLSYTNGKWQVSESSAKIRKIDMNSTTADERIIALAKEAHDTINYVRQQVGTTPAPITSYFALV

ID-59

Clone 135

TTGTCAATAAGGTTCAAATCAGCTGAAATATGATAAAATAAAACAGATTGTAAG  
TGACTGTTAAGCTGTTTCAGAGAGGTTTATGAATACAAACACAATAAAAAA  
AGGTTGTAGCGACTGGAATTGGAGCTGCACTTTTATCATTATAGGTATGCTAGTT  
AA

MSIRFQISLKYDKIKQIVSDCLSLFFREVFMNTNTIKKVVATGIGAALFIIIGMLV

ID-60

Clone 145

ATGAAACATTAAAATTCAATCGGTCTCGACATTATTGGTCCTGTTATGATTGGA  
CCATCAAGTAGTCATACTGCAGGAGCTGTCCGCATTGGTAAAGTTGTCCATTCTAT  
TTTTGGTGAACCTAGTGAAGTAACCTTCATTATACAATTCTTTGCTAAAACCTA  
CCAAGGACACGGTACTGATAAAAGCATTGGTTGCAGGGATTCTAGGAATGGATACA  
GATAATCCAGATATTAA

MKHLKFQSVDIIGPVMIGPSSSHAGAVRIGKVVHSIFGEPSEVTFHLYNSFAKTYQG  
HGTDKALVAGILGMDTDNPDI

ID-61

Clone 147

FIG. 1 CONT'D

GTGTCAGAAGGTGTTAATGTTCTAAAAGAAGATGACGTAGAGACTTTCTCA  
TATCCTGACAAATTCAATTAGCCAATTATGGCACAAATTGATTGTGTCAAGGA  
AATGATTAA

ID-62

Clone 150

ATGACCTACAAAGATTACACAGGTTAGATCGGACTGAACTTTGAGTAAAGTGCG  
TCATATGATGTCCGACAAACGTTTAA

MTYKDYTGLDRTEIJLSKVRHMMSDKRE

ID-63

### Clone S2

CTGAGTTGGGTCTTGGAAACGGTCCTGTCAATCATACTAGCTATCAAGGAGACTAA  
AATGTATTTAGAACAACTAAAAGAGGGTAAATCCTTAA

MSWVLETVLSIILAIKETKMYLEQI KEVNPI

ID=67

Clone 3-40

GTGAAAAAAAAATTAGTCTCATCACTTCTAAAGTGTCTCTAATCATT  
ATTGTTAGCTTGCTGGAGCATTGCTAGTTTGTATGAATCAT  
AATGACAATATTCCAAATGGTGGTGTCACTAAAAGTAGTAAAGTAAA  
TTATAATAACATAACGCCTACAACAAAAGCTGTTAAAAGGTACAAA  
ATAGTGTGTTCTGTTATCAATTATAACACAAGAGAGTCGTTCTG  
ACCTATCAGACTTCTATAGTCATTTTTGGTAATCAGGGGGCAACA  
CTGATAAGGGCTTACAAGTTACGGTGAAGGCTCTGGAGTCATCTAT  
AAAAAAAGATGGTAAAAATGCCTATGTTGTCACTAATAACCACGTCAT  
TGATGGGGCTAAACAAATTGAAATTCAACTAGCTGATGGCTCAAAAG  
CAGTTGGAAACTTGTGGGTCAAGATAACCTACTCTGATTTAGCCGTCG  
TCAAAATTCCATCAGATAAAGTTCAAATATTGCAAGAATTGCTGATT  
CATCAAAACTCAACATTGGTGAAGACTGCTATAGCGATCGGAAGCCCT  
CTTGGAACTGAGTATGCAAATTCTGTAACTCAAGGTATTGTATCTAGT  
TTAAAAAGAACTGTAACAATGACTAATGAAGAAGGACAAACAGTT  
CTACAAATGCTATCCAGACGGATGCTGCTATCAATCCTGGTAATTCA  
GGTGGAGCACTTATCAATATTGAAGGACAGGTTATTGGAATTAAATTCA  
TAGTAAAATTCTTCTACATCAAATCAAACCTCAGGACAATCGTCAG

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

GAAATAGCGTTGAAGGTATGGGATTGCCATTCTCAAATGATGTT  
GTTAAGATTATCAATCACTTGAGAGTAACGGACAAGTAGAGAGACC  
TGCTCTAGGTATTCTATGGCTGGATTAAGTAATTACCATCCGATGT  
TATTAGTAAACTGAAAATCCCAAGTAATGTTACTAATGGTATTGTAG  
TAGCATCTATCCAATCTGGCATGCCAGCTCAAGGCAAACAAAGAAA  
TACGATGTCATTACTAAAGTTGACGATAAAGAAGTAGCATCTCCAAG  
TGATTACAAAGTTACTCTATGCCACCAGGTAGGGGATTCCATAAA  
CAGTAACCTTTATCGTGGTAAAAATAAACAAACAGTCACTATAAAA  
CTTACTAAAAGTAGTAAAGATTAGCTAAACAAACGAGCAAATAACTA  
A

MKKKLVSSLKCSLIIVSFAGGAFAFVMNHNDNIPNGGVTKTSKVNY  
NNITPTTKAVKKVQNSVSVINYKQQESRSDFYSHFFGNQGGNTDK  
GLQVYGEFSGVYKKDGKNAYVVTNNHVIDGAKQIEQLADGSKAVGK  
LVGSDTYSDLAVVKIPSDKVSNIAEFADSSKLNIGETAIAIGSPLGTEYAN  
SVTQGIVSSLKRTVTMTNEEGQTVSTNAIQTDAAINPGNSGGALINIEGQ  
VIGINSSKISSTSNTSGQSSGNSVEGMGFAIPSNDVVKIINQLESNGQVE  
RPALGISMAGLSNLPSDLQSLLYGHQVGDSITVTFYRGENKQTVTIKLTKT  
SKDLAKQRANN\*

ID-68

Clone 3-30

ATGTTAAAATGGTATACAAACAAAGGAGGGAGGATGATAATGAAGA  
AATGTTTTGGCTATTGTTAGCTCTAGTTTTATGGTTCACT  
TCAAGCAGATGAGGTGGACTATAACATTCCCTCATTATGAGGGTAATC  
TAACATTACACAATGATAATAGTGCTGATTACAGAGAAGGGTACTT  
ACCAATTGATTGCTCTATAATGGACAGTATGTCACGTTAGGTACG  
GCGGGTAAGTTATCTGACAATTGATATAATAATAAGCCACAGGT  
TGAAGTTCAATTAAATGGTAAAGTAAGGAAAGTTAGTTACCAAGATAAG  
AAGATTGGAGGATGGCTACCGTTGAAAGTGTAAATGGTGGTGAA  
GCAGGGTATACTGTTAAAGTCATGTTAGGGAAACTAAAAAAATGT  
TCTATTATGCATAAGGATGTTGGTGAACCTAACGGATTCTTATTAG  
CGACTGGATAAAACGTTAGAGAAAGTAGATTGGATATCAACTG  
ACAAAAAAGGGTGCCTTCTCGTCTTGGGGCACTGGTTATCTTA  
AAACTCCTCCTAAAATAAGACAAAATAATAATCGTTACCATTTGACA  
GCTTTAATGTAACAAACGATTAGAATTCTATGGTTATTGGGATAG  
ATCTTATTAAATCTACCTACAAACAGTAAAATAATTACAAGAAAA  
AAATTGAACATCAAGAGAAGATAATAGAGCGTCATGGTTTATCCTA  
AGTTCTTGTAAAGGATATTACCTTCATTCTTATTATTGTGACAC  
TATTCTCATCTCAATTAGGGTGTCTGTTAGAAAAAAAGTTAATAAAT

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

ACGGGCAATTCCCTAAGGATCATCATTATATGAAGCACCTGAGGAC  
 CTTTCACCATTAGAGTTAACTCAAAGCATTATAGTATGAGCTTAAA  
 AATTTCAAGATGAGGAGAAGAAAAACTCACCTTATCAGTCAGAACAA  
 ACTCATAACAGTCAATTCTATTAGACTTGATTGATAGAAAAGTATTGA  
 ATTATGATGATAACTTGTATCTCTAGCTAACTTAGATAGAGCTTCTG  
 ATGCAGAAATAGATTATAGAGTTGCTTGCAGATTACAGAGTT  
 TGAAGCCAGATCAACTCTTCTAATTACCAATTAGTTATAAAGAAA  
 CACTACGTGAACGTGAAAAAGCAGCACAAAGGCTCAGATCTGCAAAAT  
 CAAATGAGACGCCGAGGAAGTAATGCCTTATCAAGAATTACGCGTCT  
 CACAAGGTTGATTCTAAAGACAATATAAACTCTCTAGAAGAAAAGG  
 GAATTTCATCCCCCTATCGTAAAATGTCTCAGAAGAGTCTAAAGAA  
 TTATCTAGGTTAAAAGATTCACTTACCTATCACCTCTATTCTTTG  
 TTGTTATAATTATACGCTTTTAAATTATTTACCTATTCTGTAT  
 CTATCTCTTATTGTTGGTGTATCCTGTTGAATAAAATCATT  
 ATGATGACAAGAAAAATAAGTAACGGTTATATTGTAACTGAAGATGG  
 AGCAAGTCGTGCTACCAATGGACTAGTTTAGGAACATGCTAAGGG  
 ATATCAAATCGTTGATCGTTAGAGTTAGAAAGTATCGTATTATGG  
 AATCGAATATTGGTTACGCTACTTATTGGCTACGCTGACCGTGT  
 GAGAAAGTACTCAGAGTGAACCAAATAGATATTCCAGAAAGATTGC  
 AAACATTGATAGTCATCGATTGCGATTCACTGAAATCAATCTAGTAA  
 TCATTTCACGATAACTGAAGATGTTAGTCAGCTCTAATT  
 TGTAAATTCAAGGGGTTCTCAGGTGGTTCTCAGGCAGGAGGCG  
 GCGGAGGTGGCGGTGCCTTCTAA

MLKWYTNKGRMIMKKCFLAICLALSFFMVSQADEVDYNIPHEGNL  
 TIHNDNSADFTEKVTVYQFDSSYNGQYVTLGTAGKLSDNFDINNKPV  
 SINGKVRKVSYQIEDLEDGYRLKVFNGGEAGDTVKVNVQWKLKNVLF  
 MHKDVGELNWIPISDWDKTLEKVDWFISTDKKVALSRLWGHLGYLKTP  
 PKIRQNNRYHLTAFNVNKRLEFHGYWDRSYFNLPTNSKNYYKKKIEH  
 QEKKIERHGFLSFLRILLPSFFIVTLFISIRVFLFRKVNKYGQFPKDHH  
 YEAPEDLSPLELTQSIYSMSFKNFQDEEKTHLISQEQLIQSILLDRKV  
 LNYDDNLLSLANLDRASDAEIDFIEFADSTSLKPQLFSNYQFSYKET  
 LRELKKQHKASDLQNQMRRGSNALSRTLTRLISKDNINSRRKG  
 PYRKMSSEESKELSRLKRFSYLSPLISFVVIYTLFLNYFTYFCIYLLFGV  
 LLLNKIIFMMTRKISNGYIVTEDGASRVYQWTSFRNMLRDIKSFDRSELE  
 SIVLWNRILVYATLFGYADRVEKVLRVNQDIPERFANIDSHRFAISVNQS  
 SNHFSTITEDVSHASNFSVNSGGSSGGFSGGGGGGGAF\*

ID-69

Clone 3-38

ATGATGATTGTGAATAATGGTTATCTAGAAGGGAGAAAAATGAAAAA  
AGAGACAAAAAAATATGGAGAGGGTTATCAGTTACTTACTAATCCTG

FIG. 1 CONT'D

SUBSTITUTE SHEET (RULE 26)

\_\_\_\_\_  
TCCCAAATTCCATTGGTATATTGGTACAAGGTGAAACCCAAGATA  
CAATCAAGCACTTGGAAAAGTAATTGTTAAAAAAACGGGAGACAAT  
GCTACACCATTAGGCAAAGCGACTTTGTGTTAAAAATGACAATGA  
TAAGTCAGAAACAAGTCACGAAACGGTAGAGGGTCTGGAGAAGCA  
ACCTTGAAAACATAAAACCTGGAGACTACACATTAAAGAGAAGAAA  
CAGCACCAATTGGTTATAAAAAACTGATAAAACCTGGAAAGTTAAA  
GTTGCAGATAACGGAGCAACAATAATCGAGGGTATGGATGCAGATA  
AAGCAGAGAAACGAAAAGAAGTTGAATGCCAATATCCAAAATC  
AGCTATTATGAGGATACAAAAGAAAATTACCCATTAGTTAATGTAG  
AGGGTTCAAAGTTGGTGAACAATAACAAAGCATTGAATCCAATAAAT  
GGAAAAGATGGTCGAAGAGAGATTGCTGAAGGGTGGTATCAAAAAA  
AAAATCCAGGGTCAATGATCTCGATAAGAATAATATAAAATTGAA  
TTAACTGTTGAGGGTAAAACCACTGTTGAAACGAAAGAACTTAATCA  
ACCACTAGATGTCGTTGTCTATTAGATAATTCAAATAGTATGAATA  
ATGAAAGAGCCAATAATTCTCAAAGAGCATTAAAAGCTGGGGAAAGC  
AGTTGAAAAGCTGATTGATAAAATTACATCAAATAAAGACAATAGA  
GTAGCTCTGTGACATATGCCTCAACCATTGATGGTACTGAAGCG  
ACCGTATCAAAGGGAGTTGCCGATCAAATGGTAAAGCGCTGAATG  
ATAGTGTATCATGGGATTATCATAAAACTACTTTACAGCAACTACA  
CATATTACAGTTATTAAATTAAACAAATGATGCTAACGAAGTTAA  
TATTCTAAAGTCAGAATTCAAAGGAAGCGGAGCATATAATGGG  
GATCGCACGCTCTATCAATTGGTGCACATTACTCAAAGCTCTA  
ATGAAAGCAAATGAAATTTCAGAGACACAAAGTTCTAATGCTAGAAA  
AAAACCTATTTTACGTAACTGATGGTGTCCCTACGATGTCTTATGC  
CATAAATTAAATCCTTATATATCAACATCTTACCAAACCAAGTTAA  
TTCTTTAAATAAAATACCAAGATAGAAGTGGTATTCTCAAAGAGG  
ATTTATAATCAATGGTGTGATTATCAAATAGTAAAAGGAGATGGA  
GAGAGTTAAACTGTTTCGGATAGAAAAGTCCCTGTTACTGGAGG  
AACGACACAAAGCAGCTATCGAGTACCGCAAATCAACTCTCTGTA  
TGAGTAATGAGGGATATGCAATTAAATAGTGGATATATTATCTCTATT  
GGAGAGATTACAACACTGGGTCTATCCATTGATCTAACGACAAGAAA  
GTTCTGCAACGAAACAAATCAAACACTCATGGTGAGCCAACACATT  
ATACTTTAATGGAAATATAAGACCTAAAGGTTATGACATTCTACTGT  
TGGGATTGGTGTAAACGGAGATCCTGGTGCAACTCCTCTTGAAGCTG  
AGAAAATTATGCAATCAATATCAAGTAAAACAGAAAATTATAACTAAT  
GTTGATGATACAAATAAAATTATGATGAGCTAAATAACTTTAA  
AACAAATTGTTGAGGAAAACATTCTATTGATGGAAATGTGACTG  
ATCCTATGGGAGAGATGATTGAATTCCAATTAAAAATGGTCAAAGT  
TTTACACATGATGATTACGTTGGTGGAAATGATGGCAGTCAATTAA  
AAAAATGGTGTGGCTCTTGGTGGACCAAACAGTGTGATGGGGAAATT  
AAAAGATGTTACAGTGACTTATGATAAGACATCTCAAACCATCAAAA  
TCAATCATTGAACCTAGGAAGTGGACAAAAGTAGTTCTTACCTAT  
GATGTACGTTAAAAGATAACTATATAAGTAACAAATTACAAATAC  
AAATAATCGTACAACGCTAAGTCCGAAGAGTAAAAAGAACCAAAT  
\_\_\_\_\_

FIG. 1 CONT'D

ACTATTCGTATTCCAAATCCCCAAATCGTGATGTTCTGTGAGTTT  
CCGGTACTAACCATCAGTAATCAGAAGAAAATGGGTGAGGTTGAATT  
TATTAAGTTAATAAAAGACAAACATTCAAATCGCTTTGGGAGCTA  
AGTTCAACTTCAGATAGAAAAAGATTTTCTGGGTATAAGCAATT  
GTTCCAGAGGGAAGTGTACAAACAAAGAATGATGGTAAAATTAA  
TTTAAAGCACTCAAGATGGTAACTATAAATTATATGAAATTCAA  
GTCCAGATGGCTATATAGAGGTTAAAACGAAACCTGTTGTGACATT  
ACAATTCAAAATGGAGAAGTTACGAACCTGAAAGCAGATCCAAATG  
CTAATAAAAATCAAATCGGGTATCTTGAAGGAAATGGTAAACATCTT  
ATTACCAACACTCCAAACGCCACCAGGTGTTTCTAAACAGGG  
GGGAATTGGTACAATTGTCTATATAATTAGTTGGTCTACTTTATGAT  
ACTTACCAATTGTTCTTCCCGTCGTAACAAATTGTAA

MMIVNNNGYLEGRKMKKRQKJWRGLSVTLLILSQIPFGILVQGETQDTNQ  
ALGKVIVKKTGDNATPLGKATFVLKNDNDKSETSHETVEGSGEATFENI  
KPGDYTLREETAPIGYKKTDKTWKVKVADNGATIIEGMDADKAERKE  
VLNAQYPKSAIYEDTKENYPLVNVEGSKVGEQYKALNPINGKDGRREIA  
EGWLSKKNPGVNDLKDKNKYKIELTVEGKTTVETKELNQPLDVVULLDN  
SNSMNNEERANNSQRALKAGEAVEKLIDKITSNKDNRVALVTYASTIFDG  
TEATVSKGVADQNGKALNDSVSWDYHKTTFATTHNYSYLNLTNDAN  
EVNILKSRIPKAEAHINGDRTLYQFGATFTQKALMKANEILETQSSNARK  
KLIFHVTDGVPTMSYAINFNPYISTSYQNQFNSFLNKIPDRSGILQEDFIIN  
GDDYQIVKGDGESFKLFSDRKVPVTGGTTQAAYRVPQNQLSVMNSEGY  
AINSGYIYLWWRDYNWVYPFDPKTKVSATKQIKTHGEPTTLYFNGNIR  
PKGYDIFTVGIGVNGDPGATPLEAEKFMQSISSKTENYTNVDDTNKIYDE  
LNKYFKTIVEEKHSIVDGNVTDPGMEMIEFQLKNGQSFTHDDYVLVGND  
GSQLKNGVALGGPNSDGGILKDVTVTYDKTSQTIKINHLNLGSGQKVVL  
TYDVRLKDNYISNKFYNTNNRTTLSPKSEKEPNTIRDFPIPKIRDVREFPV  
LTISNQKKMGEVEFIKVNKDKHSESLLGAKFQLQIEKDFSGYKQFVPEGS  
DVTTKNDGKJYFKALQDGNYKLYEISSPDGYIEVTKPVVTFTIQNGEVT  
NLKADPNANKNQIGYLENGKHLITNTPKRPPGVFPKTGGIGTIVYILVG  
STFMILTICSFRRKQL\*

ID-70

Clone 141

ATGAATAGAAAAGTTGAGGAAAAAATGGCTGGGAATCGTAATAACG  
ATATGAATGTCTATTGTTCA~~TTTGTGGCAAAAGCCAAGATGAAGTAA~~  
AAAAAAATTATTGCAGGTAATGGTGT~~TTTCA~~TTGTAATGAATGTGTG  
GCCTTATCACAAGAAATTATTAAGGAAGAATTAGCTGAGGAAGTACT

FIG. 1 CONT'D

**SUBSTITUTE SHEET (RULE 26)**

GGCTCATTAGCAGAAGTACCAAAACCTAAGGAACATTAGAAATAT  
 TAAATCAATATGTTGAGGGCAAGATCGTGCTAAACGTGCTTAGCA  
 GTTGCTGTCTACAATCATTACAAGCGTGTAGTTATACCGAGAGTAGT  
 GACGATGATGTAGATTGCAAAAATCCAACATTGATGATTGGTCC  
 AACTGGCTCAGGAAAAACCTTCTAGCACAAACACTGGCTAAAAGCC  
 TTAATGTACCGTTGCTATTGCAGATGCGACTTCATTGACCGAAGCAG  
 GATACGTTGGAGAAGATGTTGAGAATATTCTTCTAAATTGATTCAA  
 GCTGCTGATTATAATGTCGAACGTGCTGAGCGTGGTATTATCTACGTT  
 GATGAAATAGATAAAATTGCTAACGAAAGGCGAAAATGTTCTATCAC  
 ACGTGATGTTGCTGGTGAAGGTGTACAGCAAGCCCTTCTAAAATTAA  
 TTGAGGGTACGGTAGCAAGTGTCCCCCACAGGGTGGCGTAAACAT  
 CCTAACCAAGAAATGATTCAAATTACCAAGAACATCCTTTTATT  
 GTCGGTGGTGTCTTGATGGTATTGAAGACCTTGTGAAGCAACGTTA  
 GGCAGAAAAGTTATTGGTTTGACAGACAAGCCGAAAATTGATGA  
 CAACGCTTCTTATATGCAAGAGATAATTCTGAGGATATTCAAAAGT  
 TTGGACTGATTCCAGAGTTATTGGCCGTTACCACTAGTAGTTGCAGCGT  
 TAGAACTCTTACTGCAGAAGATCTGGTTCGTATTCTGACAGAACCA  
 CGCAATGCTTGGTAAACAATACCAAAACCTTATTATCTTATGATGGT  
 GTAGAAATTGGATTGACCAAGGATGCTTATTGGCTATCGCTGATAA  
 GGCTATCGAGCGCAAGACTGGTGCACGTGGTTACGTTCTATTATTG  
 AAGAAACGATGCTGATATCATGTTGAAATTCCAAGCCAAGAAGAT  
 GTAACAAAAGTCGTATCACAAAGGCTGCTGTTGAGGGTACTGACAA  
 GCCTGTTTAGAGACGGCTTAG

MNRKVEEKMAGNRNNDMNVYCSFCGKSQDEVKKIAGNGVFICNECV  
 ALSQEIIKEELAEEVLAHLAEVPKPKELLEILNQYVVGQDRAKRALAVA  
 VYNHYKRVSYTESSDDDVLQKSNILMIGPTSGKTFLAQTLAKSLNVP  
 FAIADATSLTEAGYVGEDVENILLKLIQAADYNVERAERGITYVDEIDKIA  
 KKGGENVSITRDVSGEGVQQALLKIIEGTVASVPPQGGRKHPNQEMIQINT  
 KNILFIVGAFDGIEDLVKQRLGEKVGFGQTSRKIDDNASYMQEIISEDI  
 QKFGLIPEFIGRLPVVALELLTAEDLVRILTEPRNALVKQYQTLLSYDG  
 VELEFDQDALLAIADKAIERKTGARGLRSIEETMLDIMFEIPSQEDVTKV  
 RITKAAVEGTDKPVLETA\*

ID-71

Clone 3-20

ATGAAAAGATTACATAACTGTTATAACCGTAATTGCTACATTAGG  
 TATGTTGGGGTAATGACCTTGGTCTTCCAACGCAGCCGCAAAACG  
 TAACGCCGATAGTACATGCTGATGTCAATTCTGTTGATACGAGC  
 CAGGAATTCAAAATAATTAAAAATGCTATTGGTAAACCTACCATT  
 TCAATATGTTAATGGTATTATGAATTAAATAATCAGACAAATT  
AAATGCTGATGTCAATGTTAAAGCGTATGTTCAAAATACAATTGACA

FIG. 1 CONT'D

ATCAACAAAGACTATCAACTGCTAATGCAATGCTTGTAGAACCAATT  
CGTCAATATCAAAATCGCAGAGATACCACTCTTCCCGATGCAAATTG  
GAAACCATTAGGTTGGCATCAAGTAGCTACTAATGACCATTATGGGC  
ATGCAGTCGACAAGGGGCATTTAATTGCCTATGCTTAGCTGGAAAT  
TTCAAAGGTTGGGATGCTTCCGTGTCAAATCCTCAAAATGTTGTCACA  
CAAACAGCTCATTCCAACCAATCAAATCAAAAAATCAATCGTGGACA  
AAATTATTATGAAAGCTTAGTCGTAAGGCGGTTGACCAAAACAAAC  
GTGTTCGTTACCGTGTAACTCCATTGTACCGTAATGATACTGATTAG  
TTCCATTGCAATGCACCTAGAAGCTAAATCACAAGATGGCACATTA  
GAATTAAATGTTGCTATTCCAAACACACAAGCATCATACACTATGGA  
TTATGCAACAGGAGAAATAACACTAAATTAA

MKRLHKLFITVIATLGMLGVMTGLPTQPQNVTPIVHADVNSSVDTSQEFQNNLKNAIGNLPFQYVNGIYELNNNQTNLNADVNVKAYVQNTIDNQQRLSTANAMLDRTIRQYQNRRDTTLPDANWKPLGWHQVATNDHYGHAVDKGHЛИЯALAGNFKGWDASVSNPQNVVTQTAHSNQSNSQKINRGQNYYESLVRKAVDQNKRVRYRVTPLYRNDTDLVPFAMHLEAKSQDGTLEFNVAIPNTQASYTMDYATGEITLN\*

ID-72

### Clone 13

ATGAAAAAACTATCGAAAACCTATTGTACTACTACTCTAACTCTTTTT  
GCCATTTTATGGGAGCATATGCTTACACGCATATTGTTGAAAAAAAG  
ATCCCTAACTAGCAATACTATTGAAAAAAACTCTACCTGTGGTAAATC  
AGATTAAGCCTCAAACCATTAAAGAATACCAAAATTACTTAACTAAG  
GTAGCTAAACGTAATGTTCTCCTGTAGACATTCTCAGGCATTAAAT  
AATGAAAAGGTAGAAATTACTGCTACTGATGGCATGCAAACATTCAC  
TTGGAATGATAAAAATAATCCTAAGCAAAAGGTTATCTTCTATGTT  
ATGGAGGATCATATATCCATCAAGCTTCCGAATTACAATAATTGTTG  
TCAATAAAACTAGCTAAAAATTAGATGCAAAAGTTGTCTTCCTATT  
ACCCTAAAGCTCCTACATATAATTATAGTGTGCTATCCCCAAAATT  
AAAAAATTATACCAAAATACATTAGCTAGCGTCACATCTCACAAACAG  
ATTATCCTAGTAGGTGAAAGTGCAGGCGGAGGCCTTGCTTAGGTAT  
TGCTGATAACCTTGCACGGAGCATATCAAACAACCAAAAGAAATTAT  
TTTAA

MKNYRKLVLLLIFFAIFMGAYAYTHIVEKRSLTSNTIEKTLPVVNQIKP  
QTIKEYQNYLTVAKRNVLPVDIPQALNNEKVEITATDGMQFTWNDK  
NNPKQKVIFYVHGGSYIHQASELQYIFVNKLAKKDAKVVFPIYPKAPT  
YNYSDAIPKIKKLYQNTLASVTSHKQIILVGESAGGGLALGIADNLARSIS  
NNQKKLF\*

FIG. 1 CONT'D

ID-73

Clone 2-19

TTGATTCTAATAACTCCTATGGGATAATATCTTATCACAAAAATTG  
 AGGGAATTATTATGAAGTTAAAACATATTGTCTAGGATTAGCCTTA  
 ACAACACTTTAGGAGTCACATTAGTAATCAAGAAGTTCAGCAAG  
 CTCAACTTCAGTAAAGTTAAAGTTGGTGTATGACCTTCTGA  
 CACTGAAAAAGCACGTTGGATAAAATTGAAAAGCTAGTAGGTGAT  
 AAAGCTAAAATCAAATTACAGAATTACAGATTATACACAACCAAA  
 TCAAGCGACAGCCAATAAGGATGTGGATATTATGCCTTCAACATT  
 ACAATTCTTAGAAAAGTGGATAAAGGAAAATAAGAAAAACTTAATT  
 CCACTTGAAAAGACTTACTTAGCTCAATTCTGATCTATTCTGAGAAG  
 GTAAAATCTCTAAAAATTGAAAAAGGCCACTATTGCAATTCC  
 AAATGATGCAACAAATGGTAGCCGTGCATTGTATGTCCTCAGTCAG  
 CAGGTTTAATCAAATTGAATGTTCTGGTAAGAAGGTTGCAACAGTT  
 GCTAATATCACATCTAATAAAAAGGATATTATATTCAAGGAGTTAGA  
 TGCGAGTCAAACACCACGTGCACTCAAAGATGTAGATGCAGCTATT  
 TTAATAATACATACATTGAGCAAGCTAATTAAAACCTTCAGATGCT  
 ATCTTGTGAGAAATCAGATAAAAATTCAAAACAATGGATTATAT  
 CATTGCGGGACGTAAAATTGGAAAAAGCAAAAGAACGCTAAAGCT  
 ATCCAAGCTATCTGGATGCTTATCACACAGATGAAGTAAAAAGT  
 TATCAAAGATACTCAGCTGATATTCCACAATGGTAA

MILITSYGIISLSQKLREFIMKLKHIVLGLALTLGVTSNQEVSASSTSS  
 KVVKVGVMFTSDTEKARWDKIEKLVDKAKIKFTEFTDYTQPNQATAN  
 KDVDINAFQHYNFLENWNKENKKNLIPLEKTYLAPIRYSEKVKSLLKKL  
 KKGATIAIPNDATNGSRALYVLQSAGLIKLNVSGKKVATVANITSNKKDI  
 NIQELDASQTPRALKDVDAAIINNTYIEQANLKPSDAIFVEKSDKNSKQW  
 INIAGRKNWKKQKNAKAIQAILDAYHTDEVKKVIKDTSDIPQW\*

ID-74

Clone 3-6

ATGTCAAATCAATATGATTATATCGTTATTGGTGGAGGTAGTGCAGG  
 CAGTGGTACCGCTAATAGGGCAGCCATGTATGGAGCAAAAGTCCTGT  
 TAATTGAAGGTGGACAAGTAGGTGGAACCTGTGTTAACTTAGGTGTT  
 GTACCTAAGAAAATCATGTGGTATGGTGCACAAGTTCTGAGACACT  
 CCATAAGTATAGTTAGGTTATGGTTTGAAGCCAATAATCTTAGTT  
 TGATTTACTACTCTAAAAGCTAATCGCGATGCTTACGTGCAGCGGTC  
 TAGACAGTCGTATGCCGCTAATTGAGCGTAATGGGGTCGAAAAGA

**FIG. 1** CONT'D

TTGATGGATTGCTCGTTATTGATAACCATACTATTGAAGTGAATG  
GTCAGCAATATAAGCTCCTCACATTACTATTGCAACAGGTGGACAC  
CCTCTTACCTGATATTATTGGAAGTGAACCTGGTGAGACTTCTGAT  
GATTTTTGGATGGGAGACCTTACCAAATTCTATATTGATTGTTGGG  
GCGGGCTATATCGCGGCAGAACTTGCTGGAGTGGTTAATGAATTAGG  
CGTTGAAACCCATCTGCATTAGAAAAGACCATATTCTACGCGGAT  
TTGATGACATGGTAACAAGTGAGGTTATGGCTGAAATGGAGAAATCA  
GGTATCTCTTACATGCTAACCATGTACCTAAATCTCTAAACCGCGAT  
GAAGGTGGCAAGTTGATTTGAAGCTGAAAATGGGAAAACGCTTGT  
CGTTGATCGTGTAAATATGGGCTATCGGCCGTGGACCAAATGTAGACA  
TGGGACTTGAAAATACCGATATTGTTAAATGATAAAAGATTATATC  
AAAACAGATGAATTGAGAATACTTCTGTAGATGGCGTGTATGCTAT  
TGGAGATGTTAATGGGAAAATTGCCCTGACACCGGTAGCAATTGCAG  
CAGGTCGTCGCTTATCAGAAAGACTTTAATCATAAAAGATAACGAA  
AAATTAGATTACCATATAATGTACCTTCAGTTATTCTACTCACCCTGTA  
ATTGGGACGGTAGGACTTCAGAAGCAGCAGCTATCGAGCAATTGG  
AAAAGATAATATCAAAGTCTATACATCAACTTTACCTCTATGTATAC  
GGCTGTTACCACTGAAATCGCCAAGCAGTTAAGATGAAGCTCATAC  
TAGGAAAAGAGGAAAAAGTTATTGGGCTTCAGTTGCTATCAAACCC  
ATTGATGAAATGATTCAAGGTTTCAGTTGCTATCAAATGGGGGC  
TACTAAAGCAGACTTGTATGATACTGTTGCTATTCAACCCAACTGGATC  
TGAGGAATTGTTACAATGCGCTAA

MSNQYDYIVIGGGSAGSGTANRAAMYGAKVLLIEGGQVGGTCVNLGC  
VPKKIMWYGAQVSETLHKYSSGYGFEANNLSFDFTTLKANRDAYVQRS  
RQSYAANFERNGVEKIDGFARFIDNHTIEVNGQQYKAPHITIATGGHPLY  
PDIIGSELGETSDDFFGWETLPNSILIVGAGYIAAELAGVVNELGVETHLA  
FRKDHLRGFDDMVTSEVMAEMEKSGISLHANHVPKSLKRDEGGKLIFE  
AENGKTLVVDRVIWAIGRGPNVDMGLENTDVLNDKDYIKTDEFENTSV  
DGVYAIGDVNGKIALTPVAlAAGRRLSERLFNHKDNEKLDYHNVPSVIF  
THPVIGTVGLSEAAAIEQFGKDNIKVYTSTFTSMYTAVTSNRQAVKMKLI  
TLGKEEKVIGLHGVGYYGIDEMIQGFSVAIKMGATKADFDDTVAIHPTGS  
EEFVTMR\*

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### Clone 3-51

ATGAGTATCAAAAAAGTGTGATTGGTTTGCCCTCGAAGCTGCAGC  
ATTATCAATGTTGCTTGTGTAGACAGTAGTCATCTGTTATGGCTGC

FIG. 1 CONT'D

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CGAGAAGGATAAAGTCGAAATTACGTGGTGGCTTTCCAACCTTA  
CTCAAGAAAAGGCTAAGGATGGAGTAGGTACTTATGAGAAAAAAAGT  
CATCAAGGCTTTGAAAAGAAAAATCCTAATATAAAAAGTAAAAGT  
AGACAATTGATTACATCTGGACCTGAAAAAAACTACAGCAATT  
GAAGCAGGGACAGCACCTGATGTGCTTTGATGCACCAGGGCGAAT  
TATTCAATATGGTAAAAATGGTAAATTAGCAGATTGAATGATTATT  
TACAGACCAATTATTAGGATGTCAATAATAAGAACATCATTCAAG  
CTTCTAAGTCTGGCGATAAACGCTACATGTATCCAATAAGTTCTGCC  
CATTTATATGGCGTCAATAAAAAAAATGCTAAAGATGCAGGAGTT  
TTGAAACTTGTAAAAGAAGGTTGGACTACTAGTGTATTGAAAAAAAGT  
ACTAAAAGCACTAAAAAATAAGGCTATACACCAGGTTCAATTCTTG  
CAAACGGGCAAGGAGGAGATCAAGGACCACGTGCATTGCTAAT  
CTTATAGTGTCCAATAACAGATAAAGAAGTAACAAAATATACCAC  
TGACACTAAAAATTCTGTAAAATCAATGAAAAAAATAGTTGAATGGA  
TTAAGAAAGGCTACTTGTATGAATGGGTCTCAGTATGATGGCTCAGCT  
GACATTCAAAACTTCGCCAATGGACAAACTGCTTCACTATCCTATG  
GGCTCCAGCTCAACCAAAACTCAAGCAAAATTATTAGAGTCAGTA  
AAGTGGATTACCTTGAAGTGCCATTCCATCAGAAGATGGAAAACCA  
GATTAGAATACCTTGTAAATGGTTGCGGTCTTAATAATAAAAGAT  
AAAAACAAAGTAAAAGCCTCAAGAAATTATCCTTATTGCTGA  
TGATAAAAATGGGACCAAAAGATGTTACGTACAGGTGCTTCC  
CAGTTAGAACATCATTGGGGATCTTATAAAAGGTGATAAACGTATG  
ATGAAGATTCAAAATGGACTCAATATTATTACCCATTACAACAC  
TATCGATGGATTCTGAAATGAGAACCTTATGGTCCCAATGGTCA  
ATCTGTATCCAATGGTGTGAAAAACCAAGCAGATGCTTGAAGACT  
TTACTCAAAAGCAAATGATACCATTAAAAAAGCAGCTAAATAA

MSIKKSVIGFCLEAAALSMFACVDSSQSVMAAEKDKVEITWWAFPTFTQ  
EKAKDGVGTYEKVKAFKKNPNIKVKLETIDFTSGPEKITTAEAGTAP  
DVLFDAPGRIIQYKGNGKLADLNDLFTDQFIKDVNKNIIQASKSGDKA  
YMPYPISSAPFYMAFNKKMLKADVLKLVKEGWTTSDFEKVLKALKNK  
GYTPGSFFANGQGGDQGPRAFFANLYSAPITDKEVTKYTTDTKNSVKSM  
KKIVEWIKKGYLMNGSQYDGSADIQNFAANGQTAFTILWAPAQPKTQAK  
LLESSKVDYLEVPFPSEDGKPDLLEYLVNGFAVFNNKDENCVKASKKFIT  
FIADDKKWGPKDVRTGAFPVRTSGFDLYKGDKRMMKISKWTQYYSPY  
YNTIDGFSEMRTLWFPMVQSVSNGDEKPADALKDFTQKANDTIKKAAK  
\*

ID-76 (Same as ID-39)

Clone 3-56

ATGAGGAAACGTTTCTTGCTAAATTATTGTTACTTTATT  
TCTTTTCTTATTCTTTCCGCTTTAAGGCCAAGATTGTCAGGT

FIG. 1 CONT'D

---

TGTTTATGCAAGTTCAAGGAGATCATTGGGACATTGTAACGCATT  
TGATTTCCGTATTCACATCGCTTGATCTCATTAAGGTAAAGAAAA  
TCAACTTACTTATAGGTTGATACAATTGCTAACAGTAAAGCCTACAC  
TGAGGATTGGAGTGATAAAAGGCCGAATTTGTTGCTCGTTAATAC  
TCAAAACCACATACATTGGAAGGATTGCAACAATTGCCTCAAACATTAT  
TAAAAAAATCATGGATACTATGCCATTAGGATGAAGGATATTCAATTG  
ATTACTTCAGTACAAGGGGTACTCAAACACTCACTTATCCAGAATTTCT  
ACTACAGGCGACTGGCAATTAGAACGGCTTTCGATGAGGAGACAAG  
CGATGTGGTGAAGTGGATATTAATCAGGATGGTAAGGATGAGTATG  
TGATCATCCAAGGTTTACGGAGATCGTTACGTATCTCACTGAAG  
ATTCGGTCGAGAATTATTCCATTATCCTGAAAAAACCCATTGGTC  
ACGCTATTGGAGTGGCGTTACTTAATCAGACTGTTCGTATTG  
GGTGGCGATCAGAAAAAGCAGAATTAGGCTTTCACTTGTAGAT  
GGCACTTGGTTCAAGAATTAGTAGATGCAAAAGCAGCTCTAGTAA  
TGTCTTAGCTTTGAAAAAGATGGAAAAGCTTATCTTCTCAGCCAA  
TAACGGACGTGGCGAAGTGCTTTATCAATTAGTAAAATAA

MRKRFSLNFIVVTFIFFFFILFPLFKAKDCQVYASFQGDHWDICNAFDF  
PYLHRFDLIGKENQLYFIGCTIANSKAYTEDWSDKGRIFVARFNTQNHT  
LEGLQQLPQTLLKNHGYYAIQDEGYSLITSVEGVLKLTYPFSTTGDWQ  
LERLFDEETSDVVVKVDINQDGKDEYVIIQGFHGDRRLIFTEDFGRELFHY  
PEKTPFGHAIWSGRLLNQTCFVFGWRSEKAELRLHFVDGHLVSELVDA  
KAASSNVLAFAEKDGKAYLFSANNRGEVALYQLVK\*

### FIG. 1 CONT'D

nucS1

Bgl II Eco RV  
5'-cgagatctgatatctcacaaacagataacggcgtaaatag -3'

nucS2

Bgl II Sma I  
5'-gaagatcttccccggatcacaaacagataacggcgtaaatag -3'

nucS3

Bgl II Eco RV  
5'-cgagatctgatccatcacaaacagataacggcgtaaatag -3'

nucR

Bam HI  
5'-cggatccttatggacctgaatcagcgttgtc -3'

NucSeq

5'-ggatgcttgcgttca -3'

pTREP<sub>F</sub>  
5'-catgatatcggtacctcaagctcatatcattgtccggcaatggtggtggctttttgttttagcggataa  
caatttcacac -3'

pTREP<sub>R</sub>

5'-gcggatccccggcttaattaatgtttaaacactagtcgaagatctcgcaatttcctgtgtgaaatt  
gttatccgcta -3'

pUC<sub>F</sub>

5'-cgccagggtttccagtcacgac -3'

v<sub>R</sub>

5'-tcagggggggcggagcctatg -3'

v<sub>1</sub>

5'-tcgtatgtgtgtggattgtg -3'

v<sub>2</sub>

5'-tccggctcgatgtgtgtggattg -3'

FIG. 2

pTREP-Nuc vectors allow cloning of genomic DNA into each frame with respect to the nuclease gene

(i)

Cloning site is indicated by an arrow.

The diagram illustrates the pTREP-nuc Cassette (iii) with the following components and features:

- Restriction sites:** Eco RI, Bgl II, Sma I or Eco RV, Kpn I, and Pst I are indicated along the cassette.
- Promoters:** The P1 promoter is located upstream of the nuc gene.
- Terminators:** Transcription terminators are positioned at both ends of the cassette.
- Sequencing primer:** A sequencing primer is located downstream of the P1 promoter.
- Gene:** The nuc gene is represented by a long horizontal bar.
- Transcription direction:** Arrows indicate transcription starting from the P1 promoter and ending at the terminators.

FIG. 3

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## GBS Vaccination - Trial 1

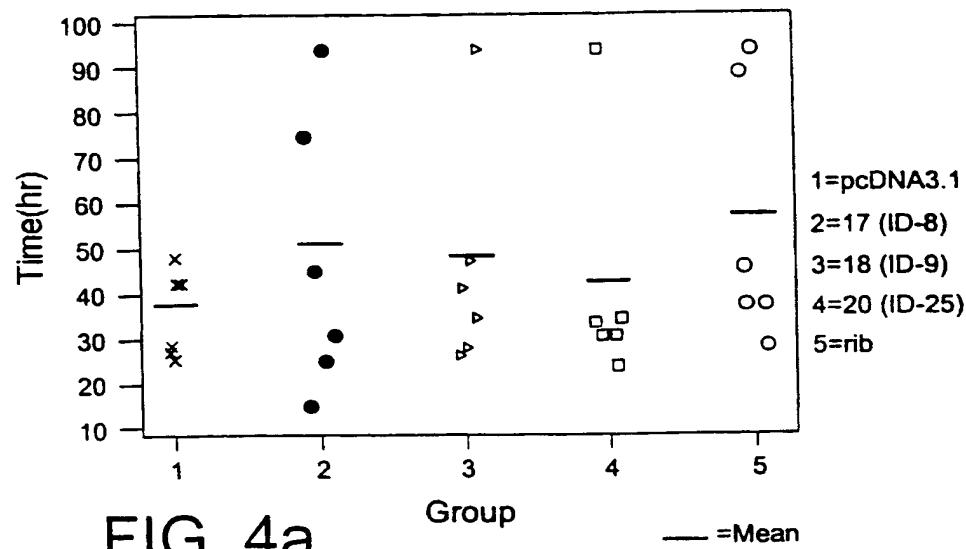


FIG. 4a

## GBS Vaccination - Trial 2

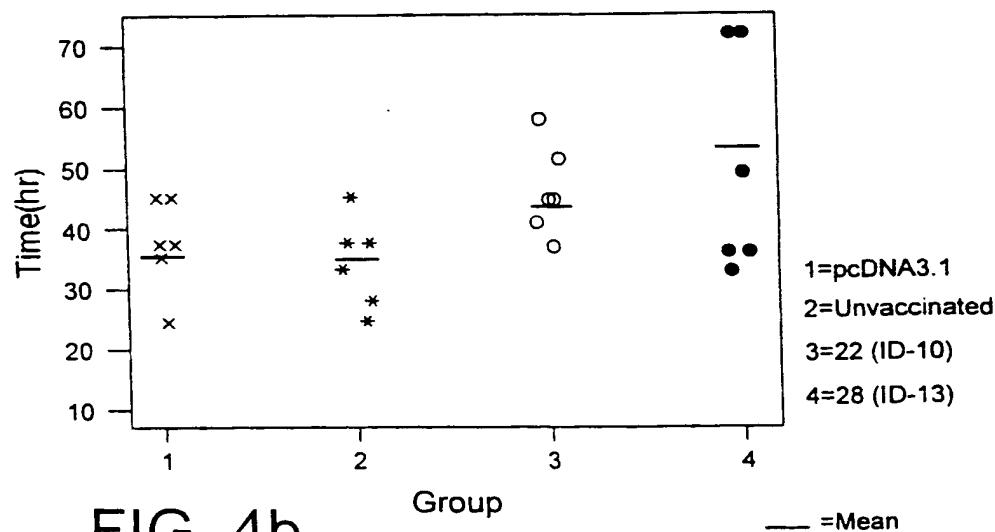


FIG. 4b

GBS Vaccination - Trial 3

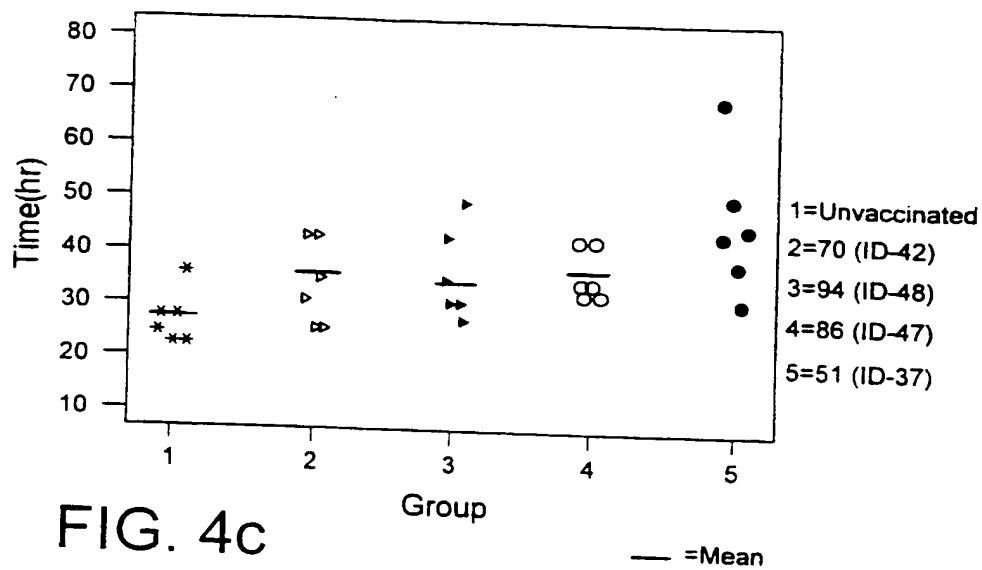


FIG. 4c

GBS Vaccination - Trial 4

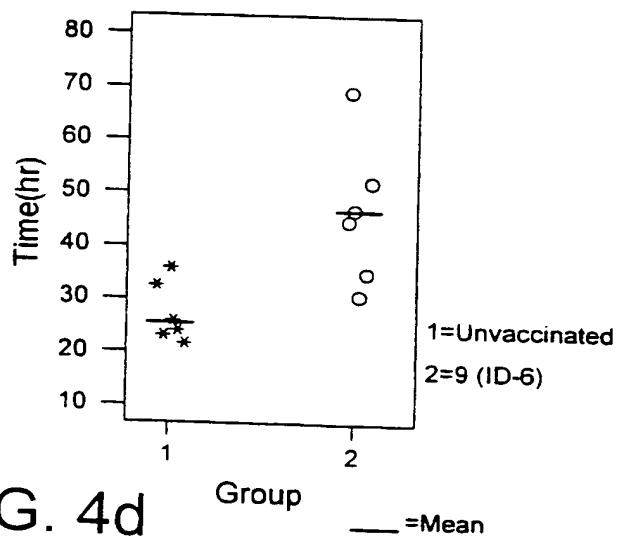


FIG. 4d

## GBS Vaccination - Trial 6

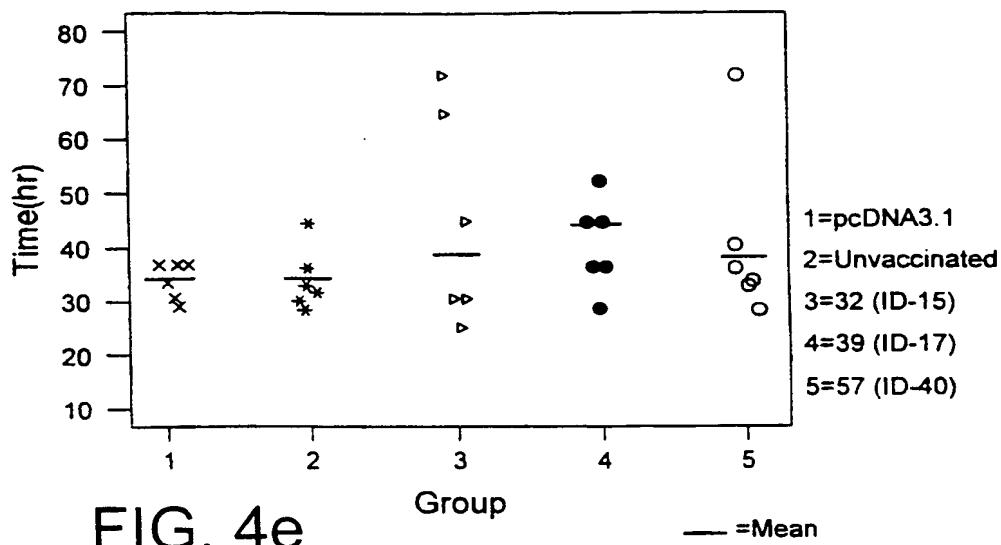


FIG. 4e

## GBS Vaccination - Trial 2

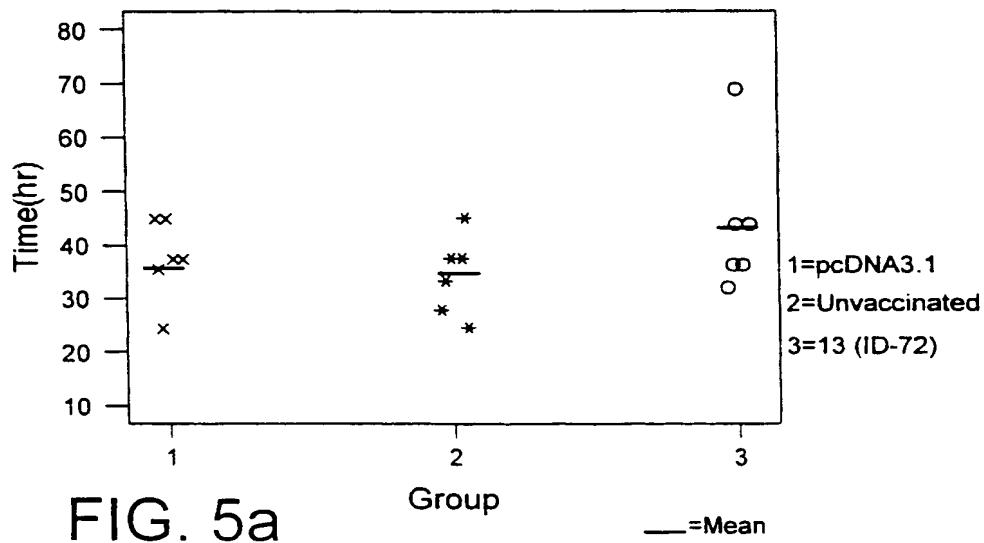


FIG. 5a

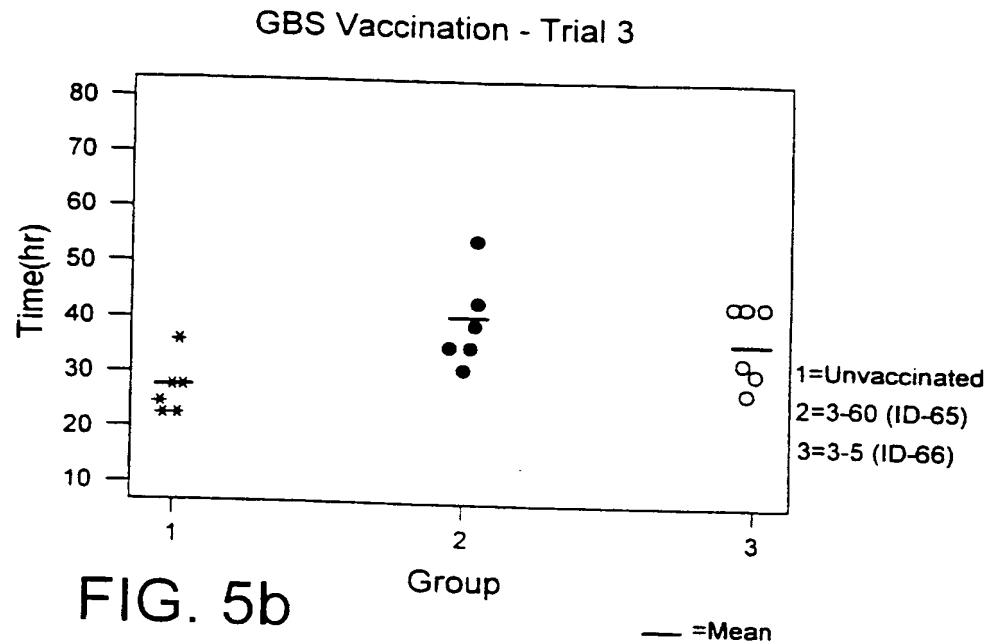


FIG. 5b

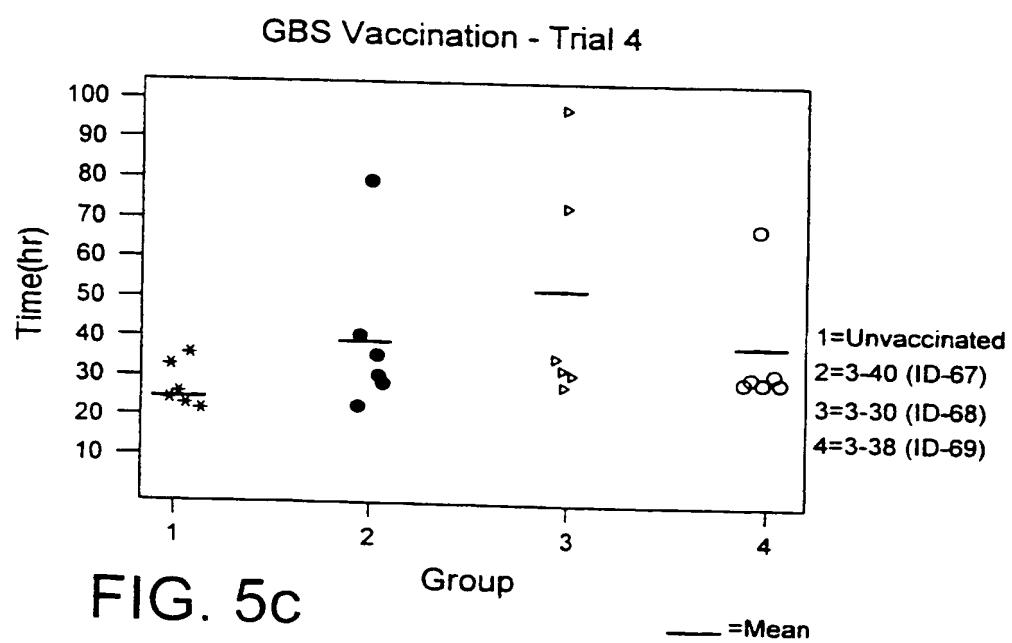


FIG. 5c

## GBS Vaccination - Trial 5

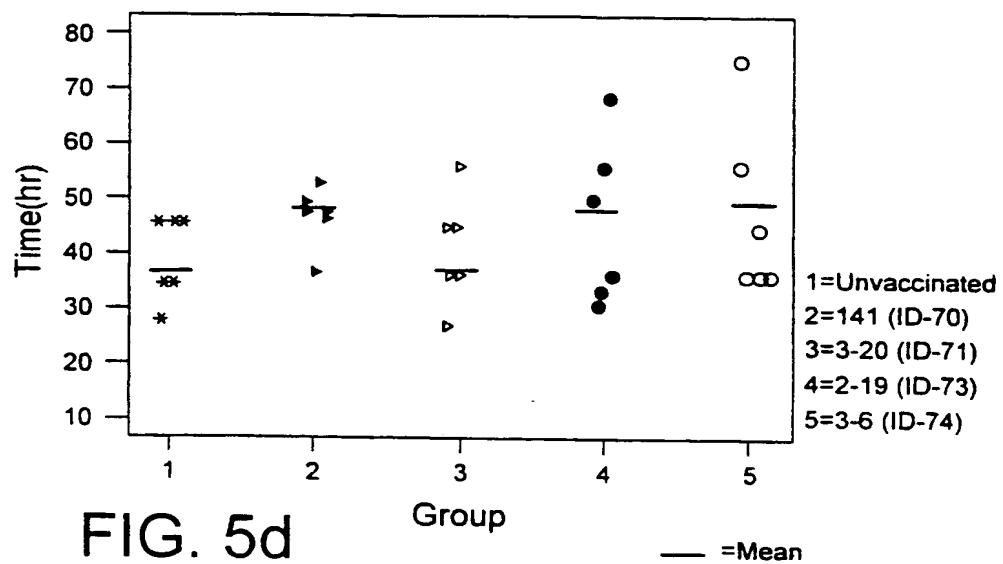


FIG. 5d

## GBS Vaccination - Trial 6

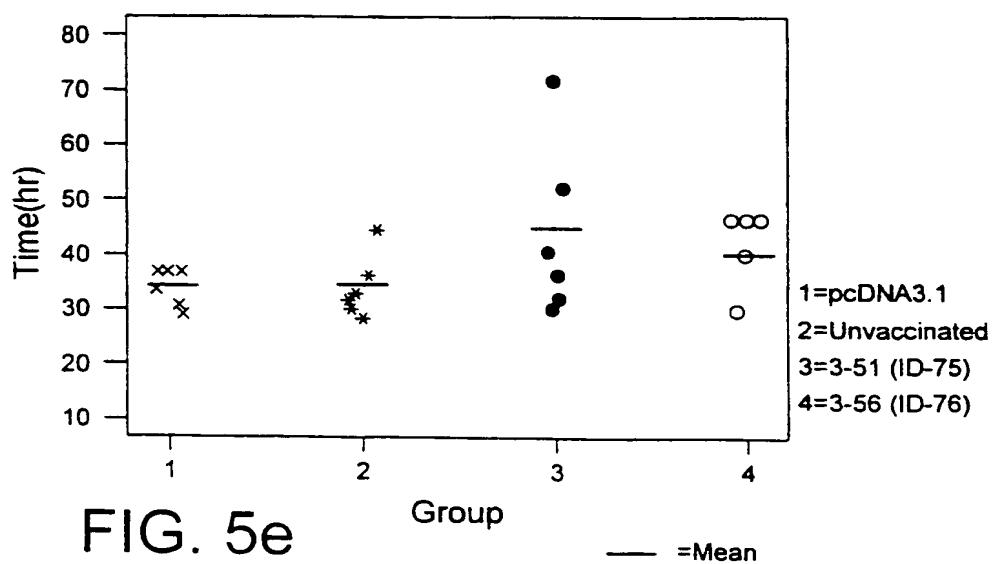


FIG. 5e

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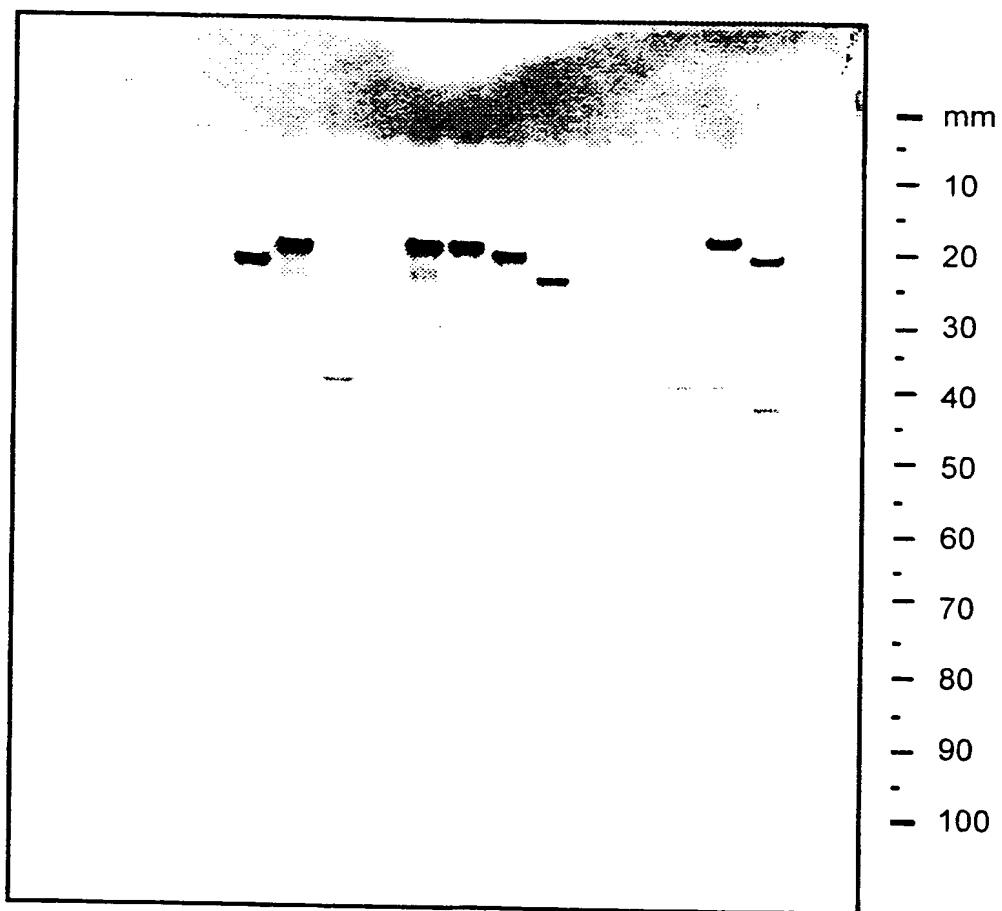


FIG. 6

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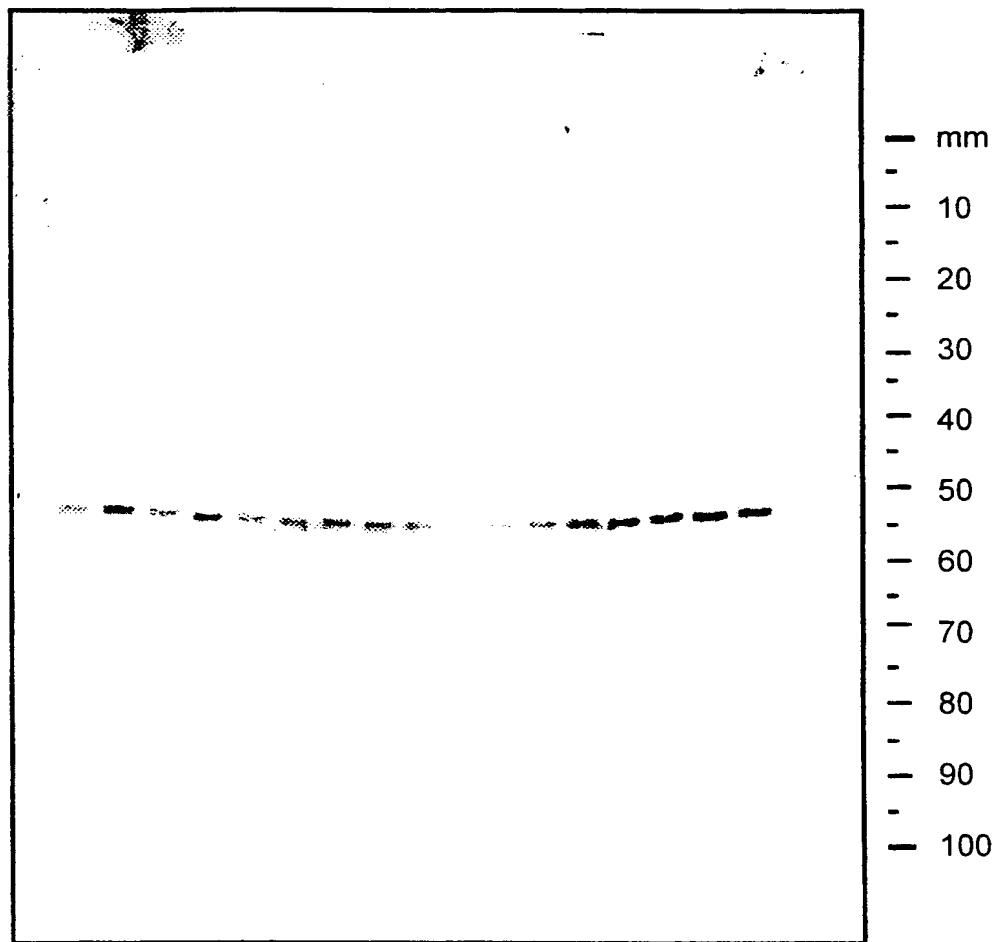


FIG. 7

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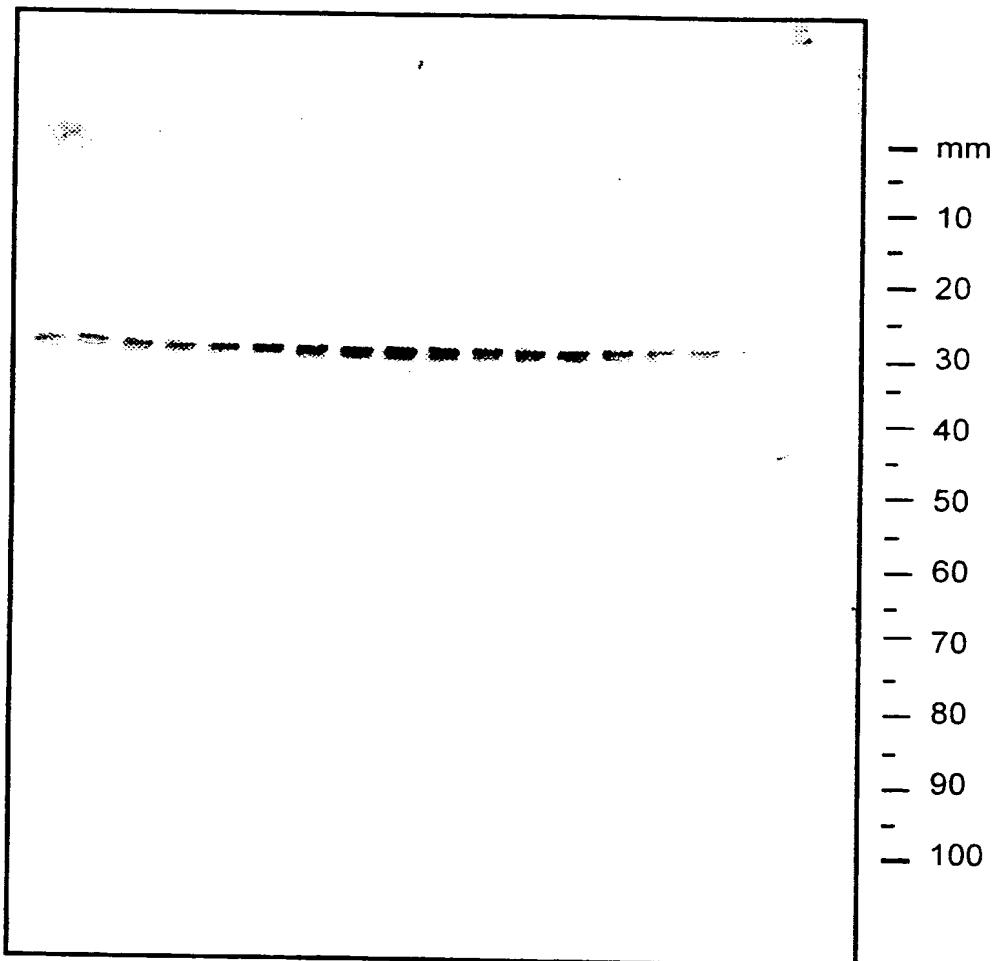


FIG. 8

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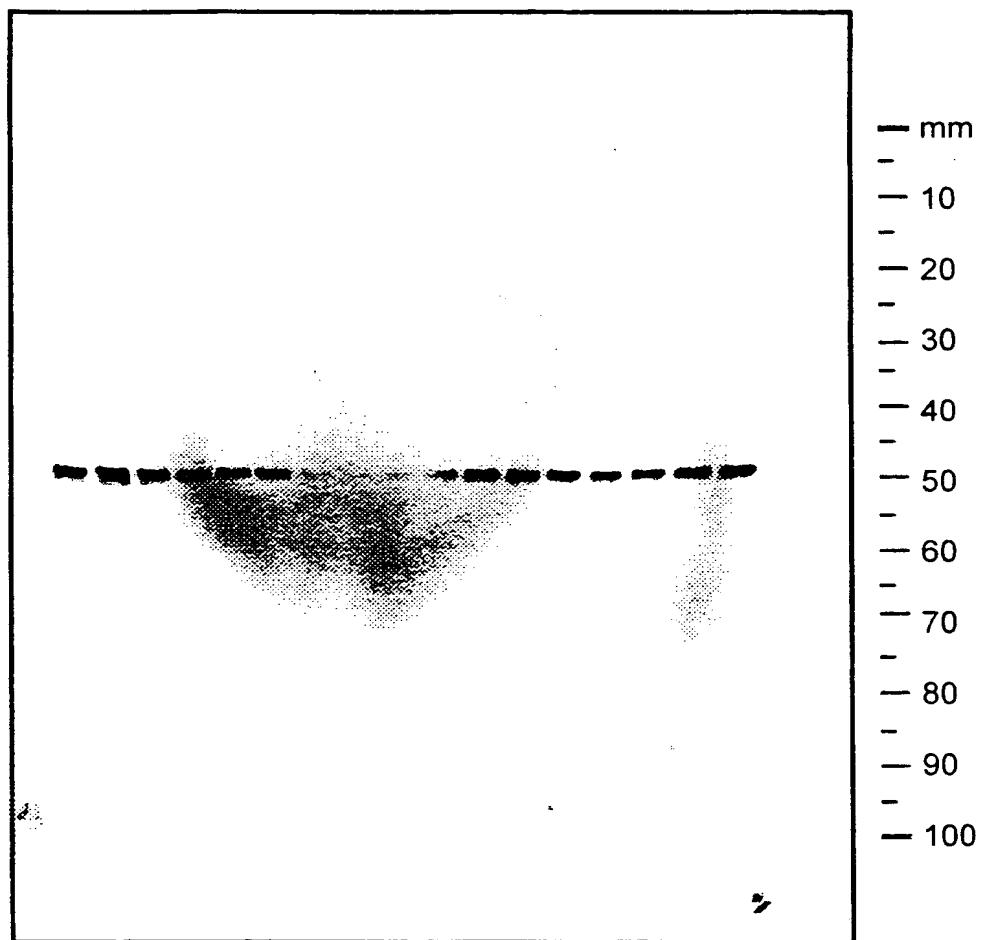


FIG. 9

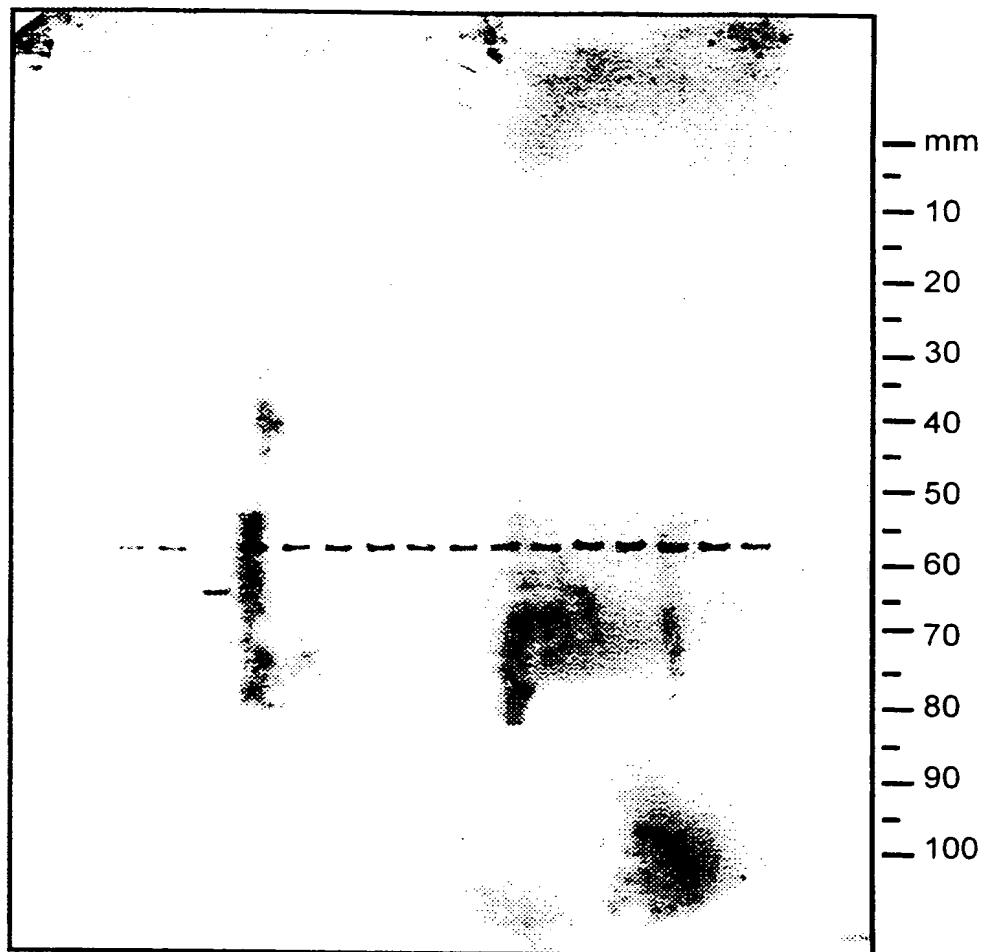
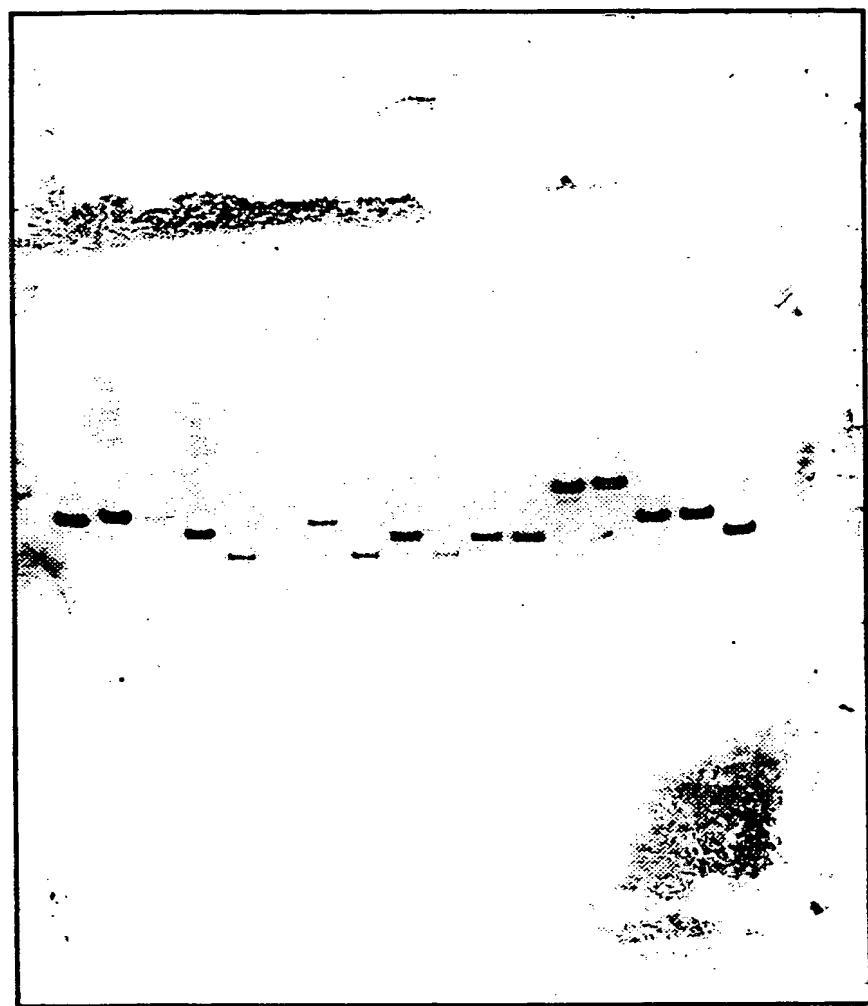
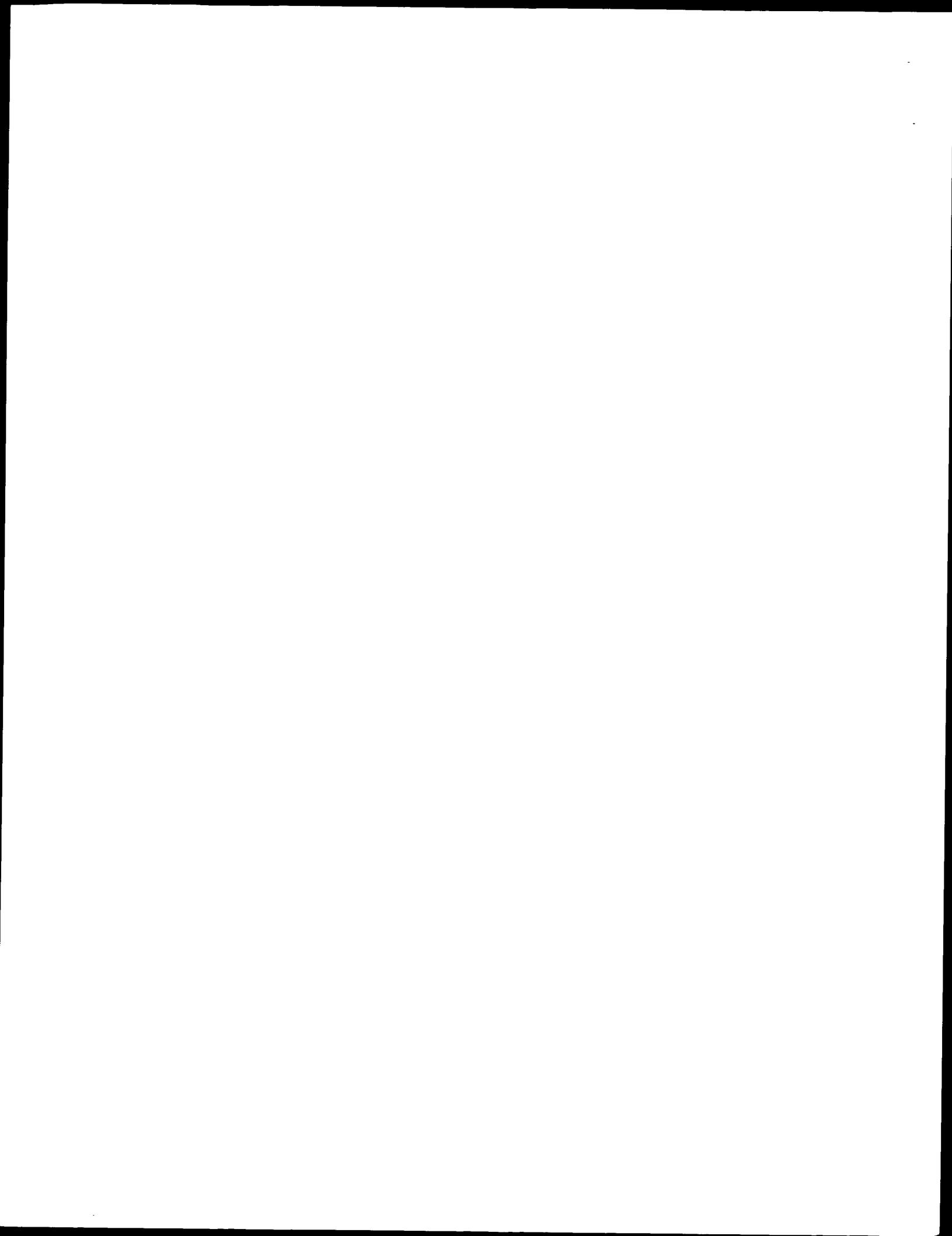


FIG. 10

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**FIG. 11**





## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification <sup>7</sup> :	A3	(11) International Publication Number: <b>WO 00/06736</b>
C12N 15/31, 15/74, 15/62, 15/10, 9/16, 1/19, 1/21, C07K 14/315, 16/12, A61K 31/70, 39/09, G01N 33/53, 33/68, C12Q 1/68		(43) International Publication Date: 10 February 2000 (10.02.00)

(21) International Application Number: PCT/GB99/02444	(81) Designated States: CA, CN, JP, US, European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).	
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(71) Applicant (for all designated States except US): MICROBIAL TECHNICS LIMITED [GB/GB]; 20 Trumpington Street, Cambridge CB2 1QA (GB).		
(72) Inventors; and		
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(74) Agents: CHAPMAN, Paul, William et al.; Kilburn & Strode, 20 Red Lion Street, London WC1R 4PJ (GB).		

(54) Title: NUCLEIC ACIDS AND PROTEINS FROM GROUP B STREPTOCOCCUS

## (57) Abstract

Novel protein antigens from Group B *Streptococcus* are described, together with nucleic acid sequences encoding them. Their use in vaccines and screening methods is also described.

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# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/GB 99/02444

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC 7	C12N15/31	C12N15/74	C12N15/62	C12N15/10	C12N9/16
	C12N1/19	C12N1/21	C07K14/315	C07K16/12	A61K31/70
	A61K39/09	G01N33/53	G01N33/68	C12Q1/68	

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**B. FIELDS SEARCHED**

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IPC 7 C12N C07K A61K G01N C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE TREMBL E.M.B.L. Databases Accession Number: Q54914, 1 November 1996 (1996-11-01) PODBIELSKI A ET AL: "ORF 1 AND ORF2 5' REGION" XP002133342 97.2% identity in 141 aa overlap with SeqIdNo.12 abstract --- -/- -	3,4

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Patent family members are listed in annex.

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- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

\*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

\*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

\*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

\*Z\* document member of the same patent family

Date of the actual completion of the international search

Date of mailing of the international search report

17 March 2000

11.04.00

Name and mailing address of the ISA

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Lonnoy, O

## INTERNATIONAL SEARCH REPORT

National Application No  
PCT/GB 99/02444

## (continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>WO 98 18930 A (HUMAN GENOME SCIENCES INC ;CHOI GIL H (US); HROMOCKYJ ALEX (US); J) 7 May 1998 (1998-05-07) SP0020: 51.9% identity in 262 aa overlap with SeqIdNo.133 &amp; DATABASE GENESEQ E.M.B.L. Databases Accession Number: W55078, 2 October 1998 (1998-10-02) CHOI G ET AL: "Streptococcus pneumoniae SP0020 protein" XP002133369 51.9% identity in 262 aa overlap with SeqIdNo.133 abstract</p> <p>---</p>	3-18,23
P,X	<p>WO 99 16882 A (MEDIMMUNE INC) 8 April 1999 (1999-04-08) &amp; DATABASE GENESEQ E.M.B.L. Databases Accession Number: Y05766, 8 April 1999 (1999-04-08) LUTTICKEN R ET AL : "Streptococcal adhesion mediator protein Lmb" XP002133343 99.7% identity in 306 aa overlap with SeqIdNo.12 abstract</p> <p>---</p>	1-18,23
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A	<p>LACHENAUER C S ET AL: "Cloning and expression in Escherichia coli of a protective surface protein from type V group B Streptococci" ADVANCES IN EXPERIMENTAL MEDICINE AND BIOLOGY, US, SPRING ST., NY, vol. 418, no. 418, 9 December 1997 (1997-12-09), page 615-618-618 XP002107261 ISSN: 0065-2598 the whole document</p> <p>---</p>	1-18,23
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National Application No  
PCT/GB 99/02444

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Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	LARSSON C ET AL: "Experimental vaccination against group B streptococcus, an encapsulated bacterium, with highly purified preparations of cell surface proteins Rib and alpha" INFECT. IMMUN., vol. 64, no. 9, September 1996 (1996-09), pages 3518-3523, XP002125783 cited in the application	
A	WO 95 06732 A (MASURE H ROBERT ; TUOMANEN ELAINE (US); PEARCE BARBARA J (US); UNIV) 9 March 1995 (1995-03-09) ---	
A	DATABASE SWISSPROT E.M.B.L. Databases Accession Number: P42422, 1 November 1995 (1995-11-01) YOSHIDA K ET AL: "Hypothetical sensor-like Histidine Kinase in IDH 3' region" XP002133344 30.6% identity in 320 aa overlap with SeqIdNo.20 abstract ---	
A	DATABASE SWISSPROT E.M.B.L. Databases Accession Number: P39845, 1 February 1995 (1995-02-01) TOGNONI A ET AL: "Peptide Synthetase 1" XP002133345 29.3% identity in 133 aa overlap with SeqIdNo.26 abstract --- -/-	

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PCT/GB 99/02444

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# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/GB 99/02444

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
  
3.  Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1.  As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
  
2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
  
3.  As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:  
1-18 and 23 (all partially) as relating to inventions 1, 6, 10, 13, 35, 41, 62, 63 and 67
  
4.  No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

### Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

URTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Invention 1: claims 1-18 and 23 (all partially)

A Group B Streptococcus protein having a sequence as depicted in SeqIdNo.2, a fragment, derivative or variant of said protein; a nucleic acid molecule comprising or consisting of SeqIdNo.1, a nucleic acid molecule complementary to said sequence, a nucleic acid molecule encoding for the a derivative or fragment of said protein; a vector comprising said nucleic acid molecule and afferent recombinant DNA practices; an antibody to said protein; an immunogenic composition comprising said protein or said nucleic acid and applications thereof; a method or kit of detection of Group B Streptococcus comprising said protein, said antibody, or said nucleic acid molecule; a method of determining whether said protein represents a potential antimicrobial target which comprises inactivating said protein and determining whether Group B Streptococcus is still viable.

2. Inventions 2-69: claims 1-18 and 23 (all partially)

Idem as subject 1 but limited to each of the polynucleotide and polypeptide sequences as depicted in SeqIdNo:3-137, wherein invention 2 is limited to SeqIdNo:3 and SeqIdNo:4, invention 3 is limited to SeqIdNo:5 and SeqIdNo:6, ..., invention 58 is limited to SeqIdNo:115, ..., and invention 69 is limited to SeqIdNo:136 and 137.

3. Inventions 70: claims 19-22 (all totally)

A method for screening for DNA encoding bacterial cell enveloppe associated or surface antigens in gram positive bacteria comprising a reporter vector including the nucleotide sequence encoding the mature form of the staphylococcus nuclease gene and an upstream promoter region with DNA from a gram positive bacterium; said method wherein the reporter vector is one of the pTREP1-nuc vectors; said method wherein the gram positive bacterium is Group B Streptococcus, Streptococcus pneumoniae, Staphylococcus aureus or pathogenic group A streptococci; said vector which is one of the pTREP1-nuc vectors

For the sake of conciseness, the first and 70th subject-matters are explicitly defined, the other subject-matters are defined by analogy to the subject-matter of invention 1.

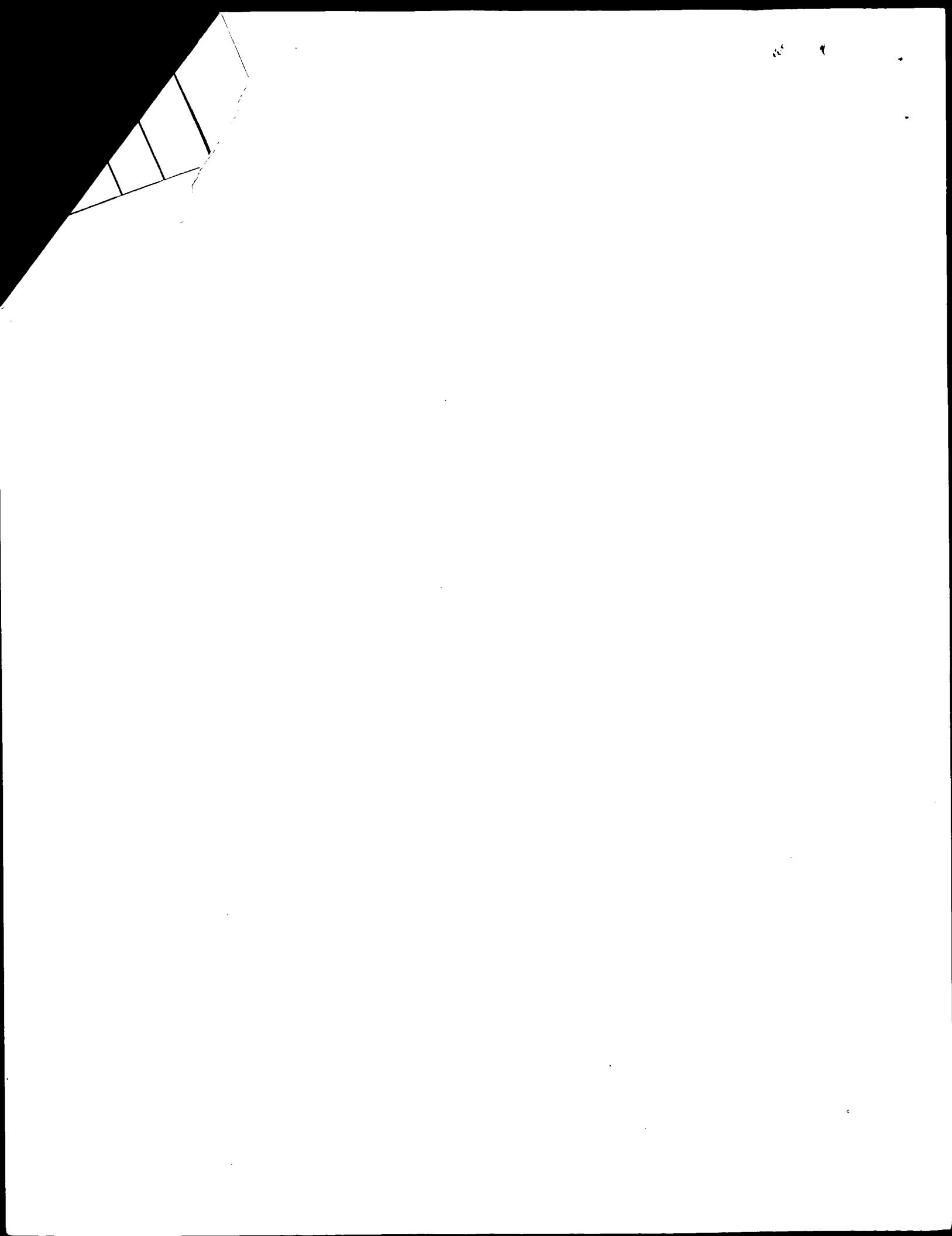
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Information on patent family members

National Application No

PCT/GB 99/02444

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